

HemaCase Clinical Case Booklet

By using Mindray Fully Automated Cellular Analysis Lines

Volume 2







HemaCases—Clinical Case Booklet | 01

We are excited to present our clinical case booklet on hematology. At Mindray, we are committed to making cutting-edge technology accessible to all, to ensure high-quality healthcare. As part of this commitment, we strive to enhance the accuracy and efficiency of hematology tests in clinical laboratories.

In this booklet, we provide real clinical cases that vividly illustrate the seamless integration of information from blood analyzers, morphological analysis, and clinical diagnostic data. With the assistance of our state-of-the-art Automated Digital Cell Morphology Analyzer MC-80, these cases showcase how early disease identification and diagnosis can be achieved.

Our goal is to empower laboratory personnel, like yourself, with invaluable insights in your daily work. By immersing yourself in these scenarios, we hope you will expand your knowledge and elevate the quality of patient care you provide.

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List of Abbreviations

WBC \rightarrow White blood cell	Eos \rightarrow Eosinophil	RET# \rightarrow Reticulocyte count
Neu \rightarrow Neutrophil	Eos# \rightarrow Eosinophil count	RET% \rightarrow Reticulocyte percentage
Neu# \rightarrow Neutrophil count	Eos $\% \rightarrow$ Eosinophil percentage	$RHE \rightarrow Reticulocyte \ hemoglobin \ expression$
Neu% \rightarrow Neutrophil percentage	Bas \rightarrow Basophil	$IRF \longrightarrow Immature\ reticulocyte\ fraction$
Lym \rightarrow Lymphocyte	Bas# \rightarrow Basophil count	$PLT \rightarrow Platelet$
Lym# \rightarrow Lymphocyte count	Bas% \rightarrow Basophil percentage	PLT-I \rightarrow Platelet counting with impedance method
Lym% \rightarrow Lymphocyte percentage	$RBC \rightarrow \; Red \; blood \; cell$	PLT-O \rightarrow Platelet counting with optical method
Mon \rightarrow Monocyte	${\sf HGB} \rightarrow {\sf Hemoglobin}$	$IPF \longrightarrow Immature platelet fraction$
Mon# \rightarrow Monocyte count	$MCV \rightarrow Mean corpuscular volume$	
Mon% \rightarrow Monocyte percentage	NRBC \rightarrow Nucleated RBCs	



Case 01 Acute promyelocytic leukemia (APL)

Clinical information

A 22-year-old female patient had fever for 1 month that worsened over the past week, accompanied by gingival bleeding. The patient was diagnosed as "thrombocytopenic purpura" by the local clinic. After treatment, gingival bleeding reoccurred and petechiae/ecchymosis in the lower limbs was observed.

G CBC results

Parameter	Flags		Result	Unit
FR-CRP		н	40.50	mg/L
WBC	&	н	15.93	10^9/L
Neu#	& R		2.99	10^9/L
Lym#			****	10^9/L
Mon#			****	10^9/L
Eos#		L	0.01	10^9/L
Bas#			0.00	10^9/L
IMG#	R		0.00	10^9/L
Neu%	& R	L	18.7	%
Lym%			****	%
Mon%			****	%
Eos%		L	0.0	%
Bas%			0.0	%
IMG%	R		0.0	%
RBC		L	2.49	10^12/L
HGB		L	75	g/L
НСТ		L	22.4	%
MCV			90.1	fL
MCH			30.2	pg
MCHC			335	g/L
RDW-CV			15.8	%
RDW-SD			49.7	fL
PLT		L	28	10^9/L
MPV			9.8	fL
PDW			17.0	
РСТ		L	0.027	%
P-LCC		L	7	10^9/L
P-LCR			26.0	%
NRBC#			0.029	10^9/L
NRBC%			0.18	/100WBC

CBC results on Apr. 18



• WBC increased, Lym and Mon results were of poor reliability and were blocked; RBC and HGB both decreased, indicating moderate anemia; PLT count decreased.

• In the DIFF scattergram, the Lym and Mon group had unclear boundaries, the particles extended towards the upper right in general, and the Mon group was barely recognized as a spindle shape. A flag was raised for abnormal cells.

		Cells pr	e-classification l
		WBC	
V	VBC	419	100%
L L	ymphocyte	52	12.4
L N	Monocyte	12	2.9
В	Basophil	1	0.2
Ν	Ayelocyte	1	0.2
! A	Abnormal promyelocyte	353	84.3
Ν	Non-WBC	257	%
Ν	Nucleated RBC	2	0.5
Ģ	Giant PLT	1	
L	arge PLT	9	
Р	PLT aggregation	1	
S	imudge cell	201	48.0
S	Sediment	43	
		PLT	
Ρ	PLT estimate	Estimated result	Estimation method
Р	PLT concentration	43*10^9/L	Manual
		RBC	
	bize	Degree	%
	Jneven erythrocyte sizes	2+	
	Macrocyte	0	0.5
	Aicrocyte	2+	19.6
	Color	Degree	%
	Hypochromic RBC	0	1.4
	Polychromatic RBC	0	0.3
	ihape	Degree	%
	Poikilocytosis	3+	
	Schistocyte	0	0.9
	chinocyte	2+	7.2
	lliptocyte	0	0.2
! C	Dvalocyte	3+	13.0
S	otomatocyte	0	0.0
L	leptocyte	0	0.0
C	Dacryocyte	0	0.6
C	Contents	Degree	%
-	Basophilic stippling	0	0.0

C Peripheral blood morphology examination

Manual microscopic result

The microscopic examination showed no presence of neutrophils, but presence of lymphocytes (12.4%), monocytes (2.9%), and abnormal promyelocytes (84.3%).

The abnormal promyelocytes varied in cell body shape, with nuclei deformity, curved and folded. The particles were fine, the cell bodies showed irregular protrusions, the nucleoli were clearly recognizable, and no Auer body was definitely observed. Karyolysis was widely observed.

Other examinations

ltem	Result	Reference range
LDH	323	120-250U/L
ESR	40	0-20mm/H
Bone marrow morphology	The microscopic bone marrow images showed acute myeloid leukemia M3, with presence of promyelocytes (86%)	
Immunophenotyping	Positive expression: HLA-DR, CD9, CD13, CD33, CD34, CD38, CD56, CD64, CD117, CD123, MPO	
FISH	PML-RARA fusion gene positive	
Fusion gene quantitative measurement	WT-1 positive	

This patient was diagnosed as APL (Acute promyelocytic leukemia) accompanied by PML-RARA (M3). She was treated with retinoic acid combined with arsenic trioxide for induction therapy; platelets and red blood cells were infused; retinoic acid was used to treat the primary disease; anti-infection therapy, liver-protection therapy, and platelet elevation therapy were ongoing. After two weeks of treatment, the patient's conditions were improved.

Parameter	Flag	JS	Result	Delta#	04-30	04-29	Unit	RBC	PLT
WBC	&	н	12.27	-1.880	14.15	16.13	10^9/L	ind c	
Neu#			****		****	****	10^9/L		
Lym#	& R		1.11	-0.190	1.30	2.10	10^9/L	11 4	1677
Mon#	R	Н	4.40	-0.810	5.21	6.72	10^9/L		
Eos#	R	L	0.00	0.000	0.00	0.00	10^9/L		
Bas#	R		0.01	0.010	0.00	0.00	10^9/L		
IMG#			****		****	****	10^9/L	0 100 200 fL	
Neu%			****		****	****	%		0 10 20 30 fL
Lym%	& R	L	9.0	-0.20	9.2	13.0	%		
Mon%	R	Н	35.9	-0.90	36.8	41.7	%		
Eos%	R	L	0.0	0.00	0.0	0.0	%	WNB	DIFF
Bas%	R		0.1	0.10	0.0	0.0	%		FL
IMG%			****		****	****	%	FS	
RBC		L	2.80	-0.050	2.85	2.94	10^12/L		1
HGB		L	85	-2.0	87	90	g/L		-
HCT		L	26.2	-0.40	26.6	27.6	%	2	
MCV			93.4	-0.10	93.5	93.9	fL	And a second second	
MCH			30.4	0.00	30.4	30.4	pg	the company of the second	200 a 100
MCHC			325	0.0	325	324	g/L	FL	SS
RDW-CV			15.2	-0.30	15.5	15.6	%		
RDW-SD			49.9	-0.90	50.8	51.4	fL	Fla	as an
PLT		L	45	5.0	40	38	10^9/L		
MPV			9.1	-0.40	9.5	9.6	fL	-WBC Abn Scattergram	
PDW			17.0	-0.30	17.3	16.6		-Blasts?	
РСТ			0.041	0.0030	0.038	0.036	%	-Immature Gran?	
P-LCC		L	10	0.0	10	10	10^9/L	-Left Shift?	
P-LCR			22.9	-2.60	25.5	26.6	%	-Monocytosis -Anemia	
NRBC#			0.022	-0.0280	0.050	0.039	10^9/L	-Anemia -Thrombocytopenia	
NRBC%			0.18	-0.180	0.36	0.24	/100WBC	monocytopenia	

CBC results on May 1

• WBC decrease was ongoing; RBC, HGB, and PLT were improving.

• Compared with the DIFF scattergram on Apr. 18, the number of particles in the Mon group decreased and the group location was significantly moving downward, indicating a decrease in abnormal promyelocytes; while in the Neu group, the number of particles increased and the group was extending towards the highly fluorescent region, indicating the presence of myelocytes and metamyelocytes.

Other examinations

		WBC	
	WBC	200	100%
L	Segmented neutrophil	17	8.5
L	Lymphocyte	20	10.0
н	Monocyte	95	47.5
!	Basophil	8	4.0
1	Myelocyte	18	9.0
1	Abnormal promyelocyte	42	21.0
	Non-WBC	18	%
	Nucleated RBC	1	0.5
	Large PLT	3	
	Smudge cell	12	6.0
	Sediment	2	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	34*10^9/L	Automated
	PLT concentration	34*10^9/L	Manual
		RBC	
	Size	Degree	%
!	Uneven erythrocyte sizes	2+	
	Macrocyte	0	0.4
!	Microcyte	2+	13.2
	Color	Degree	%
	Hypochromic RBC	0	3.7
	Polychromatic RBC	0	0.1
	Shape	Degree	%
1	Poikilocytosis	3+	
	Schistocyte	0	0.7
	Echinocyte	0	0.0
	Elliptocyte	0	0.2
!	Ovalocyte	3+	13.4
	Stomatocyte	0	0.0
	Leptocyte	0	0.0
	Dacryocyte	0	0.3
	Contents	Degree	%
	Basophilic stippling	0	0.0

Cells pre-classification by MC-80 on May 1



previous examination. Karyolysis was observed in a small number of cells.

🖹 Case analysis

Acute promyelocytic leukemia (APL) is a malignant disorder of hematopoietic tissues. It is characterized by the infinite proliferation of abundant leukemic cells in the bone marrow. It is the mostly observed malignant disease in the young people and may be caused by viruses, radioactive elements, chemical toxins, etc.

AMLL-M3 often has the following features:

- Promyelocyte cytoplasm is filled with abnormal particles
- Often accompanied with a bleeding tendency
- A large number of patients are PML/RARA fusion gene positive
- These patients are sensitive to chemotherapy, but the mortality is high in the early stage, especially when treated with cytotoxic chemotherapy, about 10%-20% of the patients died from hemorrhage during the treatment
- Retinoic acid can induce the differentiation of APL cells into mature cells, while arsenical agents can induce their apoptosis
- The sustained remission duration is long. In the past, the treatment effect and prognosis of APL was very poor, often due to severe bleeding caused by concurrent DIC or primary fibrinolysis

In this case, the patient had experienced fever, severe gingival bleeding, and ecchymosis in the lower extremities for over a month before receiving a diagnosis. The lack of immunophenotyping and genetic diagnosis may have led to a misdiagnosis of thrombocytopenic purpura. Bleeding is a common cause of mortality in APL patients, so timely identification of abnormal cells from CBC results can minimize the negative effects of delayed conditions and treatment.

Additionally, karolysis was observed at the initial diagnosis, which may have been due to an autoimmune deficiency in the patient. Leukemia patients often have malfunctions in "immune surveillance" and "self-stabilization," which can cause aging or mutant cells to accumulate in the body. In some cases, the body may even treat its own tissue cells as foreign bodies.

Case 02 Acute promyelocytic leukemia (AML-M3)

Clinical information

CBC results

A 53-year-old female patient had hemoptysis for 3 days, accompanied with gingival bleeding. She had no symptoms of pyrexia, epistaxis, gross hematuria, or melena. Nor did she have dizziness, headache, loss of consciousness, cough, productive cough, tightness of the chest, shortness of breath, palpitations, or pericordial pain. Her hemoptysis was persistent. Physical examinations: T: 36.3 °C, P: 78 bpm, R: 18 times/min, BP: 146/86 mmHg. The patient was in good general conditions, with clear breath sounds in both lungs. HR: 78 bpm. The heart rhythm was normal and auscultation of each valve showed no pathological murmur. The abdomen was soft, with no tenderness or rebound tenderness. The liver and the spleen were not palpable.

	CBC results on Apr. 28								
Parameter	Flaga	S	Result	Delta#	04-27	04-26	Unit	RBC	PLT
WBC	&		4.42	-0.510	4.93	3.31	10^9/L	nbc	1 61
Neu#	& R	L	1.30		****	0.72	10^9/L		
Lym#	& R		1.05		****		10^9/L		
Mon#	R	Н	2.02		****		10^9/L		
Eos#	R	L	0.00		****		10^9/L		
Bas#	R		0.05		****		10^9/L		
IMG#	R		0.12		****		10^9/L	0 100 200 fL	0 10 20 30 fL
Neu%	& R	L	29.4		****	21.8	%		0 10 20 50 12
Lym%	& R		23.8		****		%		
Mon%	R	Н	45.5		****	36.7	%		
Eos%	R	L	0.1		****		%	WNB	DIFF
Bas%	R	Н	1.2		****		%	FS	FL
IMG%	R		2.7		****		%		10 July 10 Jul
RBC		L	3.02	-0.250	3.27		10^12/L		ile.
HGB		L	94	-6.0	100	117	g/L	TAKE.	
НСТ		L	27.6	-2.30	29.9		%	and the second	
MCV			91.3	0.00	91.3		fL	A CONTRACTOR OF	
МСН			31.2	0.60	30.6		pg		
MCHC			342	7.0	335		g/L	FL	SS
RDW-CV			15.0	0.00	15.0		%		
RDW-SD			49.3	-0.20	49.5		fL	Ela	ags
PLT		L	54	43.0	11	19	10^9/L		.95
MPV			9.1	-2.60	11.7		fL	-Blasts?	
PDW			17.0	-1.10	18.1			-Abn Lymph/blasts?	
РСТ		L	0.049	0.0290	0.020		%	-Immature Gran	
P-LCC		L	11	7.0	4		10^9/L	-Atypical Lymph?	
P-LCR			19.5	-19.50	39.0		%	-NRBC	
NRBC#			0.054	-0.0290	0.083		10^9/L	-Monocytosis -Thrombocytopenia	
NRBC%			1.23	-0.460	1.69		/100WBC	monocytopenia	

CBC results on Apr. 28

- WBC was normal; Mon increased; mild anemia; PLT decreased.
- In the DIFF scattergram, the particles in the Neu group extended to the Mon group, while the particles in the Mon group significantly increased in number and were extending toward the highly fluorescent region, indicating the presence of abnormal cells.

Oracle Content Peripheral blood morphology examination

		WBC	
	WBC	200	100%
L	Segmented neutrophil	54	27.0
	Lymphocyte	50	25.0
L	Monocyte	2	1.0
	Basophil	1	0.5
!	Myelocyte	2	1.0
!	Myeloblast	8	4.0
!	Abnormal promyelocyte	83	41.5
	Non-WBC	29	%
	Nucleated RBC	3	1.5
	Large PLT	1	
	Smudge cell	21	10.5
	Sediment	4	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	43*10^9/L	Automated
	PLT concentration	62*10^9/L	Manual
		RBC	
	Size	Degree	%
1	Uneven erythrocyte sizes	3+	
	Macrocyte	0	0.7
	Microcyte	0	1.4
	Color	Degree	%
1	Hypochromic RBC	3+	57.5
	Polychromatic RBC	0	0.2
	Shape	Degree	%
	Poikilocytosis	0	
	Schistocyte	0	0.6
	Echinocyte	0	0.1
	Elliptocyte	0	0.0
	Ovalocyte	0	3.8
	Stomatocyte	0	0.8
	Leptocyte	0	0.0
	Dacryocyte	0	0.5
	Contents	Degree	%
	Basophilic stippling	0	0.0

Manual microscopic result

The microscopic examination showed presence of neutrophils (27%), lymphocytes (25%), monocytes (1%), and abnormal promyelocytes (45.5%).

The abnormal promyelocytes varied in cell body shape, with nuclei twisted, curved, and folded. In the cytoplasm were coarse azurophilic granules. The bundle-like Auer bodies were easily observable.

Other examinations

ltem	Parameter	Result	Reference range
	PT	14.60	10.5-14.5 s
	PT-INR	1.13	0.75-1.15
	PT activity%	81.00	70%-150%
	APTT	34.10	28-43.5 s
Coogulation	APTT ratio	1.00	0.8-1.28
Coagulation	TT	16.40	14-21 s
	TT ratio	0.96	0.8-1.24
	Fibrinogen	1.54	2-4 g/L
	Antithrombin III	85.00	80%-120%
	D-dimer	3280.00	0-500 ng/mL
MPO	١	+	
BMIS	١	Extracellular iron (+); Intracellular iron: erythroblasts, rare	
Bone marrow morphology	٨	Extremely active nucleated cell hyperplasia; presence of abnormal promyelocytes (82%), myelocytes and metamyelocytes were rarely observed. Suggested diagnosis: AML-M3a	

Case analysis

During the early stage of onset, the WBC count may still be normal in some M3 patients, hence leading to missed detection. Special alert should be paid to when abnormal Mon increase is observed. The abnormal cells are usually included in the Mon group due to high nucleic acid content and strong fluorescent signal. Special attention to the scattergram and alarms together with a comprehensive retesting rule can efficiently reduce the rate of missed detection.

Case 03 Acute myeloid leukemia (AML)

Clinical information

A 16-year-old female patient visited the local hospital due to pyrexia 2 weeks before. The CBC results showed WBC increased, Mon increased, mild anemia, and PLT decreased. The cell morphology examination showed presence of myeloblasts (46%). The patient was treated with hydroxy-urea to decrease WBCs and infused with PLTs, then transferred to our hospital.

CBC results on Mar. 8

G CBC results

	CDC TESUITS OFFINIAL. O								
Parameter	Flag	gs		Result	Delta#	02-22	Unit	RBC	PLT
WBC	&		н	14.90	-74.280	89.18	10^9/L	RDC	1 61
Neu#	&	R	L	0.55	-16.800	17.35	10^9/L		
Lym#	&	R	н	13.75	-14.980	28.73	10^9/L		
Mon#		R		0.58	-40.32	41.90	10^9/L		
Eos#			L	0.01	-0.990	1.00	10^9/L		
Bas#				0.01	-0.190	0.20	10^9/L		
IMG#		R		0.00	0.000	0.00	10^9/L		
Neu%	&	R	L	3.7	-15.80	19.5	%		
Lym%	&	R	н	92.2	60.00	32.2	%	0 100 200 fL	0 10 20 30 fL
Mon%		R		3.9	-43.10	47.0	%	0 100 200 12	0 10 20 30 12
Eos%			L	0.1	-1.00	1.1	%		
Bas%				0.1	-0.10	0.2	%		
IMG%		R		0.0	0.00	0.0	%	WNB	DIFF
RBC		R	L	2.74			10^12/L	WIND	DIT
HGB			L	84	4.0	80	g/L	FS	FL
НСТ		R	L	26.1			%		
MCV		R		95.1			fL		
MCH		R		30.5			pg		19 B
МСНС		R		321			g/L		
RDW-CV		R	н	17.2			%		
RDW-SD		R		50.0			fL		1
PLT	&	R	L	42	23.0	19	10^9/L		20 A A A A A A A A A A A A A A A A A A A
MPV		R		11.3			fL	FL	
PDW		R	н	17.6				FL.	SS
РСТ		R	L	0.055			%		
P-LCC		R	L	16			10^9/L	RET	Flags
P-LCR		R		38.1			%	THE T	i lags
IPF		R	н	12.0			%	FS	-WBC Abn Scattergram
RET#		R		0.10000			10^12/L	a second seco	-Blasts?
RET%			н	3.64			%		-Abn Lymph/blast?
IRF			н	37.2			%		-NRBC
LFR			L	62.8			%		-Lymphocytosis -Neutropenia
MFR				15.1			%		-Neutropenia -RBC
HFR			н	22.1			%	E the second second	-Anemia
RHE			н	40.3			Pg	A STATE OF A	-PLT Clump?
NRBC#				0.622			10^9/L		-Thrombocytopenia
NRBC%				4.18			/100WBC	FL	

• Compared with 2 weeks before, WBC significantly decreased, especially Neu and Mon, indicating effective WBC reduction; HGB and PLT were slightly improved; IRF increased, indicating normal erythroid hyperplasia.

• In the DIFF scattergram, the number of particles significantly decreased in the Neu group and Eos group; a large quantity of particles existed between the Lym group and Mon group, leading to a fusion of Lym and Mon particles, showing a "long eggplant" shape. The RET scattergram showed a long tail of RBC particles downward, indicating fragmented RBCs.

🖑 Peripheral blood morphology examination

		WBC	
	WBC	100	100%
L	Segmented neutrophil	2	2.0
	Lymphocyte	34	34.0
L	Monocyte	1	1.0
	Blast	63	63.0
	Non-WBC	37	%
	Nucleated RBC	13	13.0
	Giant PLT	3	
	Large PLT	1	
	Smudge cell	10	10.0
	Sediment	10	
		PLT	
	PLT estimate	Estimated result	Estimation metho
	PLT concentration	48*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	0	
	Macrocyte	0	1.9
	Microcyte	0	1.8
	Color	Degree	%
	Hypochromic RBC	3+	67.5
	Polychromatic RBC	0	1.6
	Shape	Degree	%
	Poikilocytosis	1+	
	Schistocyte	1+	2.4
	Echinocyte	0	1.0
	Elliptocyte	0	0.0
	Ovalocyte	0	4.0
	Stomatocyte	0	0.1
	Leptocyte	0	0.6
	Dacryocyte	0	1.9
	Contents	Degree	%
	Basophilic stippling	0	0.0



Manual microscopic result

The microscopic examination showed presence of myeloblasts (63%) and nucleated red blood cells 15 cells/100 WBC.

The myeloblasts had large oval cell bodies. The nuclei were large, round or oval, and irregular shapes were occasionally observed; the nuclear chromatin was fine, some had visible nucleoli. Scant cytoplasm appeared pale blue, and a very small number of particles were observed.

Other examinations

ltem	Result
Thalassemia gene mutation detection	Not detected
MDS FISH	Not detected
Bone marrow morphology	Markedly active bone marrow hyperplasia; myeloid (1%); erythroid (30%); decreased lymphocyte proportion, with normal shape; normal monocyte proportion and shape; presence of myeloblasts (63%); POX (+); compliant with the microscopic bone marrow images of acute myeloid leukemia.

Case analysis

This case was primarily diagnosed as Acute myeloid leukemia (AML). Further examinations are required, including flow cytometry, AML-related gene detection, fusion gene detection, and chromosomal examination.

Based on the peripheral blood morphology examination, the abnormal particles in the Lym and Mon regions should be myeloblasts in the CBC DIFF scattergram. These cells had low nucleic acid content due to WBC reduction therapy and were wrongly counted as Lym and Mon.

Case 04 Acute myeloid leukemia with maturation (AML-M2)

E Clinical information

A 47-year-old male patient, previously healthy, initially visited the local hospital in the early October 2022 due to pyrexia accompanied with sore throat. The highest body temperature measured was 39 °C. His body temperature returned to normal after treatment with erythromycin in local hospital. On Oct. 14, the patient reexperienced pyrexia. The CBC showed WBC: 2.59*10^9/L, HGB: 72 g/L, PLT: 27*10^9/L. The patient was treated with cefuroxime (i.v.) but did not recover, and was also treated with anti-infective therapy (piperacillin, etc.).

On Nov. 8, the patient visited the outpatient clinic due to pyrexia. The chest CT showed multiple exudative lesions in the left lower lung, subpleural atelectasis of lung tissue on the dorsal side of the right lung, and a small amount of left pleural effusion.

				CE	3C results on Nov. 8	
Parameter	Flags		Result	Unit	RBC	PLT
WBC			5.02	10^9/L		
Neu#	R	L	0.06	10^9/L		
Lym#	R	н	4.91	10^9/L		
Mon#	R	L	0.02	10^9/L		
Eos#			0.03	10^9/L		
Bas#			0.00	10^9/L		
IMG#	R		0.00	10^9/L		
Neu%	R	L	1.1	%	01 00 200 fL	0 10 20 30
Lym%	R	н	97.9	%		
Mon%	R	L	0.4	%		
Eos%			0.5	%	WNB	DIFE
Bas%			0.1	%	WIND	DIFF
IMG%	R		0.1	%	FS	
RBC		L	1.75	10^12/L		FL
HGB		L	58	g/L		270 247
HCT		L	17.1	%		
MCV			97.8	fL		
MCH			33.0	pg	A CONTRACTOR OF	
MCHC			337	g/L	and the second	
RDW-CV		Н	23.3	%	FL I	
RDW-SD		Н	83.2	fL	PL .	
PLT		L	15	10^9/L		
MPV			9.6	fL	FI	ags
PDW			16.7		-WBC Abn Scattergram	-Anemia
РСТ		L	0.015	%	-Abn Lymph/blast?	-Thrombocytopenia
P-LCC		L	4	10^9/L	-Lymphocytosis	Infombocytopenia
P-LCR			24.9	%	-Neutropenia	
NRBC#			0.000	10^9/L	-Dimorphic population	
NRBC%			0.00	/100WBC		

CBC results

• WBC normal, but Lym abnormally increased; severe anemia; PLT extremely decreased.

• In the DIFF scattergram, the Lym particles fused with the Mon particles and extended towards the highly fluorescent region, appearing as a "long eggplant" or "baseball bat".

C Peripheral blood morphology examination

	W	/BC	
	WBC	200	100%
L	Segmented neutrophil	1	0.5
	Lymphocyte	68	34.0
	Myeloblast	104	52.0
	Reactive lymphocyte	27	13.5
	Non-WBC	56	%
	Large PLT	3	
	PLT aggregation	1	
	Smudge cell	26	13.0
	Sediment	26	
	P	PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	46*10^9/L	Manual
	R	BC	
	Size	Degree	%
	Uneven erythrocyte sizes	2+	
	Macrocyte	0	3.9
	Macrocyte Microcyte	0 0	3.9 4.9
		-	
	Microcyte	0	4.9
	Microcyte Color	0 Degree	4.9 %
	Microcyte Color Hypochromic RBC	0 Degree 0	4.9 % 0.3
	Microcyte Color Hypochromic RBC Polychromatic RBC	0 Degree 0 0	4.9 % 0.3 0.0
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape	0 Degree 0 0 Degree	4.9 % 0.3 0.0
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis	0 Degree 0 0 Degree 0	4.9 % 0.3 0.0 %
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte	0 Degree 0 0 Degree 0 0	4.9 % 0.3 0.0 %
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte	0 Degree 0 Degree 0 0 0	4.9 % 0.3 0.0 %
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte	0 Degree 0 Degree 0 0 0 0 0 0 0 0 0 0 0 0 0	4.9 % 0.3 0.0 %
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Polikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte	0 Degree 0 Degree 0 0 0 0 0 0 0 0 0 0 0 0 0	4.9 % 0.3 0.0 % 0.7 0.7 0.1 0.6 10.8
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte	0 Degree 0 Degree 0 0 0 0 0 0 0 0 0 0 0 0 0	4.9 % 0.3 0.0 % 0.7 0.1 0.6 10.8 0.0
	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte Leptocyte	0 Degree 0 Degree 0 0 0 0 0 0 0 0 0 0 0 0 0	4.9 % 0.3 0.0 % 0.7 0.1 0.6 10.8 0.0 0.0



Manual microscopic result

Presence of myeloblasts (60%) with large round cell bodies and large, round or oval nuclei with fine and sand-like chromatins. Some had visible nucleoli and scant cytoplasm that appeared pale blue; thin and long single Auer body can be seen in some cytoplasm; vacuoles were observable in individual cytoplasm.

Other examinations

ltem	Result
Bone marrow morphology	Markedly active bone marrow hyperplasia, M:E = 13.54:1. Markedly blasts portion increased (approx. 88%); Auer bodies were easily recognized in cytoplasm. A total of 13 megakaryocytes were counted and no megakaryocyte platelet was observed.
Bone marrow immunophenotyping	A group of abnormal cells can be seen in the myeloid blasts extended distribution area, accounting for approx. 88.6% of nucleated cells, expressing CD7, CD13, CD34, CD38, CD117, and MPO; some cells expressed HLA-DR, CD5, and CD64. Suggested diagnosis: AML (M2 possible)
Chromosome	47, XY, +10[10]. Leukemia-related fusion gene detection was all negative. Gene mutation showed that IDH1 detected missense mutation c.394C>T(p.Arg132Cys) (mutation frequency 44.52%).

Case analysis

Auer bodies are commonly observed in AML-M2, APL, MDS, and MPN. This case can be diagnosed as AML-M2 with reference to the flow cytometry and chromosomal genotype results. For M2 patients, their peripheral blood WBCs may be increased, normal, or decreased. Special attention should be paid to the scattergram and alarms. Retesting should be timely conducted when abnormal distributions of Lym and Mon particles are observed.

Case 05 Acute myeloid leukemia (AML)

E Clinical information

A 60-year-old male patient visited the Department of Hematology and was discovered pancytopenia.

G CBC results

Parameter	Fla	igs		Result	Unit	RBC	PLT		
WBC	&		L	1.66	10^9/L				
Neu#	&	R	L	0.17	10^9/L				
Lym#	&	R		1.22	10^9/L				
Mon#		R		0.24	10^9/L				
Eos#				0.02	10^9/L				
Bas#				0.01	10^9/L				
IMG#		R		0.00	10^9/L	0 100 200 fL	0 10 20 30 fL		
Neu%	&	R	L	10.5	%				
Lym%	&	R	н	73.0	%	MAID	DIFF		
Mon%		R	н	14.7	%	WNB	DIFF		
Eos%				1.2	%	FS	DIFF 3X		
Bas%				0.6	%		4		
IMG%		R		0.0	%				
RBC			L	2.22	10^12/L	and the second second	a 👔 🦉 🖉 👘 👘		
HGB			L	77	g/L				
НСТ			L	22.9	%				
MCV			н	102.9	fL	FL	SS		
МСН			н	34.6	pg				
МСНС				336	g/L	RET	Flags		
RDW-CV			н	17.4	%				
RDW-SD			н	63.9	fL	FS	-Pancytopenia -Blasts?		
PLT	&		L	40	10^9/L	and the state of the	-Blasts? -Abnormal Lymph/blast?		
RET#				0.0205	10^12/L		-NRBC		
RET%				0.92	%		-Neutropenia		
IRF				23.8	%	and the second sec	-Leukocytopenia -Anemia		
NRBC#				0.103	10^9/L	FL	-Thrombocytopenia		
NRBC%				6.20	/100WBC				

• WBC decreased, the retest rule was triggered, Lym% and Mon% increased; HGB and PLT both decreased.

• The DIFF scattergram showed that the Mon particles were in the highly fluorescent region, half a group position higher than the normal Mon group, which were suspected as abnormal cells.

C Peripheral blood morphology examination

		WBC	
	WBC	189	100%
	Segmented neutrophil	13	6.9
	Band neutrophil	2	1.1
ł	Lymphocyte	141	74.7
	Monocyte	8	4.2
	Basophil	1	0.5
	Metamyelocyte	1	0.5
	Myeloblast	15	7.9
	Reactive lymphocyte	3	1.6
	Others	5	2.6
	Non-WBC	54	%
	Nucleated RBC	2	1.1
	Giant PLT	9	
	Large PLT	7	
	Smudge cell	15	13.2
	Sediment	21	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	41*10^9/L	Automated
	PLT concentration	58*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	2+	
	Macrocyte	0	4.8
	Microcyte	0	5.2
	Color	Degree	%
	Hypochromic RBC	0	0.1
	Polychromatic RBC	0	0.0
	1 ofgenionatie noe	-	010
	Shape	Degree	%
	•		
	Shape	Degree	
	Shape Poikilocytosis	Degree 3+	%
	Shape Poikilocytosis Schistocyte	Degree 3+ 0	% 0.3
	Shape Poikilocytosis Schistocyte Echinocyte	Degree 3+ 0	% 0.3 0.0
	Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte	Degree 3+ 0 0 0	% 0.3 0.0 1.1
	Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte	Degree 3+ 0 0 0 3+	% 0.3 0.0 1.1 25.6
	Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte	Degree 3+ 0 0 3+ 0 0 3+ 0 0 3+ 0 3+ 0	% 0.3 0.0 1.1 25.6 0.0
	Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte Leptocyte	Degree 3+ 0 0 3+ 0 3+ 0 3+ 0 0 0 0 0 0 0 0 0	% 0.3 0.0 1.1 25.6 0.0 0.0



Manual microscopic result

Presence of myeloblasts (7.9%), with uneven cell body sizes, most round or oval; moderate amount of cytoplasm, which appeared blue or dark blue, with darker edges; a small number of cells contained few azurophilic granules; the nuclei were round or oval and few were retracted, folded, or kidney-shaped; the chromatins were fine and evenly distributed, some had visible nucleoli.

Other examinations

ltem	Result
Bone marrow morphology	Markedly active bone marrow hyperplasia, M:E = 0.48:1; presence of myeloblasts (35.0%); POX: 13% positive. Suggested diagnosis: microscopic bone marrow images of acute leukemia.
Flow cytometry	Presence of abnormal immature myeloid cells (29.2%); expressing: CD38, CD117, CD13, CD33 bri; partially expressing: CD7; not expressing: MPO, CD15, CD64, CD11b, CD11c, CD56, CD19, CD24, cCD79a. Suggested diagnosis: AML
Gene detection	A total of 42 leukemia-related fusion genes were detected: WT1 copy number 855337.0, ABL copy number 3518728, WT1 quantitative measurement 24.3081%, the rest were all negative. The myeloid leukemia prognostic gene detection showed NPM1 positive.

Case analysis

NPM1 mutation is one of the most common gene mutations in AML. NPM1 protein is a nucleolar protein. The mutant protein can be released from the nucleoli into cytoplasm, hence resulting in AML. Studies have shown that the NPM1 mutant protein (NPM1c) can bind to the chromatin, induce Hox gene high expression, and promote the occurrence of AML. At present, there is no specific treatment regimen for NPM1 gene mutant AML. The treatment is mainly referring to the clinical treatment regimen for AML.

Case 06 Acute myeloid leukemia (AML)

Clinical information

A 68-year-old male patient had dizziness for 2 months and aggravated for 3 days. The physical examinations in April reported anemia and platelets decreased. No symptoms of melena or active hemorrhage were observed.

G CBC results

Parameter	Fla	gs		Result	Unit
WBC			L	2.37	10^9/L
Neu#		R	L	1.37	10^9/L
Lym#		R	L	0.37	10^9/L
Mon#		R		0.62	10^9/L
Eos#			L	0.00	10^9/L
Bas#				0.01	10^9/L
IMG#		R		0.01	10^9/L
Neu%		R		57.7	%
Lym%		R	L	15.5	%
Mon%		R	н	26.4	%
Eos%				0.1	%
Bas%				0.3	%
IMG%		R		0.6	%
RBC			L	1.91	10^12/L
HGB			L	51	g/L
НСТ			L	15.9	%
MCV				83.0	fL
МСН			L	26.4	pg
MCHC			L	318	g/L
RDW-CV			н	17.6	%
RDW-SD				51.9	fL
PLT	&	R	L	26	10^9/L
RET#			L	0.0083	10^12/L
RET%				0.44	%
IRF				14.1	%
NRBC#				0.000	10^9/L
NRBC%				0.00	/100WBC











• Pancytopenia, Mon% increased.

• In the DIFF scattergram, the Mon group extended from lower left to upper right, which were suspected as abnormal cells; in the RET scattergram, there were few PLT particles and most of them were located in the highly fluorescent region, indicating low PLT count and predominantly immature PLTs.

FS

		WBC	
	WBC	223	100%
	Segmented neutrophil	110	49.3
	Band neutrophil	2	0.9
	Lymphocyte	30	13.5
_	Monocyte	2	0.9
	Metamyelocyte	4	1.8
	Myeloblast	75	33.6
	Non-WBC	187	%
	Giant PLT	18	
	Large PLT	43	
	Smudge cell	21	9.4
	Sediment	92	
	Megakaryocyte	13	5.8
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	41*10^9/L	Automated
	PLT concentration	58*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	2+	
	Macrocyte	0	0.9
	Microcyte	2+	17.8
	Color	Degree	%
	Hypochromic RBC	2+	12.6
	Polychromatic RBC	0	0.0
	Shape	Degree	%
	Poikilocytosis	3+	
	Schistocyte	3+	4.8
	Echinocyte	0	1.6
	Elliptocyte	0	1.8
	Ovalocyte	3+	26.0
	Stomatocyte	0	0.0
	Leptocyte	0	0.0
	Dacryocyte	0	3.1
	Contents	Degree	%
	Basophilic stippling	0	0.0

Manual microscopic result

Presence of myeloblasts (33.6%), with uneven cell body sizes, most round or oval; moderate amount of cytoplasm, which appeared caeruleous, with darker edges; no azurophilic granules were observed; the nuclei were round or oval; the chromatins were fine, some had 1–2 visible nucleoli. Presence of megakaryocytes (5.8%).



Other examinations

ltem	Result					
Bone marrow morphology	Decreased bone marrow hyperplasia, M:E = 10.57; presence of myeloblasts (23.0%); myeloids accounted for approx. 37.0% of total nucleated cells; a total of 27 megakaryocytes were observed, and binucleated, multinucleated, and micro-megakaryocytes were observed. POX negative, NAE weakly positive, NAE-NaF inhibition, NAP score: 256; Suggested diagnosis: microscopic bone marrow images of acute leukemia, the cell morphology and cytochemical staining features were unclear, please refer to flow cytometry, gene detection, and other examinations for further comprehensive diagnosis and treatment.					
Flow cytometry	Abnormal immature myeloid cells (34.6%). Expressing: CD34, CD117, CD13, HLA-DR, CD7, CD123, etc.; not expressing: MPO, CD15, CD38, CD14, CD56, CD4, CD24, cCD79a, CD16, CD19, CD41, CD42B. Suggested diagnosis: AML possible.					
Gene detection	A total of 42 leukemia-related fusion genes were detected: WT1 copy number 814.00, ABL copy number 32462.00, WT1 quantitative measurement 2.5075%, the rest were all negative. The myeloid leukemia prognostic gene detection showed NPM1 negative.					

Case analysis

The patient was diagnosed with Acute myeloid leukemia (AML). Normally, megakaryocytes cannot pass through the blood sinusoids and appear in the peripheral blood. When there are megakaryocytes in the peripheral blood, it may be due to:

- Excessive production of megakaryocytes caused by myeloproliferative disorders;
- Bone-marrow barrier damage caused by inflammation, tumor factors, or other reasons;
- Extramedullary hematopoiesis;
- Impaired splenic clearance;
- Abnormalities in apoptosis or apoptosis-inducing factors.

Case 07 Acute myelomonocytic leukemia (AMMOL)

E Clinical information

The patient was a 52-year-old female.

G CBC results

Parameter	Flags		Result	Unit
WBC		L	1.55	10^9/L
Neu#	R	L	0.25	10^9/L
Lym#	R	L	0.66	10^9/L
Mon#	R		0.63	10^9/L
Eos#		L	0.01	10^9/L
Bas#			0.00	10^9/L
IMG#	R		0.01	10^9/L
Neu%	R	L	16.4	%
Lym%	R	н	41.9	%
Mon%	R	н	40.8	%
Eos%			0.8	%
Bas%			0.1	%
IMG%	R		0.4	%
RBC		L	1.48	10^12/L
HGB		L	52	g/L
НСТ		L	15.1	%
MCV		н	101.7	fL
МСН		н	34.8	pg
MCHC			342	g/L
RDW-CV		н	16.8	%
RDW-SD		н	60.0	fL
PLT		L	34	10^9/L
MPV			8.5	fL
PDW			15.9	
РСТ		L	0.029	%
P-LCC		L	4	10^9/L
P-LCR			13.2	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC





Flags



PLT

20

30

fL

10

-Pancytopenia -Blasts? -Abn Lymph/blast? -Atypical Lymph?

• Pancytopenia, Lym% and Mon% increased.

• DIFF-3X was initiated due to WBC decreased. In the DIFF scattergram, the Lym particles fused with the Mon particles and extended towards the highly fluorescent region, appearing as a "pencil" shape, indicating presence of myeloblasts.

🖑 Peripheral blood morphology examination

	WBC						
WBC	110	100%					
Segmented neutrophil	3	2.7					
Band neutrophil	2	1.8					
Lymphocyte	26	23.6					
Monocyte	7	6.4					
Metamyelocyte	1	0.9					
Myeloblast	71	64.6					
Non-WBC	23	%					
Nucleated RBC	1	0.9					
Smudge cell	1	0.9					
Sediment	21						
PLT							
PLT estimate	Estimated result	Estimation method					
PLT concentration	40*10^9/L	Automated					
PLT concentration	22*10^9/L	Manual					
	RBC						
Size	Degree	%					
Uneven erythrocyte sizes	0						
Macrocyte	0	0.3					
Microcyte	3+	45.1					
Color	Degree	%					
Hypochromic RBC	3+	73.2					
Polychromatic RBC	0	0.2					
Shape	Degree	%					
Poikilocytosis	2+						
Schistocyte	0	0.3					
Echinocyte	0	0.2					
Elliptocyte	0	0.2					
Ovalocyte	2+	5.7					
Stomatocyte	0	0.1					
Leptocyte	0	0.1					
Dacryocyte	0	0.3					
Contents	Degree	%					
Basophilic stippling	0	0.1					



Manual microscopic result

Presence of myeloblasts (65%), with large cell body and irregular shapes; the chromatins were coarse particles, with 1–2 large and pronounced nucleoli; the cytoplasm was dark blue, and cytoplasm of some cells contained dust-like granules.

Other examinations

ltem	Result
Bone marrow morphology	Extremely active bone marrow hyperplasia; presence of leukemia cells (73%), with round cell bodies, round nuclei, some twisted or folded, the chromatins were fine. 1–3 nucleoli were visible. Scant cytoplasm appeared dusty blue. Fine purple particles were visible in some cells. Cytochemical staining: POX (+), PAS (+), AS-DCE (partial +), a-NAE (+), NAF (partial inhibition), a-NBE (partial +); the eosinophil portion increased (16%), particle decreased was observed in some cells, uneven amount of basophilic particles were visible. Suggested diagnosis: M4EO.
Flow cytometry	Presence of abnormal myeloid blasts (61.19%) (nucleated cells). Expressing: CD34, CD117, CD13, CD33, CD38, cMPO; partially expressing: CD15, HLA-DR, CD64; not expressing: CD10, CD7, CD19, CD56, cCD3, cCD79a, etc. Compliant with the immunophenotype of AML, probable AML-M5 or M4.
Chromosomal examination	47, XX, inv(16)(p13q22), +22[14]/46, XX, inv(16)(p13q22)[6]

Case analysis

A unique type of AML is acute myelomonocytic leukemia with eosinophilia (AML-M4EO). This subtype is an M4 with good prognosis, so increasing the detection rate of AML-M4EO is of great clinical significance. Most M4EO cases are indicated with abnormalities of chromosome 16 structure, and the more distinct and common type is chromosome 16 inversion. inv(16) is a characteristic abnormal karyotype of M4EO. It is involved in the formation of CBF β -MYH11 fusion gene, which can be used as a molecular biology indicator for M4EO. This has important clinical value for the clinical diagnosis and prognosis prediction of M4.

Case 08 Acute monocytic leukemia (AMOL)



Clinical information

A 52-year-old male patient was admitted to the hospital due to infection accompanied by anemia.

G CBC results

Parameter	Ala	rm		Result	Unit
WBC	&		Н	14.87	10^9/L
Neu#	&	R		4.00	10^9/L
Lym#	&	R		3.01	10^9/L
Mon#		R	Н	7.83	10^9/L
Eos#				0.03	10^9/L
Bas#				0.00	10^9/L
IMG#		R		0.56	10^9/L
Neu%	&	R	L	26.9	%
Lym%	&	R		20.2	%
Mon%		R	Н	52.7	%
Eos%			L	0.2	%
Bas%				0.0	%
IMG%		R		3.8	%
RBC			L	1.76	10^12/L
HGB			L	63	g/L
НСТ			L	20.3	%
MCV			Н	115.2	fL
МСН			н	35.8	pg
MCHC			L	311	g/L
RDW-CV			н	17.9	%
RDW-SD			н	72.8	fL
PLT			L	91	10^9/L
MPV				9.7	fL
PDW				16.9	
PCT			L	0.088	%
P-LCC			L	21	10^9/L
P-LCR				23.6	%
NRBC#				0.257	10^9/L
NRBC%				1.73	/100WBC



• WBC increased, mainly Mon; HGB decreased; low PLT count.

• In the WNB scattergram, marked red particles were visible, indicating nucleated red blood cells; in the DIFF scattergram, the number of particles markedly increased in the Mon region and most particles were in the highly fluorescent region, indicating presence of immature monocytes.

C Peripheral blood morphology examination

		WBC	
	WBC	110	100%
L	Segmented neutrophil	17	15.5
	Band neutrophil	3	2.7
	Lymphocyte	38	34.5
н	Monocyte	45	41.0
!	Myelocyte	2	1.8
1	Myeloblast	5	4.5
	Non-WBC	45	%
1	Nucleated RBC	6	5.5
	Giant PLT	1	
	Large PLT	9	
	Smudge cell	6	5.5
	Sediment	23	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	90*10^9/L	Automated
	PLT concentration	50*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	0	
			2.0
	Macrocyte	0	3.9
	Macrocyte Microcyte	0 0	3.9 10.3
!	Microcyte	0	10.3
!	Microcyte Color	0 Degree	10.3 %
!	Microcyte Color Hypochromic RBC	0 Degree 3+	10.3 % 40.8
! ! !	Microcyte Color Hypochromic RBC Polychromatic RBC	0 Degree 3+ 2+	10.3 % 40.8 6.4
! ! !	Microcyte Color Hypochromic RBC Polychromatic RBC Shape	0 Degree 3+ 2+ Degree	10.3 % 40.8 6.4
! ! !	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis	0 Degree 3+ 2+ Degree 1+	10.3 % 40.8 6.4 %
! ! !	 Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte 	0 Degree 3+ 2+ Degree 1+ 1+	10.3 % 40.8 6.4 %
! ! !	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte	0 Degree 3+ 2+ Degree 1+ 1+ 0	10.3 % 40.8 6.4 % 0.7 0.7
! ! !	Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte	0 Degree 3+ 2+ Degree 1+ 1+ 0 0	10.3 % 40.8 6.4 % 0.7 0.7 0.1 0.2
	 Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte 	0 Degree 3+ 2+ Degree 1+ 1+ 0 0 0 0	10.3 % 40.8 6.4 % 0.7 0.7 0.1 0.2 3.3
	 Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte 	0 Degree 2+ Degree 1+ 1+ 0 0 0 0 0	10.3 % 40.8 6.4 % 0.7 0.7 0.1 0.2 3.3 0.5
1	 Microcyte Color Hypochromic RBC Polychromatic RBC Shape Poikilocytosis Schistocyte Echinocyte Elliptocyte Ovalocyte Stomatocyte Leptocyte 	0 Degree 2+ Degree 1+ 0 0 0 0 0 0 0 0 0 0 0 0 0	10.3 % 40.8 6.4 % 0.7 0.1 0.2 3.3 0.5 1.1

Manual microscopic result

Results from manual re-differentiation: Myeloblasts accounted for 45.5%.

The abnormal cells can be differentiated into two clusters morphologically:

One cluster of the cells had large cell bodies, round or irregular; the chromatins were fine and loose, the nuclei were retracted, twisted, or folded, like wrinkles, the nucleoli were barely visible; the cytoplasm was blue, with no granules inside but vacuoles visible. These cells were probably monoblasts.

The other cluster of the cells had large cell bodies, most were irregular; the nuclei were twisted or folded in a more marked manner, the nucleoli were barely visible; the cytoplasm was dusty blue, with azurophilic granules and vacuoles visible. These cells were probably promonocytes. The automated cell morphology analyzer wrongly differentiated most monoblasts and promonocytes as monocytes.

Other examinations

ltem	Result
Bone marrow morphology	Active bone marrow hyperplasia; presence of leukemia cells (54%), cytochemical staining results: POX (+); PAS (+), AS- DCE (+), a-NAE (+), NAF (inhibition), a-NBE (+). Suggested diagnosis: AML-M5 probable.
Flow cytometry	CD34+CD117+ myeloid blasts accounted for 0.37% of the nucleated cells, with no marked abnormality in phenotypes. Immature monocytes accounted for 26.5% of the nucleated cells. Mature monocytes accounted for 35.17% of the nucleated cells. Granulocytes accounted for 15.43% of the nucleated cells. Nucleated red blood cells accounted for 14.98% of the nucleated cells. The results were compliant with the immunophenotype of AML. Suggested diagnosis: AML-M5 possible.

Case analysis

This patient was diagnosed with AML-M5.

AML-M5 is acute monocytic leukemia and can be classified into two subcategories:

• M5a (undifferentiated): In the bone marrow, marked hyperplasia of monoblasts can be observed, accounting for not less than 80% (NEC), and promonocytes are rarely observed. The leukemia cells are uniformed, quasi-circular or irregular shaped. The nuclei are retracted or folded, the chromatins are loose, fine, and net-like. The nucleoli are clearly visible. The cytoplasm is dusty blue, with pseudopods. Thin and long Auer bodies can be observed in some cytoplasm.

M5b (partially differentiated): In the bone marrow, leukemia cells of variable differentiation mononuclear lineage can be seen. Monoblasts account for less than 80% (NEC), and promonocytes are significantly more than that in M5a, reaching more than 20%. The leukemia cells vary in size, are irregular in shape, and can be with a lagged tail; The nuclei are folded and are shown in various shapes, such as pen-holder, horseshoe, kidney-like, S-shape, etc., the chromatins are reticular or cord-like, the cell mass is rich in dusty blue, purple-red granules at different thickness are visible, and obvious pseudopods can be observed.

Hepatomegaly, splenomegaly, lymphodenopathy, and hemorrhage are mainly observed in clinical practice.

Clinical manifestations:

- Gingival hyperplasia, swelling, bleeding, ulcers, necrosis, etc., are common;
- Nasal mucosal infiltration, nasal obstruction, hyposmia, palatal ulcers, asphyxia caused by pharyngeal/laryngeal edema, etc.
- Leukemia cutis lesions are common, with manifestations of diffuse macular/papular, hard nodules, masses, dermatitis pustular/bullous/exfoliative, etc.
- Intestinal wall infiltration, ulcer, gastrointestinal dysfunction, etc., are relatively more observed;
- Renal failure and proteinuria are common, which are related to the formation of lysozyme anemia and lysozyme urine caused by the rich lysozymes in monocytes and granulocytes;
- Joint pain and swelling are relatively common;
- The treatment efficacy of acute myelomonocytic leukemia and acute monocytic leukemia is poor compared with the other acute non-lymphocytic leukemias.

Case 09 B-lymphoblastic lymphoma

Clinical information

A 64-year-old male patient was admitted to the hospital due to "mass in the left subcostal region for more than 2 months". The upper abdominal CT (dynamic with contrast) showed markedly enlarged spleen. The lower abdominal CT (dynamic with contrast) showed multiple enlarged retroperitoneal lymph nodes.

Sec results

Parameter	Flags		Result	Unit
WBC		Н	10.91	10^9/L
Neu#	R	L	1.57	10^9/L
Lym#	R	Н	7.66	10^9/L
Mon#	R	Н	1.59	10^9/L
Eos#			0.05	10^9/L
Bas#			0.04	10^9/L
IMG#	R		0.00	10^9/L
Neu%	R	L	14.4	%
Lym%	R	Н	70.1	%
Mon%	R	Н	14.6	%
Eos%			0.5	%
Bas%			0.4	%
IMG%	R		0.0	%
RBC			4.06	10^12/L
HGB			129	g/L
НСТ		L	38.8	%
MCV			95.5	fL
МСН			31.8	pg
MCHC			332	g/L
RDW-CV			12.9	%
RDW-SD			45.2	fL
PLT	&	L	81	10^9/L
MPV			11.0	fL
PDW			16.8	
PCT		L	0.089	%
P-LCC		L	26	10^9/L
P-LCR			32.4	%
IPF			5.4	%
RET#			0.0840	10^12/L
RET%			12.07	%
IRF			13.7	%
LFR			86.3	%
MFR			11.5	%
HFR			2.2	%
RHE			31.5	pg
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC













PLT

20

DIFF

10

0

fL

30

- -Abn Lymph/blast -Atypical Lymph? -Monocytosis -Lymphocytosis
- WBC increased, Lym and Mon significantly increased; erythroids normal; low PLT count.
- In the DIFF scattergram, the Lym particles extended towards the highly fluorescent region, appearing as a "knife", the abnormal cells in the higher region were counted as Mon.

C Peripheral blood morphology examination

	14	BC	
	WBC	200	100%
L		200	12.0
2	Segmented neutrophil	24 50	25.0
L	Lymphocyte	5	2.5
L	Monocyte	2	
	Eosinophil	2	1.0 0.5
1	Basophil	88	0.5 44.0
	Myeloblast		
!	Reactive lymphocyte Non-WBC	30	15.0 %
		66	%
	Giant PLT	1	
	Large PLT	1	
	Smudge cell	18	9.0
	Sediment	46	
	· · · · · · · · · · · · · · · · · · ·	LT Estimated	Estimation
	PLT estimate	esult	ethod
	PLT concentration	64*10^9/L	Automated
	PLT concentration	80*10^9/L	Manual
	R	BC	
	Size	Degree	%
!	Uneven erythrocyte size	s 2+	
	Macrocyte	0	0.4
!	Microcyte	3+	0.1
	Color	Degree	%
!	Hypochromic RBC	2+	11.2
	Polychromatic RBC	0	0.1
	Shape	Degree	%
	Poikilocytosis	0	
	Schistocyte	0	0.2
	Echinocyte	0	7.1
	Elliptocyte	0	1.2
	Ovalocyte	0	10.5
	Stomatocyte	0	0.0
	Leptocyte	0	0.0
		0	0.5
	Dacryocyte	0	0.5
	Dacryocyte Contents	Degree	%



Manual microscopic result

The re-differentiation showed presence of lymphocytes (25%) and abnormal lymphocytes (59%).

These abnormal lymphocytes had small cell bodies and irregular shape. The chromatins were more fine compared with the normal lymphocytes, and nucleoli were visible in some cells. Scant cytoplasm appeared blue.

Other examinations

ltem	Result
Bone marrow morphology	Active nucleated cell hyperplasia in bone marrow; myeloid hyperplasia was poor, mainly metamyelocytes; presence of immature lymphocytes (28.5%) and mature lymphocytes (41.5%), where abnormal lymphocytes were common.
Flow cytometry	Abnormal shape of B lymphocytes were observed, FSC and SSC were not strong, compliant with the signs of CD5-CD10-B cells, and not compliant with HCL phenotype.
Pathological diagnosis	CD5-CD10- small B-cell lymphoma

Case analysis

The patient had enlarged spleen and multiple enlarged retroperitoneal lymph nodes. In combination with the peripheral blood morphology results, the primary diagnosis was small B-cell lymphoma. The diagnosis of CD5-CD10- small B-cell lymphoma was confirmed by flow cytometry. Further genetic detections, lymph node biopsy, and other relevant examinations are required for immunophenotyping.

Case 10 T-lymphoblastic lymphoma

Clinical information

A 9-year-old female patient was admitted to the hospital due to "asthenia for 10 days, WBC increased for 1 day".

G CBC results

	CBC results on Aug. 4								
Parameter	Fla	gs		Result	Unit	RBC	PLT		
WBC	&		Н	114.76	10^9/L	ind c			
Neu#	&	R	Н	34.00	10^9/L				
Lym#				****	10^9/L	1			
Mon#				****	10^9/L				
Eos#		R	н	0.55	10^9/L				
Bas#		R		0.03	10^9/L				
IMG#		R		11.17	10^9/L	0 100 200 fL			
Neu%	&	R	L	29.6	%	0 100 200 fL	0 10 20 30 fL		
Lym%				****	%				
Mon%				****	%				
Eos%		R		0.5	%	WNB	DIFF		
Bas%		R		0.0	%				
IMG%		R		9.7	%	FS	FL		
RBC			L	1.41	10^12/L				
HGB			L	49	g/L				
НСТ			L	14.6	%				
MCV			Н	103.6	fL				
MCH			Н	34.8	pg				
MCHC				336	g/L	FL	SS		
RDW-CV			Н	18.6	%				
RDW-SD			Н	67.5	fL				
PLT			L	29	10^9/L	F	lags		
MPV				9.9	fL				
PDW			Н	18.0		-WBC Abn Scattergram -Blasts?	-Leukocytosis -Dimporphic Population		
РСТ			L	0.029	%	-Abn Lymph/blast?	-Dimporphic Population -Anemia		
P-LCC			L	8	10^9/L	-Immature Gran?	-Thrombocytopenia		
P-LCR				27.2	%	-Left Shift?			
NRBC#				0.347	10^9/L	-Neutrophilia			
NRBC%				0.30	/100WBC				

• WBC markedly increased; RBC and PLT decreased.

• In the WNB scattergram, nucleated red blood cells were observed; in the DIFF scattergram, the number of particles markedly increased, the Lym group extended towards the highly fluorescent region and covered the Mon group, so that the Lym and Mon groups were indistinguishable, indicating a large amount of myeloblasts, possibly lymphocytes; the Neu group also extended upward, indicating the presence of immature granulocytes.

C Peripheral blood morphology examination

	WI	BC	
	WBC	200	100%
-	Segmented neutrophil	46	23.0
	Band neutrophil	2	1.0
-	Lymphocyte	29	14.5
	Monocyte	14	7.0
	Eosinophil	1	0.5
	Basophil	1	0.5
	Metamyelocyte	5	2.5
	Myelocyte	5	2.5
	Myeloblast	95	47.5
	Reactive lymphocyte	2	1.0
	Non-WBC	93	%
	Nucleated RBC	1	0.5
	Smudge cell	52	26.0
	Sediment	40	
	PI	т	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	19*10^9/L	Manual
	RE	BC	
	Size	Degree	%
	Uneven erythrocyte size	s 3+	
	Macrocyte	2+	13.2
	Microcyte	0	3.4
	Color	Degree	%
	Hypochromic RBC	3+	27.2
	Polychromatic RBC	0	0.0
	Shape	Degree	%
	Poikilocytosis	3+	
	Schistocyte	3+	9.2
	Echinocyte	0	0.6
	Elliptocyte	0	0.9
	Ovalocyte	0	14.6
	Stomatocyte	0	0.3
	Leptocyte	0	0.0
	Dacryocyte	0	1.7
	Contents	Degree	%
	Contents Basophilic stippling	Degree 0	% 0.0



Manual microscopic result

Presence of myeloblasts (47.5%). The cell bodies were small in round or oval shapes. The nuclei were large, round or oval. The chromatins were fine and granule-like with clear boundaries, slightly coarser than those in the myelomonocytes. Nucleoli were visible in some cells. Extremely scant cytoplasm appeared pale blue, the boundaries were clear, and a very small amount of azurophilic granules were observed.

Other examinations

Item	Result
Flow cytometry	The abnormal cells accounted for 53.1% of the nucleated cells, expressing HLA-DR, CD2, CD7, CD38, CD99, CD117, and cCD3; partially expressing CD13, CD34, and TdT; and not expressing CD1a, CD4, CD5, and CD8. These cells were immature T-lymphocytes, where a group of HLA- DRpart+CD64part+MPOpart+CD13+CD33+CD38+ myeloid cells accounted for 0.6% of the nucleated cells; and the granulocytes showed abnormal differentiation in the CD15-CD11b, CD16-CD13, and CD11b-CD13 scattergrams. Conclusion: Suggested diagnosis: T-cell acute lymphoblastic leukemia (ETP-ALL possible), the involvement of myeloid cells is to be excluded.

Case analysis

T-cell acute lymphoblastic leukemia is a heterogeneous diseases characterized by the proliferation of malignant immature T lymphocytes, accounting for 10%–15% of childhood ALL. Although the current high-intensity chemotherapy has led to an induction response rate of more than 95% in children with T-ALL, it is prone to early relapse and difficult to response after relapse, so the long-term prognosis is still worse than that of pediatric precursor B-cell ALL. Early T-cell precursor-acute lymphoblastic leukemia (ETP-ALL) was first noted by Coustan-Smith et al. from St Jude Children's Hospital in 2009. The investigators screened out samples of patients with T-ALL with similar gene expression profiles to normal early T-precursor cells, and found that they had characteristic immunophenotypes. These patients had low response rates and short survival.

ETP-ALL was classified as an independent subtype by the WHO Classification of Tumors of Haematopoietic and Lymphoid Tissues in 2016. It has a unique immunophenotype and gene composition, and retains some characteristics of bone marrow stem cells at the immunophenotype and gene level. It is clinically highly aggressive and has a poor long-term prognosis. Once a patient is diagnosed, induction chemotherapy should be administered in clinical practice and the CBC should be monitored regularly.



CBC results on Aug. 21

- The CBC results on Aug. 21 showed significant WBC decrease. In the DIFF scattergram, the particle groups distributed normally, indicating no abnormal cells.
- The flow cytometry also indicated no leukemia residual cells with significantly abnormal immunophenotype.

Case analysis

Parameter	Flag	s	Result	Delta#	8-21	8-14	Unit
WBC		L	0.63	-0.890	1.52	11.19	10^9/L
Neu#	R	L	0.01	-0.690	0.70	8.68	10^9/L
Lym#	R	L	0.62	-0.190	0.81	2.05	10^9/L
Mon#	R	L	0.00	0.000	0.00	0.46	10^9/L
Eos#	R	L	0.00	-0.010	0.01	0.00	10^9/L
Bas#			0.00	0.000	0.00	0.00	10^9/L
IMG#	R		0.00	-0.010	0.01	0.99	10^9/L
Neu%	R	L	2.3	-44.30	46.6	77.6	%
Lym%	R	н	97.1	44.10	53.0	18.3	%
Mon%	R	L	0.0	0.00	0.0	4.1	%
Eos%	R		0.6	0.20	0.4	0.0	%
Bas%			0.0	0.00	0.0	0.0	%
IMG%	R		0.0	-0.60	0.6	8.8	%
RBC		L	1.73	-0.030	1.76	2.24	10^12/L
HGB		L	52	-3.0	55	70	g/L
НСТ		L	14.9	-0.80	15.7	21.4	%
MCV			86.3	-3.00	89.3	95.4	fL
MCH			29.9	-1.30	31.2	31.3	pg
MCHC			347	-3.0	350	328	g/L
RDW-CV			12.8	-1.90	14.7	18.3	%
RDW-SD			39.4	-7.10	46.5	61.3	fL
PLT		L	22	-2.0	24	13	10^9/L
MPV			9.5	0.40	9.1	10.3	fL
PDW			15.8	-0.30	16.1	17.3	
PCT		L	0.020	-0.0020	0.022	0.013	%
P-LCC		L	4	0.0	4	4	10^9/L
P-LCR			20.8	4.50	16.3	31.9	%
NRBC#			0.000	0.0000	0.000	0.000	10^9/L
NRBC%			0.00	0.000	0.00	0.00	/100WBC

The CBC results on Aug. 28 showed progressive WBC decrease, and the differentiation results showed a flag of "suspected". In the DIFF scattergram, the Neu group extended toward the highly fluorescent region, indicating presence of immature granulocytes. No other marked abnormalities were observed, but the analyzer raised an alarm of abnormal cells. A small amount of myeloblasts were observed in microscopic examination.

The flow cytometry showed that the cCD3+ cells accounted for approx. 1.99% of the nucleated cells; no leukemia residual cells with significantly abnormal immunophenotype were observed (leukemia residual cells < 10-4) (Note: when pro-T, pre-T, cortical T, and medullary T are observed, the cCD3 expression is regarded as +).



CBC results on Aug. 28



30 fL

SS

Case analysis

-	-							
Parameter	FI	ag	5	Result	Delta#	9-11	09-04	Unit
WBC	&		L	1.78	-0.610	2.39	0.74	10^9/L
Neu#				****		1.31	0.38	10^9/L
Lym#	&	R	L	0.53	-0.540	1.07	0.36	10^9/L
Mon#				****		0.01	0.00	10^9/L
Eos#		R		0.08	0.080	0.00	0.00	10^9/L
Bas#		R		0.01	0.010	0.00	0.00	10^9/L
IMG#				****		0.01	0.02	10^9/L
Neu%				****		54.3	51.5	%
Lym%	&	R		29.8	-15.10	44.9	48.2	%
Mon%				****		0.6	0.2	%
Eos%		R		4.6	4.50	0.1	0.1	%
Bas%		R		0.6	0.50	0.1	0.1	%
IMG%				****		0.2	2.9	%
RBC				3.55	1.360	2.19	2.08	10^12/L
HGB			L	101	41.0	60	56	g/L
НСТ			L	30.2	12.30	17.9	16.8	%
MCV				84.9	3.50	81.4	80.8	fL
MCH				28.3	1.10	27.2	27.0	pg
MCHC				333	-1.0	334	334	g/L
RDW-CV				15.1	-0.70	15.8	16.3	%
RDW-SD				46.1	-0.20	46.3	47.4	fL
PLT			L	54	-19.0	73	32	10^9/L
MPV				10.1	1.80	8.3	8.9	fL
PDW				16.7	0.90	15.8	15.7	
РСТ			L	0.054	-0.0070	0.061	0.029	%
P-LCC			L	17	5.0	12	7	10^9/L
P-LCR				31.1	15.00	16.1	21.3	%
NRBC#				0.085	0.0850	0.000	0.000	10^9/L
NRBC%				4.77	4.770	0.00	0.00	/100WBC

CBC results on Oct. 23



The results after 2 months still showed WBC decreased, but the DIFF scattergram was significantly different. The Mon and Neu groups markedly extended toward the highly fluorescent region and fused together, showing grayish-white color, and the analyzer raised the alarm of myeloblasts and abnormal lymphocytes; in the WNB scattergram, the presence of nucleated red blood cells was observed, indicating condition aggravated.

The flow cytometry showed that the CD45dimCD7st cells accounted for approx. 4.75% of the nucleated cells, also expressing CD117 and cCD3, and partially expressing CD99. Suggested diagnosis was abnormal immature T lymphocytes.

In this case, multiple CBC tests and flow cytometry examinations were conducted from diagnosis to treatment. The CBC results were consistent with the flow cytometry results. The progression of disease can be seen from the changes in the DIFF scattergrams below.



Case 11 Plasma cell leukemia (PCL)

Clinical information

A 51-year-old male patient was admitted to the hospital on Nov. 29, 2022 due to "palpitations for more than 3 years, definite diagnosis of multiple myeloma for 3 years, recurrence 3 months after transplantation".

G CBC results

Parameter	Flags		Result	Unit	RBC	PLT
WBC			7.66	10^9/L	NDC	L I
Neu#	R	L	0.67	10^9/L		
Lym#			****	10^9/L		
Mon#			****	10^9/L		
Eos#			0.02	10^9/L		
Bas#			0.01	10^9/L		
IMG#	R		0.02	10^9/L		
Neu%	R	L	8.8	%		
Lym%			****	%	0 100 200 fL	0 10 20 30
Mon%			****	%		
Eos%		L	0.3	%		
Bas%			0.1	%		
IMG%	R		0.3	%	WNB	DIFF
RBC		L	1.76	10^12/L	VVIND	DIFF
HGB		L	50	g/L		
НСТ		L	14.8	%	FS	FL
MCV			84.1	fL		
MCH			28.7	pg		
МСНС			341	g/L		
RDW-CV			15.4	%		
RDW-SD			47.7	fL		1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
PLT	&	L	3	10^9/L		100
MPV			****	fL	FL	Š
PDW			****			
РСТ			****	%		
P-LCC			****	10^9/L	RET	Flags
P-LCR			****	%		i lags
IPF			6.7	%		-WBC Abn Scattergram
RET#		L	0.0024	10^12/L	FS	-Blasts?
RET%		L	0.14	%		-Abn Lymph/blast?
IRF			6.7	%		-Atypical Lymph?
LFR			93.3	%		-Neutropenia
MFR			6.7	%		-Fragments? -Anemia
HFR			0.0	%	-Anemia -PLT Histogram Abn. -Thrombocytopenia	
RHE			28.6	Pg		
					FL	
NRBC#			0.000	10^9/L	FL	

- WBC normal, but the differentiation results were of poor reliability and were blocked; RBC decreased; severe anemia; extremely low PLT count.
- In the DIFF scattergram, the Mon particles were completely elevated to above the Lym particles and were tilting to the right, indicating the presence of a large amount of abnormal cells.

🖑 Peripheral blood morphology examination

		WBC								
	WBC	190	100%							
L	Segmented neutrophil	3	1.6							
	Band neutrophil	6	3.2							
	Lymphocyte	40	21.1							
L	Monocyte	5	2.6							
	Eosinophil	2	1.1							
1	Metamyelocyte	5	2.6							
1	Plasma cell	129	67.8							
	Non-WBC	45	%							
	Giant PLT	2								
	Smudge cell	40	21.1							
	Sediment	3								
PLT										
	PLT estimate	Estimated result	Estimation method							
	PLT concentration	8*10^9/L	Automated							
	PLT concentration	8*10^9/L	Manual							
		RBC								
	Size	Degree	%							
1	Uneven erythrocyte sizes	2+								
I.	Macrocyte	2+	11.2							
	Microcyte	0	2.2							
	Color	Degree	%							
1	Hypochromic RBC	3+	25.1							
	Polychromatic RBC	0	0.5							
	Shape	Degree	%							
	Poikilocytosis	0								
	Schistocyte	0	0.2							
	Echinocyte	0	0.0							
	Elliptocyte	0	0.2							
	Ovalocyte	0	5.6							
	Stomatocyte	0	0.0							
	Leptocyte	0	1.3							
	Dacryocyte	0	0.4							
	Contents	Degree	%							
	Basophilic stippling	0	0.0							



Manual microscopic result

The microscopic examination showed a large amount of plasma cells/ lymphoma cells. The cells were large, most of them were round shaped. The nuclei were large and round, not centrally-located. The chromatins aggregated to lumps. Nucleoli were observed in some cells. The cytoplasm was abundant and appeared in dark blue, foamy. Rouleux red blood cells were observed.

Other examinations

ltem		Result	Reference range
	Urobilinogen	1+	Normal
Urinalysis	Urine bilirubin	1+	Negative
	Urine protein	1+	Negative
	ALT	162.7	9.0-50.0U/L
	AST	192.0	15.0-40.0U/L
	ALP	165.4	45.0-125.0U/L
	γ-GGT	137.2	10.0-60.0U/L
	TP	98.0	65.0-85.0g/L
	GLO	71.7	20.0-40.0g/L
lepatic function	ALB	26.3	40.0-55.0g/L
	A/G	0.4	1.2-2.4
	TBil	80.8	0.0-26.0µmol/L
	inDBil	22.5	0.0-18.0µmol/L
	DBil	58.3	0.0-8.0µmol/L
	Prealbumin	0.10	0.17-0.42g/L
	GA	39.46	0.0-10.0mg/L
	Light chain κ	9450.00	598-1329mg/dL
erum light chain panel	Light chain λ	124.00	298-665 mg/dL
	κ/λ	76.21	1.35-2.65
	IgG	9150.00	751.00-1560.00 mg/dL
	IgA	54.20	82.00-453.00 mg/dL
mmunoglobulin	IgM	39.50	30.00-220.00 mg/dL
	Total IgE	19.90	0.00-165.00IU/mL



Case analysis

This patient was diagnosed with refractory/recurrent multiple myeloma, with secondary plasma cell leukemia, accompanied with infections, organ function abnormal (liver, lung, hear, Gl tract, etc.), severe anemia, and platelets decreased.
Case 12 Hairy cell leukemia (HCL)

Clinical information

The patient was a 65-year-old female.

G CBC results

Parameter	Flags		Result	Unit
WBC		н	14.65	10^9/L
Neu#	R		3.84	10^9/L
Lym#	R	н	10.58	10^9/L
Mon#	R	L	0.02	10^9/L
Eos#			0.20	10^9/L
Bas#			0.01	10^9/L
IMG#	R		0.02	10^9/L
Neu%	R	L	26.2	%
Lym%	R	Н	72.4	%
Mon%	R	L	0.1	%
Eos%			1.3	%
Bas%			0.0	%
IMG%	R		0.1	%
RBC			4.33	10^12/L
HGB			135	g/L
НСТ			39.6	%
MCV			91.3	fL
МСН			31.3	pg
MCHC			342	g/L
RDW-CV			13.5	%
RDW-SD			46.7	fL
PLT			145	10^9/L
MPV			11.3	fL
PDW			16.6	
РСТ			0.163	%
P-LCC			51	10^9/L
P-LCR			34.9	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC



• High WBC count, Lym increased; erythroids and platelets normal.

• In the DIFF scattergram, the number of Lym particles markedly increased in the highly fluorescent region, indicating the potential presence of abnormal lymphocytes.

	WBC							
	WBC	200	100%					
L	Segmented neutrophil	54	27.0					
	Band neutrophil	1	0.5					
L	Lymphocyte	36	18.0					
L	Monocyte	2	1.0					
	Eosinophil	1	0.5					
	Basophil	1	0.5					
1	Abnormal lymphocyte	105	52.5					
	Non-WBC	46	%					
	Large PLT	6						
	Smudge cell	38	19.0					
	Sediment	2						



Manual microscopic result

The microscopic examination showed abnormal lymphocytes (52.5%). The cells were large, about 1–2 times the volume of mature lymphocytes. The nuclei were oval, not centrally located. Some nuclei were round, elliptical, kidney-shaped, or horseshoe-like. The cytoplasm was abundant, appeared in pale blue to dusty blue, fluffy. Hair-like protrusions were observed at the periphery of cell membrane.

Case analysis

Based on the peripheral blood morphology examination, the patient was likely to have hairy cell leukemia (HCL). MICM related examinations are required.

For HCL, the median age of onset is 63 years old, few cases are observed in adolescence and more are observed in males. Typical HCL is easy to identify but atypical HCL is usually misdiagnosed as CLL. HCL often manifests clinically as pancytopenia or cytopenia of one lineage. The disease is often observed in the middle-aged and the elderly, with a chronic disease course, repeated infections, no enlargement of lymph nodes, but enlarged spleen. Monocytes are significantly decreased or even missing.

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Case 13 Chronic lymphocytic leukemia (CLL)

Clinical information

This patient was a 67-year-old male, with WBC increased of unknown reasons.

G CBC results

Parameter	Flags		Result	Unit
WBC		н	19.97	10^9/L
Neu#	R		6.59	10^9/L
Lym#	R	н	12.82	10^9/L
Mon#	R		0.32	10^9/L
Eos#			0.20	10^9/L
Bas#			0.04	10^9/L
IMG#	R		0.08	10^9/L
Neu%	R	L	33.0	%
Lym%	R	Н	64.2	%
Mon%	R	L	1.6	%
Eos%			1.0	%
Bas%			0.2	%
IMG%	R		0.4	%
RBC		L	3.00	10^12/L
HGB		L	82	g/L
НСТ		L	25.6	%
MCV			85.2	fL
MCH			27.5	pg
MCHC			320	g/L
RDW-CV		Н	19.8	%
RDW-SD		Н	60.6	fL
PLT	&	Н	591	10^9/L
MPV		L	6.3	fL
PDW			15.0	
PCT		Н	0.357	%
P-LCC			30	10^9/L
P-LCR		L	5.0	%
IPF		L	0.8	%
RET#			0.0753	10^12/L
RET%			2.51	%
IRF			19.5	%
LFR			80.5	%
MFR			15.7	%
HFR			3.8	%
RHE			25.7	pg
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC



- WBC increased, predominantly Lym; RBC decreased, moderate anemia; PLT increased.
- In the DIFF scattergram, some Lym particles extended to the highly fluorescent region, indicating the potential existence of abnormal cells or abnormal shape of lymphocytes.

		WBC				
	WBC	100	100%			
4	Segmented neutrophil	78	78.0	18		
	Band neutrophil	5	5.0			
_	Lymphocyte	16	16.0			-
	Eosinophil	1	1.0			12
	Non-WBC	172	%			1
	Nucleated RBC	1	1.0			
	Large PLT	1				
	PLT aggregation	1				
	Smudge cell	153	153.0	32		
	Sediment	16				
		PLT				-
	PLT estimate	Estimated result	Estimation method		1	
	PLT concentration	611*10^9/L	Automated			100
	PLT concentration	665*10^9/L	Manual			
		RBC				
	Size	Degree	%			
	Uneven erythrocyte sizes	0				
	Macrocyte	0	1.2			
	Microcyte	0	5.8	3		
	Color	Degree	%			945
	Hypochromic RBC	3+	60.0		8	
	Polychromatic RBC	0	0.0		ST.	1
	Shape	Degree	%		2.91	6. ···
	Poikilocytosis	0			-	.4
	Schistocyte	0	0.3			and all
	Echinocyte	0	2.6	-		
	Elliptocyte	0	0.5	42		-
	Ovalocyte	0	7.8			1
	Stomatocyte	0	0.0		34	1
	Leptocyte	0	0.7		-37	. 1
	Dacryocyte	0	0.2			12
	Contents	Degree	%		1	3
	Basophilic stippling	0	0.3			

Manual microscopic result

The microscopic examination showed that the lymphocytes had scant cytoplasm, with dense and regularshaped nuclei, the chromatins were aggregated, with a shape similar to the normal mature lymphocytes or with a slightly larger nucleus or cell body. A small number of lymphocytes had moderate cytoplasm, loose chromatins, and irregular nuclei, and the shape of these cells was similar to that of the "abnormal shape of lymphocytes". Smudge cells markedly increased.



ltem	Result
Flow cytometry immunofluorescence analysis	The flow cytometry showed that in the samples, there were approx. 63.4% of abnormal mature B lymphocytes, expressing CD5, CD19, CD23, and CD200; partially expressing CD22, CD20, and IgM; and not expressing CD10, CD34, CD38, CD79b, PMC7, sKappa, and sLambda. The flow cytometry results indicated CLL immunophenotyping (CLL score: 5).
Pathology examination report	(Posterior superior iliac spine): The HE and PAS staining showed uneven marrow hyperplasia. Most part showed extremely low hyperplasia (< 10%), a small part showed normal hyperplasia (approx. 40%). The lymphocytes increased in number, were scattered or focal, with small cell body, scant cytoplasm, round or slightly irregular nuclei, and coarse chromatins; myeloid and erythroid cells at various stages were visible, mainly myelocytes or at earlier stages. There was quite an amount of megakaryocytes, mainly segmented; reticulocyte staining was observed (MF-2, focal). The immunohistochemistry results are as follows: lymphocyte CD3 (-), CD5 (partial weak +), CD20 (+), PAX-5 (-), CD10 (-), CD23 (+), Cyclin D1 (-), SOX-11 (-), CD43 (-), CD38 (-), CD138 (-), MPO (-), TdT (-). Conclusion: Suggested diagnosis: CLL/small B-cell lymphocytic lymphoma. Further diagnosis is to be made in combination with clinical manifestations and other examinations.
Chromosome karyotyping	No clonal structure and number abnormalities were observed.

Case analysis

The patient's final diagnosis was CLL.

Case 14 Plasmodium malariae infection

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Clinical information

A 52-year-old male patient visited the hospital due to intermittent pyrexia (pyrexia onset every 72 hours) and pain for 1 week. This patient had a history of visiting Guinea a month before.

Physical examination: T 36.4 °C, P 86 bpm, R 20 times/min, BP 128/90 mmHg. The patient was conscious. He had no rash on the skin and mucous membranes, his superficial lymph nodes were not palpable or enlarged. The auscultation showed clear breath sounds in both lungs, with no dry or moist rales heard. HR: 86 bpm. The heart rhythm was normal and auscultation of each valve showed no pathological murmur. The abdomen was flat and soft, with no abdominal vein varices, gastrointestinal movement, or abdominal effusion. There was no tenderness or rebound tenderness. The liver and spleen were not palpable. The Murphy's sign was negative. The percussion test showed tympanic sound. There was no percussion pain in the liver region or kidney region. The shifting dullness was negative. The bowel sounds were normal with no gas and water sounds. Physiological reflexes were all normal and pathological reflexes were all negative.

S CBC results

Parameter	Flags	Result	Unit
WBC		4.95	10^9/L
Neu#		3.65	10^9/L
Lym#	L	0.67	10^9/L
Mon#		0.37	10^9/L
Eos#		0.24	10^9/L
Bas#		0.02	10^9/L
IMG#		0.01	10^9/L
Neu%	Н	73.9	%
Lym%	L	13.5	%
Mon%		7.4	%
Eos%		4.7	%
Bas%		0.5	%
IMG%		0.3	%
RBC		4.26	10^12/L
HGB		139	g/L
HCT		40.6	%
MCV		95.4	fL
MCH		32.7	pg
MCHC		343	g/L
RDW-CV		11.9	%
RDW-SD		42.0	fL
*InR#		0.23	0^9/L
*InR‰		0.05	%0
PLT		137	10^9/L
MPV		9.9	fL
PDW		16.0	
PCT		0.136	%
P-LCC		33	10^9/L
P-LCR		24.2	%
NRBC#		0.000	10^9/L
NRBC%		0.00	/100WBC



- The blood cells were generally normal, high Neu%.
- In the DIFF scattergram, there was a small amount of green particles above the Mon group, indicating potential existence of abnormal cells or abnormal shape of lymphocytes. The Eos group was more inclined to the lower left distribution than the normal model, and was generally flat, located in the lowly fluorescent region. Microscopic examination is required.

		WBC	
	WBC	100	100%
	Segmented neutrophil		70.0
1	Band neutrophil	11	11.0
L	Lymphocyte	11	11.0
	Monocyte	5	5.0
н	Basophil	2	2.0
1	Myelocyte	1	1.0
	Non-WBC	34	%
1	Nucleated RBC	1	1.0
	Large PLT	11	
	Smudge cell	13	13.0
	Sediment	9	
		PLT	
	PLT estimate		Estimation method
	PLT concentration	183*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	0	
	Macrocyte	0	0.1
1	Microcyte	2+	11.0
	Color	Degree	%
	Hypochromic RBC	0	0.2
	Polychromatic RBC	0	0.6
	Shape	Degree	%
1	Poikilocytosis	2+	
	Schistocyte	0	0.8
1	Echinocyte	2+	43.2
	Elliptocyte	0	0.3
	Ovalocyte	0	4.4
	Stomatocyte	0	0.0
	Leptocyte	0	0.0
	Dacryocyte	0	0.2
	Contents	Degree	%
	Basophilic stippling	0	0.1

Manual microscopic result

The microscopic examination showed red blood cells infected by *Plasmodium malariae*, mainly schizonts and trophozoites. Schizont almost occupied all red blood cells. The cytoplasm was dark blue and the merozoites were arranged in ring form. The malarial pigment was gathered at the center. The ring of trophozoites was thick, about 1/3 the diameter of the red blood cell. The parasite body had 1–2 red nuclei, dark blue cytoplasm, and was ring-like.

Other examinations

ltem	Result
Plasmodium detection at CDC	Plasmodium malariae

🖹 Case analysis

Plasmodium malariae parasites in small red blood cells. There are usually 6–12 merozoites arranged in a chrysanthemumlike pattern, with central aggregation; The gametocytes are round with a dark red or pale red nucleus on one side or in the center, and dark brown malarial pigments scatter evenly. Data have shown that once infected, *Plasmodium malariae* may be carried lifelong. Its incubation period in humans may last from several years to even decades, and it can survive 20 to 30 years in human bone marrow before causing malaria episodes. Due to the long development duration in red blood cells and the small number of mature merozoites, the detection of *Plasmodium malariae* is easily missed in the peripheral blood smear examination at the early stage of onset.

Case 15 Plasmodium falciparum infection

Clinical information

This patient was a 48-year-old male.

G CBC results

Parameter	Flags	Result	Unit
WBC	Н	17.17	10^9/L
Neu#	RΗ	12.20	10^9/L
Lym#	RΗ	4.08	10^9/L
Mon#	R	0.56	10^9/L
Eos#	R	0.05	10^9/L
Bas#	RΗ	0.28	10^9/L
IMG#	R	1.02	10^9/L
Neu%	R	70.9	%
Lym%	R	23.8	%
Mon%	R	3.3	%
Eos%	R L	0.3	%
Bas%	RΗ	1.7	%
IMG%	R	5.9	%
RBC	L	2.14	10^12/L
HGB	L	62	g/L
НСТ	L	18.1	%
MCV		84.6	fL
MCH		28.9	pg
MCHC		342	g/L
RDW-CV		14.8	%
RDW-SD		43.4	fL
*InR#		0.04	0^9/L
*InR‰		0.02	%0
PLT	& L	61	10^9/L
MPV		****	fL
PDW		****	
PCT		****	%
P-LCC		****	10^9/L
P-LCR		****	%
RET#	Н	0.2499	10^12/L
RET%	Н	11.68	%
IRF		2.4	%
LFR		97.6	%
MFR		2.3	%
HFR		0.1	%



- WBC increased; RBC and PLT both decreased.
- In the DIFF scattergram, the particle groups all diffused toward the highly fluorescent region; in the RET scattergram, there was a group of highlighted particles in the LFR region.

		WBC	
	WBC	200	100%
L	Segmented neutrophil	91	45.5
!	Band neutrophil	38	19.0
L	Lymphocyte	17	8.5
	Monocyte	11	5.5
	Basophil	1	0.5
!	Metamyelocyte	26	13.0
	Promyelocyte	1	0.5
!	Myeloblast	3	1.5
	Reactive lymphocyte	12	6.0
	Non-WBC	161	%
	Nucleated RBC	11	5.5
	Giant PLT	1	
	Large PLT	30	
	PLT aggregation	2	
	Smudge cell	97	48.5
	Sediment	11	
	Plasmodium - trophozoite	8	
	Plasmodium - schizont	1	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	139*10^9/L	Automated
	PLT concentration	100*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte sizes	2+	
	Macrocyte	0	1.2
	Microcyte	2+	13.0
	Color	Degree	%
	Hypochromic RBC	0	1.6
	Polychromatic RBC	0	0.3
	Shape	Degree	%
!	Poikilocytosis	1+	
!	Schistocyte	1+	1.7
	Echinocyte	0	4.2
	Elliptocyte	0	1.7
	Ovalocyte	0	16.7
	Stomatocyte	0	0.0
	Leptocyte	0	0.2
	Dacryocyte	0	0.4
	Contents	Degree	%
	Basophilic stippling	0	0.3
	Basophilic stippling		

Manual microscopic result

The microscopic examination showed red blood cells infected by *Plasmodium falciparum*, mainly trophozoites. The ring was thin, taking up about 1/5 the red blood cell. There were 1–2 nuclei. Two or more protozoons were inside each red blood cell. The parasite body was usually located at the edge of the red blood cell. A small amount of schizonts were visible, irregularly arranged, and the malarial pigments were gathered. Neutrophils ingesting the *Plasmodium* were visible.

Case analysis

In recent years, due to the increasing mobility of the population, malaria is mainly caused by *Plasmodium falciparum* and *Plasmodium vivax*, while cases of *Plasmodium malariae* are less common. Once a patient is infected, the *Plasmodium* in the red blood cells lyses and proliferates, damaging the red blood cells and leading to anemia. The schizonts released and the other metabolic products are engulfed by phagocytic cells, resulting in abnormal monocyte and neutrophil contents. This is shown in the scattergram as the Neu and Mon groups extending toward the highly fluorescent and high lateral scattered light regions. Sometimes, the Neu particles in the high lateral scattered light region are counted as Eos by the analyzer. For real eosinophils, their eosinophilic granules specifically bind to the reagents in the analyzer so that the lateral scattered light is markedly enhanced. This can be used for the differentiation from the neutrophils engulfing the *Plasmodium*.

In addition, the infected red blood cells carry the nucleic materials of the *Plasmodium*, so the infected red blood cells showed certain fluorescent signals in the RET scattergram, leading to false RET increase.

Case 16 Essential thrombocythemia (ET)

Clinical information

This patient was a 48-year-old male.

G CBC results

Parameter	Flag	gs		Result	Unit
WBC			н	59.17	10^9/L
Neu#		R	Н	54.22	10^9/L
Lym#				1.68	10^9/L
Mon#			н	1.41	10^9/L
Eos#		R	н	1.62	10^9/L
Bas#		R	н	0.24	10^9/L
IMG#				1.75	10^9/L
Neu%		R	Н	91.7	%
Lym%			L	2.8	%
Mon%			L	2.4	%
Eos%		R		2.7	%
Bas%		R		0.4	%
IMG%				2.9	%
RBC		R	L	3.72	10^12/L
HGB			L	75	g/L
НСТ		R	L	25.9	%
MCV		R	L	69.6	fL
МСН		R	L	20.1	pg
MCHC		R	L	288	g/L
RDW-CV		R	н	19.5	%
RDW-SD		R		49.5	fL
PLT	&	@	н	5126	10^9/L
MPV		R		8.1	fL
PDW		R		16.1	
PCT		R	н	3.129	%
P-LCC		R	Н	832	10^9/L
P-LCR		R		16.2	%
IPF		R		3.8	%
RET#		R	Н	0.3437	10^12/L
RET%			Н	9.24	%
IRF				23.7	%
LFR			L	76.3	%
MFR				13.7	%
HFR			Н	10.0	%
RHE			L	19.1	pg
NRBC#				0.000	10^9/L
NRBC%				0.00	/100WBC

RBC



DIFF

SS

FL

WNB





FL

FS

Flags -Immature Gran? -Left Shift? -Basophilia -Eosinophilia -Neutrophilia -Neutrophilia -Leukocytosis -RBC Agglutination -Fragments -Hypochromia -Iron Deficiency?

- -Microcytosis
- -Reticulocytosis
- -PLT Clump

PLT extremely increased.Histograms and scattergrams were basically normal.

• WBC increased, predominantly Neu; RBC decreased, microcytic hypochromic anemia;

		WBC				RBC	
	WBC	100	100%		Size	Degree	%
н	Segmented neutrophil	86	86.0	I.	Uneven erythrocyte sizes	3+	
	Band neutrophil	4	4.0		Macrocyte	0	2.0
L	Lymphocyte	1	1.0	I.	Microcyte	2+	17.1
L	Monocyte	1	1.0		Color	Degree	%
	Eosinophil	4	4.0	Hypochromic RBC	3+	42.6	
н	Basophil	3	3.0 ! 1.0		Polychromatic RBC	2+	2.0
	Metamyelocyte	1			Shape	Degree	%
					Poikilocytosis	0	
	Non-WBC	10	%		Schistocyte	0	1.1
	Large PLT	4			Echinocyte	0	0.0
	PLT aggregation	1	4.0		Elliptocyte	0	0.5
	Smudge cell	4			Ovalocyte	0	7.7
	Sediment	1			Stomatocyte	0	2.3
	PLT					0	1.4
	PLT estimate	Estimated result	Estimation method		Dacryocyte	0	0.4
	PLT concentration	4347*10^9/L	Automated		Contents	Degree	%
	PLT concentration	4953*10^9/L	Manual		Basophilic stippling	0	0.0

Manual microscopic result

The microscopic examination showed WBCs increased, mainly mature neutrophils. The platelets were abnormally increased, scattered, and distributed in small clusters. The cell sizes were uneven, microplatelets, large platelets, and giant platelets were visible. A small amount of platelets were lack of granules.



Other examinations

ltem	Result
Bone marrow morphology	Active bone marrow hyperplasia; active myeloid hyperplasia, the ratio of cells at each stage was normal; active erythroid hyperplasia; a total of 51 megakaryocytes were observed; the platelets were scattered and visible in clusters; no other pathological cells or parasites were observed. Suggested diagnosis: thrombocytosis.
Gene detection	JAK2 14exon: JAK2 V617F 40% mutation.

Case analysis

Essential thrombocythemia (ET) adds to the risk of embolism and hemorrhage. About 1/3 of the ET patients have gene mutations, where JAK2 V617F mutations account for 60%. In 2008, WHO listed JAK2 V617F gene mutation as one of the primary diagnostic indicator for myeloproliferative neoplasms (MPN). In the WHO MPN classification guideline of 2016, JAK2 analysis becomes the main evaluation criteria for all MPN diagnoses.

Case 17 Hypogranular platelet

Clinical information

CBC results

Flags

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Parameter

A 59-year-old male patient visited the hospital due to chronic colitis, gastroenteritis, etc. Subsequent pathological diagnosis showed benign colon neoplasm. The initial CBC results showed: PLT-I: 220*10^9/L, PLT-F: 29*10^9/L. The difference was significant, the scattergrams are shown as follows, so retesting was conducted using another analyzer.

Unit

Result

RBC **PLT** 100 fL 200 20 0 0 10 30 fL **WNB** DIFF FL SS Flags - PLT Clump?

• The blood cells were generally normal.

0.00

/100WBC

• The PLT histogram had a zigzag tail, the alarm of platelet aggregation was raised. Since the PLT results from different tests varied, microscopic examination was required for retest.

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NRBC%

WBC			6.64	10^9/L
Neu#			4.14	10^9/L
Lym#			1.83	10^9/L
Mon#			0.49	10^9/L
Eos#			0.15	10^9/L
Bas#			0.03	10^9/L
IMG#			0.02	10^9/L
Neu%			62.5	%
Lym%			27.5	%
Mon%			7.3	%
Eos%			2.3	%
Bas%			0.4	%
IMG%			0.3	%
RBC			4.75	10^12/L
HGB			155	g/L
НСТ			47.2	%
MCV			99.3	fL
MCH			32.6	pg
MCHC			328	g/L
RDW-CV			12.4	%
RDW-SD			44.3	fL
PLT	R		219	10^9/L
MPV	R	н	13.6	fL
PDW	R		16.3	
РСТ	R	Н	0.297	%
P-LCC	R	н	114	10^9/L
P-LCC P-LCR	R R	H H	114 52.1	10^9/L %





		WBC					
	WBC	200	100%				
	Segmented neutrophil	131	65.5				
	Band neutrophil	5	2.5				
L	Lymphocyte	23	11.5				
	Monocyte	12	6.0				
н	Eosinophil	23	11.5				
н	Basophil	6	3.0				
	Non-WBC	450	%				
	Giant PLT	6					
	Large PLT	6					
	Smudge cell	388	194.0				
	Sediment	50					
		PLT					
	PLT estimate	Estimated result	Estimation method				
	PLT concentration	214*10^9/L	Automated				
	PLT concentration	209*10^9/L	Manual				
	RBC						
	Size	Degree	%				
	Uneven erythrocyte sizes	5 0					
	Macrocyte	0	10.7				
	Microcyte	0	0.8				
	Color	Degree	%				
	Hypochromic RBC	0	0.1				
	Polychromatic RBC	0	0.1				
	Shape	Degree	%				
	Poikilocytosis	2+					
	Schistocyte	2+	1.7				
	Echinocyte	0	0.2				
	Elliptocyte	0	1.4				
	Ovalocyte	2+	15.0				
	Stomatocyte	0	0.0				
	Leptocyte	0	0.0				
	Dacryocyte	0	1.5				
	Contents	Degree	%				
	Basophilic stippling	0	0.0				



Manual microscopic result

The PLT estimate was consistent with the second CBC result. A large amount of grayish-white platelets scattered.

Case analysis

This is a case of fluorescence-based false PLT count decreased caused by platelet granule deficiency. Platelet granule deficiency is observed in congenital disorders, such as gray platelet syndrome. The platelets and megakaryocytes have abnormal shapes and the cellular granules markedly decrease. Since the PLT cellular granules carry a large amount of nucleic materials, the lack of these granules leads to the weakening of PLT fluorescence signal, which is a possible cause of PLT false decrease.

Case 18 EDTA-induced pseudothrombocytopenia (EDTA-PTCP)

Clinical information

This patient was a 66-year-old male. The gastroscopic examination on Jul. 3, 2023 showed progressive gastric cancer and chronic atrophic gastritis. The patient was then admitted to the hospital due to "gastric malignancy". Since onset, the patient was in good general conditions, with normal urination and defecation, but weight loss.

Sec results

Parameter	Flags		Result	Unit
WBC	R		8.14	10^9/L
Neu#	R		5.91	10^9/L
Lym#			1.02	10^9/L
Mon#			0.45	10^9/L
Eos#	R	Н	0.69	10^9/L
Bas#			0.07	10^9/L
IMG#			0.05	10^9/L
Neu%	R	Н	72.7	%
Lym%		L	12.5	%
Mon%			5.5	%
Eos%	R	н	8.4	%
Bas%			0.9	%
IMG%			0.6	%
RBC			4.86	10^12/L
HGB			153	g/L
НСТ			46.7	%
MCV			96.1	fL
MCH			31.4	pg
MCHC			327	g/L
RDW-CV			13.5	%
RDW-SD			46.0	fL
PLT-O	R		175	10^9/L
PLT-I	R	L	38	
MPV	R		9.8	fL
PDW	R		16.6	
PCT	R	L	0.037	%
P-LCC	R		49	10^9/L
P-LCR	R		28.2	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC



- The blood cells were generally normal. The difference between PLT-O and PLT-I was significant, possibly due to PLT aggregation.
- The PLT histogram had a zigzag tail, the alarm of platelet aggregation was raised. In the WNB scattergram, the blood cell particle group on the left bottom corner extended toward the right upper direction and even surpassed the WBC group, and the analyzer raised the alarm of abnormal scattergram. In the DIFF scattergram, the Neu group extended toward the high lateral scattered light region and invaded the Eos region, so some Neu particles were counted as Eos.

		WBC	
	WBC	201	100%
L	Segmented neutrophil	67	33.3
	Lymphocyte	68	33.8
Н	Monocyte	40	19.9
н	Eosinophil	20	10.0
н	Basophil	5	2.5
	Metamyelocyte	1	0.5
	Non-WBC	252	%
	Giant PLT	1	
	Large PLT	14	
	PLT aggregation	61	
	Smudge cell	63	31.3
	Sediment	113	
		PLT	
	PLT estimate	Estimated result	Estimation method
	PLT concentration	214*10^9/L	Automated
	PLT concentration	209*10^9/L	Manual
		RBC	
	Size	Degree	%
	Uneven erythrocyte size		
	Macrocyte	0	10.7
	Microcyte	0	0.8
	Color	Degree	%
	Hypochromic RBC	0	0.1
	Polychromatic RBC	0	0.1
	Shape	Degree	%
1	Poikilocytosis	2+	
1	Schistocyte	2+	1.7
	Echinocyte	0	0.2
	Elliptocyte	0	1.4
1	Ovalocyte	2+	15.0
	Stomatocyte	0	0.0
	Leptocyte	0	0.0
	Dacryocyte	0	1.5
	Contents	Degree	%
	Basophilic stippling	0	0.0

Case analysis

Platelet satellitism is commonly reported in samples from various cancer patients and are thought to be related to platelet activation induced by ethylenediaminetetraacetic acid anticoagulants (EDTA). This EDTA-induced pseudo-thrombocytopenia is known as EDTA-PTCP in clinical practice.

PTCP can be observed in patients with autoimmune disorders, inflammations, tumors, or pregnancy, and even in the healthy population. This indicated that these autoantibodies are not related to the onset of diseases.

To investigate this case for the relationship between PLT satellitism and the blood sampling time/ anticoagulants, blood samples were collected in EDTA-tubes and citrate tubes, respectively, and tested at 5 min, 30 min, 1 h, 2 h, and 3 h after blood sampling. The results are as follows:



Case analysis

EDTA sample							
Time after blood sampling	5min	30min	1h	2h	3h		
PLT-O	240	220	197	162	99		
PLT-I	224	157	83	47	27		
WBC-O	5.26	5.34	5.18	5.62	4.42		
WBC-D	6.12	6.05	6.95	7.59	8.29		
WBC-N	6.01	6.17	6.14	7.70	8.13		
PLT histogram	0 10 20 30 ft	0 10 20 30 m	0 10 20 30 TL		0 10 20 30 ft		
Neu	8		3		S.		
Lym		0					
Mon	0						

The histogram within 5 min after blood sampling showed that there was no pattern of platelet aggregation. However, with the prolongation of the test time, the histogram at 30 min after blood sampling already showed an elevating trend of the tail, together with zigzag patterns. The histogram at 3 h after blood sampling clearly indicated PLT aggregation.

The CDR test results showed that there was almost no difference in PLT-I and PLT-O results within 5 min after blood sampling. With the prolongation of the test time, the PLT-O results were much greater than PLT-I, indicating that the testing of EDTA-dependent platelet aggregation samples could be improved by RET channel; however, the PLT-O results also decreased with the increase of time, indicating that the ability of RET channel to depolymerize platelets continuously decreased with the increase of time.

The WBC results also increased over time, possibly because the PLT aggregation group had scattered light and fluorescence signals similar to WBC and were hence counted as WBCs.

According to the film reading results, satellitism and phagocytosis were also more pronounced over time.



Case analysis

Citrate sample								
Time after blood sampling	5min	30min	1h	2h	3h			
PLT-O	225	185	170	140	116			
PLT-I	189	174	129	115	93			
WBC-O	5.54	4.22	4.70	4.02	3.94			
WBC-D	5.60	5.35	5.25	5.92	6.30			
WBC-N	5.62	5.36	5.26	5.42	5.41			
PLT histogram	0 10 20 30 TL	0 10 20 30 TL	0 10 20 30 fL	0 10 20 30 fL	0 10 20 30 n.			
Neu	63	3	ġ					
Lym								
Mon	Ċ							

As shown in the histogram, PLT results, and cytogram, the PLT aggregation trend in the citrate tubes was significantly weaker than that in the EDTA tubes.

In this case, severe EDTA-dependent PLT satellitism was observed along with chronic gastric cancer. It is worth noting that PLT satellitism usually occurs around neutrophils, but it is rare to see satellitism and phagocytosis around lymphocytes and monocytes, such as in this case. A possible reason for PLT satellitism around lymphocytes may be platelet adsorption caused by the lymphocyte surface protein molecular abnormality.

PLT satellitism generally occurs under the following conditions.

1. EDTA-K2 can induce immune mediation in blood. Antiplatelet antibodies are then produced, leading to PLT satellitism and low PLT count.

2. The complement mediated cell lysis (C3) is activated and the major product, C3b, attaches onto the surface of the platelets and binds with the C3b receptors on the neutrophils, so platelets are engulfed, leading to low PLT count.

3. WBCs and PLTs interact through adhesion protein molecules.

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