

HemaCase Clinical Case Booklet

By using Mindray Fully Automated Cellular Analysis Lines

Volume 1



Preface

We are delighted to present you with this clinical case booklet on hematology. At Mindray, our mission has always been to bring cutting-edge technology within reach, ensuring that high-quality healthcare is available to all. As part of this commitment, we have dedicated ourselves to enhancing the accuracy and efficiency of hematology tests in clinical laboratories.

Through the pages of this booklet, we aim to provide you with real clinical cases that vividly illustrate the seamless integration of information from blood analyzers, morphological analysis, and clinical diagnostic data. With the assistance of our state-of-the-art Automated Digital Cell Morphology Analyzer MC-80, these cases showcase how early disease identification and diagnosis can be achieved.

Our ultimate aspiration is for these cases to empower laboratory personnel, like yourself, to gain invaluable insights in their daily work. By immersing yourself in these scenarios, we hope you will not only expand your knowledge but also elevate the quality of patient care you provide.

Contents

List of Abbreviations 04
Scattergrams of 5-part differential hematology analyzer
Case 01 05 Acute myeloblastic leukemia without maturation (M1)
Case 02 11 Acute promelocytic leukemia (APL)
Case 03 15 Acute myeloblastic leukemia with maturation (M2)
Case 04 17 Acute myelomonocytic leukemia (AMML)
Case 05 19 Thrombotic thrombocytopenic purpura (TTP)
Case 06 21 Chronic myelomonocytic leukemia (CMML)
Case 07 23 MDS with single lineage dysplasia (MDS-SLD)
Case 08
Case 09 27 Multiple myeloma (MM)
Case 10 29 B-cell lymphoblastic leukemia/lymphoma (B-ALL)
Case 11
Case 12
Case 13
Case 14
Case 15
Case 16 42 Green neutrophilic inclusion
References 45

List of Abbreviations

WBC \rightarrow White blood cell	Eos \rightarrow Eosinophil	RET# \rightarrow Reticulocyte count
Neu \rightarrow Neutrophil	Eos# \rightarrow Eosinophil count	RET% \rightarrow Reticulocyte percentage
Neu# \rightarrow Neutrophil count	Eos $\% \rightarrow$ Eosinophil percentage	$\begin{array}{rcl} RHE & \to & \underset{expression}{Reticulocyte hemoglobin} \end{array}$
Neu% \rightarrow Neutrophil percentage	Bas \rightarrow Basophil	$IRF \rightarrow Immature\ reticulocyte\ fraction$
Lym \rightarrow Lymphocyte	Bas# \rightarrow Basophil count	$PLT \rightarrow Platelet$
Lym# \rightarrow Lymphocyte count	Bas% \rightarrow Basophil percentage	PLT-I \rightarrow Platelet counting with impedance method
Lym% \rightarrow Lymphocyte percentage	$RBC \longrightarrow Red blood cell$	$\label{eq:PLT-O} \begin{array}{l} \text{Platelet counting with optical} \\ \text{method} \end{array}$
Mon \rightarrow Monocyte	HGB \rightarrow Hemoglobin	IPF o Immature platelet fraction
Mon# \rightarrow Monocyte count	$MCV \ \longrightarrow \ Mean\ corpuscular\ volume$	
Mon% $ ightarrow$ Monocyte percentage	NRBC \rightarrow Nucleated RBCs	

Scattergrams of 5-part differential hematology analyzer



Case 01 Acute myeloblastic leukemia without maturation (M1)

Clinical information

One year ago, the patient presented with a history of recurrent dry coughing for over six months, with worsening symptoms in the last month. A chest CT scan revealed a nodule in the right middle lung, and further testing below confirmed a diagnosis of AML-M1.

ltem	Parameter	Result
	WBC	195.4*10^9/L
CBC	HGB	92g/L
	PLT	11*10^9/L
Bone marrow morphology	Bone marrow morphology	Highly active bone marrow with 95% of myeloblasts, indicating acute myeloid leukemia of the AML-M1 subtype.
Immunophenotyping	Immunophenotyping	Immunophenotyping suggested AML
Chromosome	Chromosome	46, XX[20]
	FLT3-ITD mutation	AR=0.271
Bone marrow NGS	CEBPA p.Thr310dup	45.6%
	CEBPA p.Gln83fs	47.1%
	WT1 p.Gly356Ter	44.7%

After more than 4 months of chemotherapy, the patient improved and was discharged. However, the patient relapsed 2 months later and underwent 6 months of continued chemotherapy. Bone mzarrow morphology suggested that the disease was not resolved, so the patient was readmitted for treatment on Jun. 2.

S CBC results

Parameter	AI	arn	n	Result	Unit
WBC	&		L	0.85	10^9/L
Neu#	&	R	L	0.26	10^9/L
Lym#	&	R	L	0.57	10^9/L
Mon#		R	L	0.02	10^9/L
Eos#			L	0.00	10^9/L
Bas#				0.00	10^9/L
IMG#		R		0.00	10^9/L
Neu%	&	R	L	30.6	%
Lym%	&	R	н	66.0	%
Mon%		R	L	2.5	%
Eos%				0.6	%
Bas%				0.3	%
IMG%		R		0.5	%
RBC			L	2.01	10^12/L
HGB			L	75	g/L
HCT			L	22.8	%
MCV			н	113.3	fL
MCH			н	37.0	pg
MCHC				327	g/L
RDW-CV			н	22.3	%
RDW-SD			н	90.3	fL
PLT		R	L	4	10^9/L
MPV				****	fL
PDW				****	
PCT				****	%
P-LCC				****	10^9/L
P-LCR				****	%
NRBC#				0.130	10^9/L
NRBC%				15.28	/100WBC

CBC results on Jun. 1



- WBC decreased with inverted Neu% and Lym%; decreased RBC and HGB; extremely low PLT count.
- NRBCs were visible in the WNB channel; due to the decreased WBC, the DIFF 3X channel was used to increase the particle count by 3 times, thereby improving the repeatability of classification results and the ability to detect abnormal cells. The scattergram showed that Mon particles extended upward into the abnormal lymphocytes/blasts region. The instrument also gave an alarm.

Peripheral blood morphology examination

	Whit	e blood cells	
	White blood cells	100	100%
L	Segmented neutrophils	18	18.0
	Band neutrophils	4	4.0
Н	Lymphocytes	70	70.0
	Eosinophils	1	1.0
!	Blasts	7	7.0
	Non-white blood cells	45	%
1	Nucleated RBCs	12	12.0
	Large platelets	1	
	Smudge cells	9	9.0
		Platelet	Ectimation
	PLT estimate	Estimated result	method
	Platelet concentration	6*10^9/L	Automated
	Platelet concentration	9*10^9/L	Manual
	Re	d blood cells	
	Size	Degree	%
1	Anisocytosis	3+	
	Macrocytes	0	6.7
	Microcytes	0	5.3
	Color	Degree	%
1	Hypochromic cells	2+	15.0
	Polychromasia	0	0.8
	Shape	Degree	%
!	Poikilocytosis	3+	0.6
!	Schistocytes	1+	0.6
	Elliptocytes	0	0.0
1	Ovalocytes	3+	21.9
	Stomatocytes	0	1.6
	Target cells	0	0.2
	Teardrop cells	0	2.2

Pre-classification by the reader on Jun. 1



Results from manual re-classification

Segmented neutrophils accounted for 16%, band neutrophils accounted for 3%, metamyelocytes accounted for 3%, lymphocytes accounted for 70%, and myeloblasts accounted for 7%. The blasts had small round cell bodies and large, round or oval nuclei with fine chromatins. Some blasts had visible nucleoli and scant cytoplasm that appeared pale blue. The PLT estimate was consistent with the CBC result.

Case analysis

- At the initial diagnosis, the patient had a bone marrow blast percentage of 95% and was diagnosed with Acute myeloblastic leukemia (AML-M1). There were no chromosomal abnormalities, but CEBPA mutation was present. Generally, AML patients with biallelic CEBPA mutation have a good prognosis, but in this case, the patient also had an FLT3-ITD (FMS-like tyrosine kinase 3-internal tandem duplication) mutation, which is a strong indicator of poor prognosis as it leads to increased tyrosine kinase activity and the activation of various tumors, especially AML. Therefore, the patient relapsed after only two months following improvement with treatment, and subsequent treatment results were not ideal.
- The DIFF scattergram of the CBC showed that the upper part of the Lym cluster was pointed and slightly extended to the
 upper right; although the Mon particles were scarce, they were significantly elevated, some of which entered the abnormal
 lymphocytes/myeloblast region, and was shifted to the right overall, indicating the possible presence of abnormal cells. This
 corresponds to the results of the peripheral blood smear reading. Blasts were observed in 7% of peripheral blood cells.
 Compared with typical myeloblasts (as shown in the figure below), blasts have scant cytoplasm and less fine and loose
 chromatins, which to some extent affected the binding of nucleic acid fluorescent dyes, resulting in relatively weak FL signals of
 the corresponding particles in the DIFF scattergram.

E Case analysis

Reference figures

of blasts



After the patient was admitted to the hospital, the chemotherapy regimen was changed. We tracked the CBC and peripheral blood morphology results as follows:

The CBC results from Jul. 6, Jul. 13, and Aug. 3 are shown from right to left

Parameter	A	lar	m	Result	Delta#	07-13	07-6	Unit
WBC	&		L	2.35	0.040	2.31	1.96	10^9/L
Neu#	&	R	L	0.62	-0.200	0.82	0.55	10^9/L
Lym#	&	R		1.70	0.310	1.39	1.35	10^9/L
Mon#		R	L	0.02	-0.010	0.03	0.03	10^9/L
Eos#		R	L	0.01	-0.060	0.07	0.03	10^9/L
Bas#		R		0.00	0.000	0.00	0.00	10^9/L
IMG#		R		0.08	-0.050	0.13	0.08	10^9/L
Neu%	&	R	L	26.2	-9.40	35.6	28.2	%
Lym%	&	R	Н	72.2	12.40	59.8	68.8	%
Mon%		R	L	1.0	-0.40	1.4	1.3	%
Eos%		R		0.5	-2.50	3.0	1.6	%
Bas%		R		0.1	-0.10	0.2	0.1	%
IMG%		R		3.2	-2.50	5.7	4.2	%
RBC			L	2.83	0.350	2.48	2.05	10^12/L
HGB			L	96	7.0	89	76	g/L
НСТ			L	30.9	2.40	28.5	24.1	%
MCV			н	109.4	-5.50	114.9	117.4	fL
MCH			н	34.1	-1.70	35.8	36.9	pg
MCHC			L	312	1.0	311	315	g/L
RDW-CV				14.9	-2.70	17.6	19.1	%
RDW-SD			Н	63.5	-15.00	78.5	81.2	fL
PLT	&		L	13	4.0	9	18	10^9/L
MPV		R		10.6	-1.00	11.6	15.2	fL
PDW		R		16.3	0.10	16.2	17.8	
PCT		R	L	0.008	-0.0010	0.009	0.027	%
P-LCC		R	L	6	2.0	4	11	10^9/L
P-LCR		R		44.8	-2.00	46.8	61.3	%
IPF			Н	26.9	-1.10	28.0		%
RET#				0.1974	-0.1371	0.3345		10^12/L
RET%			Н	6.98	-6.500	13.48		%
IRF				24.4	-15.70	59.9		%
LFR			L	75.6	15.70	59.9		%
MFR				19.8	-9.70	29.5		%
HFR				4.6	-6.00	10.6		%
RHE				34.6	0.00	34.6		pg



• The results from the three tests were similar. Compared with the results before admission, there was an increase in WBC and HGB. The DIFF scattergram showed that Lym particles further shifted towards the upper right corner, with an increase in the density of particles in the upper region, suggesting that abnormal cells still existed and their percentage might be increasing.

Peripheral blood morphology results from Jul. 6 to Aug. 3

Analysis time 09:07, Jul. 6, 2022

White blood cells									
	White blood cells	302	100%						
L	Segmented neutrophils	48	15.9						
	Band neutrophils	20	6.6						
	Lymphocytes	97	32.1						
L	Monocytes	3	1.0						
	Eosinophils	4	1.3						
	Metamyelocytes	1	0.3						
ł	Myelocytes	17	5.6						
1	Blasts	112	37.2						
	Non-white blood cells	116	%						
l	Nucleated RBCs	93	30.8						
	Giant platelets	1							
	Large platelets	3							
	Smudge cells	14	4.6						

Blasts



Analysis time 08:14, Jul. 13, 2022

	White blood ce	lls	
	White blood cells	301	100%
L	Segmented neutrophils	58	19.3
	Band neutrophils	20	6.6
	Lymphocytes	90	29.9
L	Monocytes	3	1.0
	Eosinophils	11	3.7
	Metamyelocytes	3	1.0
l	Myelocyte	15	5.0
	Promyelocytes	1	0.3
l	Blasts	99	32.9
	Reactive lymphocytes	1	0.3
	Non-white blood cells	116	%
I.	Nucleated RBCs	66	21.9
	Large platelets	2	
	Smudge cells	14	4.7

White blood cells 300 White blood cells 100% L Segmented neutrophils 50 16.7 Band neutrophils 15 50 H Lymphocytes 133 44.4 4 L Monocytes 1.3 ! Myelocytes 10 3.3 Promyelocytes 4 1.3 83 277 ! Blasts Reactive lymphocytes 1 0.3 Non-white blood cells 65 % ! Nucleated RBCs 33 11.0 Giant platelets 1 Large platelets 10 Smudge cells 11 3.7

Analysis time 08:51, Aug. 8, 2022



- Three peripheral blood morphology results were consistent with the CBC results. Myeloblasts and immature granulocytes at all stages of development were visible. Due to reasons including drug-induced cell differentiation, myeloblast morphology was atypical, with small cell bodies and less fine and loose chromatins, but nucleoli and Auer bodies were still visible.
- Despite almost 3 months of treatment, the patient's condition further deteriorated.



CBC results on Aug. 24

- WBC increased with classification results of poor reliability that were blocked; RBC and HGB decreased; extremely low PLT count. •
- In the WNB channel, a large number of NRBCs were observed, and some WBC particles extended upwards, indicating the presence of a large amount of abnormal cells. In the DIFF scattergram, the particles in the Lym and Neu regions extended together towards the high-fluorescence area and eventually merged, causing particle clusters to be indistinguishable and presenting a droplet-like appearance, indicating the presence of a large number of immature granulocytes and blasts.

Peripheral blood morphology results on Aug. 24							
Analysis time 08:57, A	ug. 24, 2022	2			3		
White bloc	od cells					and the second	
White blood cells	325	100%			and the second s		
Segmented neutrophils	2	0.6			1000		
Band neutrophils	2	0.6					
Lymphocytes	11	3.4				Concellant of the second	
Monocytes	2	0.6		Immature			
Eosinophils	1	0.3		granulocytes:			
Metamyelocytes	4	1.2		5 ,	600		
Myelocytes	39	12.0				A Report	
Promyelocytes	49	15.1				the second	
Blasts	215	66.2					
Non-white blood cells	92	%			AAAU		
Nucleated RBCs	8	2.5			SVO-		
Smudge cells	79	24.3					

09 HemaCase—Clinical Case Booklet

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Blasts



• Pseudo Pelger–Huët anomaly was observed in some neutrophilic metamyelocytes and band neutrophils. The blasts accounted for 66.2%, varied in sizes, and were generally larger compared with previous results, with 2–3 nucleoli visible in some blasts.

Discussion

• Although scattergrams cannot directly reflect the morphology of cells, the changes in their optical signal values can faithfully reflect changes in cell morphology. For example, in this case, although blasts appeared frequently, the monoclonal proliferation was inhibited by the drug and the transcriptional and translational activities of nucleic acids were weakened. Meanwhile, the cell volume was reduced and chromatin was aggregated. As a result, the binding of fluorescent dye was not as pronounced as in typical blasts during the CBC, and Lym and Mon particle cluster distribution were slightly suppressed in the fluorescence direction on the DIFF scattergram. Nevertheless, the Lym particle cluster was more skewed to the right than that under normal circumstances, indicating the presence of abnormal cells. In the last examination, a large number of myeloblasts and immature granulocytes proliferated and were released, rendering the DIFF scattergram more distinct features.

Case 02 Acute promelocytic leukemia (APL)

Clinical information

The patient was a 52-year-old female, previously healthy, who presented to the hospital on Nov. 1 with left ear bleeding without any obvious cause (which later stopped spontaneously). One week prior to the visit, the patient had experienced fatigue without obvious cause. The skin showed no petechiae or ecchymoses, and there was no hepatosplenomegaly or superficial lymphadenopathy.

Sec results

ParameterAlarmResultUnitWBC&H93.0510.9/LNeu#&RL1.1510.9/LLym#60.0010.9/LMon#6.0010.9/LBas#H0.9010.9/LMG#R0.8010.9/LNeu%&RL1.2%Mon%Neu%&RL0.00%Bas#-0.00Non%Neu%&RL0.00%Bas#-0.9MG%MG%-1.0MG%-1.0MG%-1.0MCH-3.3g/LMCHRDW-SD-5.0RDW-SD-5.0PCTL17PCTL0.015PCTL0.015PLC-1.00%/LPALC2.3.3%PLC-1.00%/LPLCL1.00%/LPLC-1.00%/LPLC-8.9RDW-SD-2.3.3MCH-1.2.3PCTL1.00%/LPLC-1.00%/LPLC-1.00%/LPLC-1.00%/LPLC-1.00%/LPLC-1.00%/LPLC<						CBC	C results on Nov. 1	
WBC & H 93.05 10^9/L Neu# & R L 1.15 10^9/L Neu# & **** 10^9/L Mon# **** 10^9/L Bas# H 0.00 10^9/L Sas# H 0.90 10^9/L Neu# & R 0.80 10^9/L Neu% R L 1.2 % MG% R 0.90 10^9/L Neu% L 0.00 % Mon% R 0.9 % MG% R 0.9 % MCC 3.1.2 pg MCH 3.2.0 fL PUT<	Parameter	A	larm		Result	Unit	RRC	PLT
Neu## 8 R L 1.15 10^9/L Lym# - - 10^9/L Mon# - L 0.00 10^9/L Bas# - L 0.00 10^9/L Bas# - No 0.09 10^9/L Bas# - L 0.00 10^9/L Neu%6 & R 0.00 10^9/L Neu%6 & R 0.00 10^9/L Mon% - - **** % Mon%6 - 0.00 % Bas% L 1.2 % Mon% - 0.0 % Bas% L 0.0 % Mon% R 0.0 % Mo% R 0.0 % Mo% R 26.6 % McHC S2.0 ft	WBC	&		н	93.05	10^9/L	RBC	1 21
	Neu#	&	R	L	1.15	10^9/L		
Mon# **** 10^9/L Eos# L 0.00 10^9/L Bas# H 0.90 10^9/L Bas# H 0.90 10^9/L Mon% R 0.80 10^9/L Won% **** % Mon% 0.9 % Bas# 1.0 % MG% R 0.9 Bas# 1.0 % MG% R 0.9 Bas# 1.0 % MCV 9.3.6 fL MCHC 31.2 pg MCHC 33.3 g/L RDW-CV 15.1 % RDW-CV 5.2.0 fL PLT L 17 10^9/L MPV 8.9 fL PLC L 4 10^9/L PLC L 4 10.9/L	Lym#				****	10^9/L		
Eos# L 0.00 10^9/L Bas# H 0.90 10^9/L MG# R 0.80 10^9/L Neu% & R 12 yym% **** % Mon%6 * 12 Eos% L 0.0 % Mon%6 * 0.0 Sas% 1.0 % MG% R 0.9 % RBC L 28.4 10^12/L Ho 93.6 fl 10.4 MCH 31.2 pg MCHC 33.3 g/L RDW-CV 52.0 fl MPV 8.9 fl PLT L 17 PLC L 0.015 PLCC L 4 10^9/L PLCC L 4 10/9/L PLCC 22.3 % -Abn. Lymph/blast? - Nameture Gran? -Basophilia - Leukocytosis -Anemia - Thrombocytopeni </td <td>Mon#</td> <td></td> <td></td> <td></td> <td>****</td> <td>10^9/L</td> <td></td> <td></td>	Mon#				****	10^9/L		
Bas# H 0.90 10^9/L MG# R 0.80 10^9/L Neu% & R L 1.2 %6 Mon%	Eos#			L	0.00	10^9/L		
MG# R 0.80 10^9/L Neu% & R L 1.2 % Mon%	Bas#			н	0.90	10^9/L		
Neugé & R L 1.2 % Lym% **** % Mon% **** % Mon% **** % Eos% L 0.0 % Bas% 1.0 % MG% R 0.9 % RBC L 2.84 10^12/L HGB L 89 g/L HCT L 26.6 % MCH 31.2 pg MCH 33.3 g/L MCH 33.3 g/L MCH 52.0 fL PPLT L 17 10^9/L MPV 8.9 fL -Abn. WBC scattergram PLCC L 4 10^9/L P-LCC L 4 10^9/L P-LCC L 4 10^9/L NRBC# 0.195 10^9/L NRBC# 0.21 /100WBC	MG#		R		0.80	10^9/L		
Lymp% **** % Mon% **** % Mon% L 0.0 % Eos% L 0.0 % Bas% 0.10 % DIFF MG% R 0.9 % MG% R 0.9 % MG% L 2.84 10^12/L HGB L 8.9 g/L MCT L 2.6.6 % MCH 31.2 pg MCH 33.3 g/L NCHC 33.3 g/L NCH 52.0 f. PUT L 17 NPV 8.9 f. PDW H 18.0 PCT L 4 PCT	Neu%	&	R	L	1.2	%	0 100 200 fL	0 10 20 30 fL
Mon% **** % Eos% L 0.0 % Eos% 1.0 % MG% R 0.9 % MG% R 0.9 % MG% R 0.9 % HGB L 89 g/L HCT L 26.6 % MCH 31.2 pg MCHC 333 g/L NCHC 333 g/L NCHC 52.0 fL PUT L 17 10^9/L MCY 8.9 fL PDW H 18.0 PCT L 4 10^9/L PCT L 4 10^9/L P-LCC L 4 10^9/L P-LCC L 4 10/9/L NBBC# 0.195 10/9/L NBRC# 0.21 /100WBC	Lym%				****	%		
Eos% L 0.0 % Bas% 1.0 % MG% R 0.9 % RBC L 2.84 10^12/L HGB L 89 g/L HCT L 2.6.6 % MCH 31.2 pg MCHC 33.3 g/L NCHC 33.3 g/L NCHC 52.0 fL PUT L 17.4 NEV 8.9 fL PDW H 18.0 PCT L 0.015 P-LCC L 4 P-LCC L 4 NRBC# 0.195 10.9/L NRBC# 0.21 10.9/L	Mon%				****	%		
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RDW-SD 52.0 fL PLT L 17 10^9/L MPV 8.9 fL Alarm PDW H 18.0 Alarm PCT L 0.015 % - Abn. WBC scattergram - Blasts? - Basophilia - Leukocytosis - Anemia - Mnemia - Anemia - Thrombocytopeni NRBC# 0.21 /100WBC - Immature Gran? - Thrombocytopeni	RDW-CV				15.1	%	SS	FI
PelT L 17 10^9/L MPV 8.9 fL Alarm PDW H 18.0 - Abn. WBC scattergram - Basophilia P-LCC L 4 10^9/L - Abn. WBC scattergram - Basophilia P-LCR 22.3 % - Abn Lymph/blast? - Anemia NRBC# 0.195 10^9/L - Immature Gran? - Thrombocytopeni	RDW-SD				52.0	fL		
MPV 8.9 fL Alarm PDW H 18.0 - Abn. WBC scattergram - Basophilia PCT L 0.015 % - Abn. WBC scattergram - Basophilia P-LCC L 4 10^9/L - Abn Lymph/blast? - Anemia NNBC# 0.195 10^9/L - Immature Gran? - Thrombocytopeni	PLT			L	17	10^9/L		
PDW H 18.0 PCT L 0.015 % - Abn. WBC scattergram - Basophilia P-LCC L 4 10^9/L - Blasts? - Leukocytosis P-LCR 22.3 % - Abn Lymph/blast? - Anemia NRBC# 0.195 10^9/L - Immature Gran? - Thrombocytopeni	MPV				8.9	fL	Ala	arm
PCTL0.015%- Abn. WBC scattergram- BasophiliaP-LCCL410^9/L- Blasts?- LeukocytosisP-LCR22.3%- Abn Lymph/blast?- AnemiaNRBC#0.19510^9/L- Immature Gran?- ThrombocytopeniNRBC%0.21/100WBC- Abn Lymph/blast?- Anemia	PDW			Н	18.0			
P-LCCL410^9/L- Blasts?- LeukocytosisP-LCR22.3%- Abn Lymph/blast?- AnemiaNRBC#0.19510^9/L- Immature Gran?- ThrombocytopeniNRBC%0.21/100WBC- Immature Gran?- Immature Gran?	РСТ			L	0.015	%	- Abn. WBC scattergram	- Basophilia
P-LCR 22.3 % - Abn Lymph/blast? - Anemia NRBC# 0.195 10^9/L - Immature Gran? - Thrombocytopeni NRBC% 0.21 /100WBC - Immature Gran? - Thrombocytopeni	P-LCC			L	4	10^9/L	- Blasts?	- Leukocytosis
NRBC#0.19510^9/L- Immature Gran?- InfrombocytopeniNRBC%0.21/100WBC	P-LCR				22.3	%	- Abn Lymph/blast?	- Anemia
NRBC% 0.21 /100WBC	NRBC#				0.195	10^9/L	- Immature Gran?	- Thrombocytopenia
	NRBC%				0.21	/100WBC		

• WBC increased with poor classification reliability; RBC and HGB decreased; extremely low PLT count.

• The WNB channel showed the presence of NRBCs, and some particles extended upward. The DIFF channel showed particles in the Neu region extending upward to the left and merging with particles in the Mon region to advance towards the high-fluorescence region, leading to some particles entering the abnormal lymphocytes/blasts region. The instrument also gave an alarm.

C Peripheral blood morphology examination

	White	blood cells	
	White blood cells	500	100%
L	Segmented neutrophils	6	1.2
	Band neutrophils	1	0.2
L	Lymphocytes	5	1.0
L	Eosinophils	2	0.4
	Metamyelocytes	1	0.2
	Promyelocytes	4	0.8
ļ	Blasts	473	94.6
	Reactive lymphocytes	8	1.6
	Non-white blood cells	70	%
!	Nucleated RBCs	4	0.8
	Smudge cells	60	12.0
	Р	latelets	
	PLT estimate	Estimated result	Estimatio method
	Platelet concentration	29*10^9/L	Manual
	Red	blood cells	
	Size	Degree	%
ļ	Anisocytosis	3+	
	Macrocytes	0	1.2
!	Microcytes	3+	40.4
	Color	Degree	%
	Hypochromic cells	0	3.1
	Polychromasia	0	0.6
	Shape	Degree	%
ļ	Poikilocytosis	1+	
ļ	Schistocytes	1+	1.7
	Echinocytes	0	0.2
	Elliptocytes	0	0.4
	Ovalocytes	0	8.7
	Stomatocytes	0	0.0
	Target cells	0	0.0
	Teardrop cells	0	1.5



Results from manual re-classification

Segmented neutrophils accounted for 1.2%, band neutrophils accounted for 0.2%, metamyelocytes accounted for 0.2%, promyelocytes accounted for 0.6%, lymphocytes accounted for 1%, and abnormal promyelocytes accounted for 94.8%. Abnormal promyelocytes exhibited irregular cell bodies and nuclei with twisting and folding patterns. Chromatins were fine and nucleoli were visible. The internal and external plasma was visible in few cells, with indistinct granules while Auer's bodies were visible in the cytoplasm of some abnormal cells.

Other examinations

ltem	Parameter	Result	Reference range	ltem	Result
	PT	16.3	9.4-12.5	Bone	APL cells accounted for 86%, with POX-5, +37, +++28, ++++30/100, suggesting
	APTT	23.8	25.4-38.4	marrow	APL. Please take into account clinical findings and other genetic tests.
Congulation	ТТ	18.6	11.0-17.8		The immature myeloid cells accounted for 94.55% of the nucleated cells in the
Coagulation	FIB	118	200-400		bone marrow, with large side-scatter angles; strongly positive for CD9:
	DD	2574	<250	Flow	positive for CD117, CD45, CD34, CD64, CD13, CD33, CD123, and cMPO;
	FDP	45.90	<5.00	cytometry	weakly positive for CD7 and CD71; negative for CD22, CD10, CD19, CD38, CD14, CD15, CD11b, CD25, CD11c, DR,
	К	2.94	3.50-5.30mmol/L		GlyA, cCD79a, cCD3, and other antigens
Chemistry	LDH	691	120-250U/L	Chromosome	46, XX, t(15; 17)(q24; q21)
	CRP	6.2	<3.0mg/L	Gene	PML-RARA 74.49%, FLT-ITD (high) 44.23%

Case analysis

The main clinical manifestations of the patient were asthenia and bleeding; CBC showed anemia with markedly elevated WBCs and markedly reduced platelets. The coagulation results were abnormal, with markedly increased D-dimer, suggesting a tendency for DIC. Lactate dehydrogenase (LDH) was markedly elevated, while typical abnormal promyelocytes were observed in both peripheral blood and bone marrow. The diagnosis of acute promyelocytic leukemia (high-risk type) was further confirmed with the results of flow cytometry and karyotype analysis.

Starting on Nov. 1, platelets and fresh frozen plasma (Fib supplementation) were given for supportive treatment to correct DIC tendency.

Hydroxyurea, cytarabine, and mitoxantrone were temporarily given from Nov. 1 to Nov. 4 for WBC reduction therapy;

Arsenic + ATRA induction therapy was started on Nov. 3;

Parameter	Alarm	1	Result	Unit
WBC			4.48	10^9/L
Neu#	R	L	0.77	10^9/L
Lym#			****	10^9/L
Mon#			****	10^9/L
Eos#		L	0.00	10^9/L
Bas#		н	0.12	10^9/L
IMG#	R		0.04	10^9/L
Neu%	R	L	17.2	%
Lym%			****	%
Mon%			****	%
Eos%		L	0.1	%
Bas%		н	2.8	%
IMG%	R		1.0	%
RBC		L	2.36	10^12/L
HGB		L	74	g/L
НСТ		L	22.1	%
MCV			93.8	fL
МСН			31.6	pg
MCHC			337	g/L
RDW-CV			14.5	%
RDW-SD			50.2	fL
PLT		L	49	10^9/L
MPV			8.3	fL
PDW			16.6	
PCT		L	0. 041	%
P-LCC		L	8	10^9/L
P-LCR			15.5	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC

CBC results on Nov. 5

FL



DIFF



WNB





Alarm

- Abn. WBC scattergram
- Blasts?
- Abn Lymph/blast?
- Atypical Lymph?
- Neutropenia
- Anemia
- Thrombocytopenia

Parameter	Alarm	Result	Unit
WBC	L	1.11	10^9/L
Neu#	L	0.70	10^9/L
Lym#	L	0.38	10^9/L
Mon#	L	0.01	10^9/L
Eos#		0.02	10^9/L
Bas#		0.00	10^9/L
IMG#		0.00	10^9/L
Neu%		62.2	%
Lym%		34.4	%
Mon%	L	1.0	%
Eos%		2.2	%
Bas%		0.2	%
IMG%		0.1	%
RBC	L	2.87	10^12/L
HGB	L	91	g/L
НСТ	L	27.2	%
MCV		94.8	fL
МСН		31.6	pg
MCHC		333	g/L
RDW-CV	Н	18.5	%
RDW-SD	Н	64.7	fL
PLT		123	10^9/L
MPV		8.3	fL
PDW		16.0	
РСТ	L	0.103	%
P-LCC	L	19	10^9/L
P-LCR		15.0	%
NRBC#		0.000	10^9/L
NRBC%		0.00	/100WBC



After treatment, some improvement in anemia and thrombocytopenia was achieved, and the WBC concentration also decreased significantly. The scattergram showed that the decreased WBCs were mainly particles in the region of monocytes and abnormal cells. Because abnormal cells have fewer granules and a higher content of intracellular nucleic acid material, they typically fall in the Mon region of the DIFF scattergram, leading to a false increase in Mon. Therefore, in practice, it is advisable to set corresponding retesting rules for Mon elevation and to perform morphology screening of abnormal cells for samples that trigger the rules.

Discussion

Acute promyelocytic leukemia (APL) is an acute myeloid leukemia featured by malignant proliferation of abnormal promyelocytes and cytogenetic abnormalities t(15;17)(q22;q12) and PML-RARa. Its morphological characteristics are equivalent to those of acute promyelocytic leukemia (M3 subtype) in the FAB classification scheme. AML-M3 emphasizes abnormal promyelocyte morphology, with morphological features including internal and external plasma, "buttock" and "butterfly" cells, as well as faggot cells.

In addition to the common clinical features of AML, APL has the following characteristics:

- Granules in the cytoplasm of abnormal promyelocytes contain a large amount of procoagulant enzymes, often causing patients to develop DIC. Skin and mucous membrane bleeding is the most obvious sign, followed by gastrointestinal tract, urinary tract, respiratory tract, and vaginal bleeding, while intracranial bleeding, as the most severe case, is one of the causes of death. In this case, the patient experienced bleeding in the ear canal that stopped spontaneously, but DIC occurred soon after hospitalization. Fortunately, the diagnosis and hospitalization were timely.
- All-trans retinoic acid (ATRA) can induce the differentiation of APL cells into mature cells, while arsenic trioxide (ATO) can induce their apoptosis, resulting in a high remission rate. Therefore, early diagnosis and treatment of APL are key to improving the remission rate.

Case 03 Acute myeloblastic leukemia with maturation (M2)

Clinical information

The patient, a 60-year-old female, was admitted to the hospital on Aug. 16, 2021 with dizziness and anemia.

OBC results

Parameter	A	larn	n	Result	Unit		R	3C				PLT	
WBC	&		Н	34.99	10^9/L								
Neu#	&	R	Н	13.25	10^9/L								
Lym#				****	10^9/L				;	110			
Mon#				****	10^9/L								1
Eos#		R		0.04	10^9/L						_		1
Bas#		R	Н	0.27	10^9/L								
IMG#		R		3.56	10^9/L	LL,		<u> </u>		1		~	
Neu%	&	R	L	37.8	%	0	100	200	fL	0	10	20	30
Lym%				****	%								
Mon%				****	%								
Eos%		R	L	0.2	%		D	IEE				W/NI	R
Bas%		R		0.8	%		D					VVINI	J
IMG%		R		10.2	%	FL I				FS			
RBC			L	2.70	10^12/L								
HGB			L	89	g/L		601.0						
НСТ			L	27.2	%			2			52	A State	
MCV			Н	100.8	fL			ALC: NO					1
MCH				33.1	pg						07075		
MCHC				328	g/L		3				1		
RDW-CV				14.1	%				SS				F
RDW-SD				51.5	fL								
PLT			L	31	10^9/L				٨١٦	rm			
MPV				10.5	fL				Ala				
PDW			Н	17.1									
PCT			L	0.033	%	- Ak	on. WBC	scattergr	am				
P-LCC			L	10	10^9/L	- BI	asts?	h/blact?					
P-LCR				31.3	%	- At	on Lymp mature	Gran					
NRBC#				2.762	10^9/L	- 111	matule	Gran					
NRBC%				7.89	/100WBC								

- WBC increased with poor classification reliability; moderate anemia; PLT decreased.
- In the WNB channel, the NRBC region was significantly dense, and the WBC, NRBC, and Bas particle clusters were
 clearly distinguished, ensuring the accuracy of WBC. In the DIFF scattergram, the particles in the Mon region were
 dense and could not be effectively distinguished from Lym, with a large number of particles extending up to the
 abnormal cell region, suggesting the presence of abnormal cells. In addition, the left margin of the mixed Lym and
 Mon particle cluster was concave to the right, suggesting that the abnormal cells were possibly of myeloid lineage.
 The Neu particle cluster extended toward the high-fluorescence region, suggesting a left shift of the nucleus.

Peripheral blood morphology examination

	White bl	ood cells	
	White blood cells	200	100%
L	Segmented neutrophils	25	12.5
	Band neutrophils	3	1.5
L	Lymphocytes	7	3.5
	Monocytes	б	3.0
н	Basophils	13	6.5
1	Metamyelocytes	15	7.5
1	Myelocytes	5	2.5
	Promyelocytes	1	0.5
1	Blasts	125	62.5
	Non-white blood cells	70	%
1	Nucleated RBCs	15	7.5
	Large platelets	1	
	Smudge cells	41	20.5
	Plat	elets	Estimation
	PLI estimate	Estimated res	uit method
	Platelet concentration	12*10/9/L	Manual
	Platelet concentration		IvidiTudi
	Size	Degree	%
1	Anisocytosis	2+	
	Macrocytes	0	2.0
1	Microcytes	2+	11.9
	Color	Degree	%
1	Hypochromic cells	2+	14.9
	Polychromasia	0	1.0
	Shape	Degree	%
	Poikilocytosis	0	
	Schistocytes	0	1.0
	Echinocytes	0	0.2
	Elliptocytes	0	0.0
	Elliptocytes Ovalocytes	0 0	0.0 4.4
	Elliptocytes Ovalocytes Stomatocytes	0 0 0	0.0 4.4 0.4
	Elliptocytes Ovalocytes Stomatocytes Target cells	0 0 0	0.0 4.4 0.4 0.2

Results from manual re-classification

The immature granulocytes accounted for 10.5%, which were mainly metamyelocytes; blasts accounted for 62.5%; NRBCs accounted for 7.5%, with occasional large platelets; no platelet aggregation was observed. The PLT estimate was generally consistent with the CBC result. Blasts showed oval cell bodies, eccentric nucleus, fine sandy chromatins, 2–3 nucleoli, and blue transparent cytoplasm. Abnormal myelocytes were also visible with pink cytoplasm, but the nucleoli were still visible in the nucleus, showing old nuclei and young cytoplasm.

Bone marrow cytology examination

- Blasts accounted for 84.5% and histochemical staining showed POX+.
- Acute myeloid leukemia with a predilection for M2, further subtyping is recommended through integration of flow cytometry and relevant molecular biology testing.

Case analysis

Characteristics of M2 (acute myeloid leukemia with partial differentiation):

- Hemogram: Significant anemia, moderate leukocytosis similar to M1, mainly myeloblasts and promyelocytes, and moderate to severe thrombocytopenia.
- Myelogram: A diagnosis of this subtype can be made with 30%–89% blasts in bone marrow (NEC) with morphological abnormalities; monocytes < 20%, promyelocytes and cells in earlier stages > 10%.
- Cell histochemical staining: POX and SBB staining were positive; PAS staining: most blasts were negative, most promyelocytes were weakly positive with fine granular positivity; neutrophilic alkaline phosphatase (NAP): activity was obviously decreased, even disappearing. When accompanied by infection, NAP score might increase transiently.

Based on the myelogram characteristics and histochemical staining, the diagnosis of acute myeloid leukemia M2 subtype was favored. Further subtyping requires the results of flow cytometry and relevant genetic testing.

Case 04 Acute myelomonocytic leukemia (AMML)

Clinical information

The patient was a 49-year-old female who presented with asthenia more than 10 days ago with no obvious cause. She had a medical history of "malignant breast tumor" for 7 years and had undergone surgical treatment, with 1 month of post-operative radiotherapy and 8 cycles of chemotherapy.

CBC results

Parameter	Al	arm		Result	Unit
WBC	&		L	0.95	10^9/L
Neu#	&	R	L	0.39	10^9/L
Lym#	&	R	L	0.46	10^9/L
Mon#		R	L	0.06	10^9/L
Eos#			L	0.00	10^9/L
Bas#				0.04	10^9/L
IMG#		R		0.01	10^9/L
Neu%	&	R	L	40.6	%
Lym%	&	R	н	47.9	%
Mon%		R		6.8	%
Eos%			L	0.1	%
Bas%			н	4.6	%
IMG%		R		1.2	%
RBC			L	1.43	10^12/L
HGB			L	50	g/L
НСТ			L	15.6	%
MCV			н	109.1	fL
МСН			Н	35.3	pg
MCHC				323	g/L
RDW-CV				15.8	%
RDW-SD			н	62.6	fL
PLT			L	30	10^9/L
MPV				11.8	fL
PDW			н	18.2	
РСТ			L	0.035	%
P-LCC			L	10	10^9/L
P-LCR				33.0	%
NRBC#				0.012	10^9/L
NRBC%				1.24	/100WBC



• Decreased trilineage, severe anemia, increased Bas%.

• The particle clusters in the Diff scatter plot were clearly defined, and the Mon particle clusters were distributed in the high-fluorescence region, which were suspected to be blasts.

Peripheral blood morphology examination

	White blood	cells	
	White blood cells	200	100%
L	Segmented neutrophils	64	32
	Band neutrophils	6	3.0
	Lymphocytes	53	26.5
L	Monocytes	1	0.5
н	Basophils	4	2.0
1	Metamyelocytes	4	2.0
1	Myelocytes	2	1.0
1	Blasts	66	33.0
	Non-white blood cells	59	%
1	Nucleated RBCs	2	1.0
	Giant platelets	1	
	Large platelets	7	
	Smudge cells	10	5.0
	Platelets	5	Estimation
	PLT estimate	Estimated resu	method
	Platelet concentration	40*10^9/L	Automated
	Platelet concentration	62*10^9/L	Manual
	Red blood	cells	
	Size	Degree	%
1	Anisocytosis	2+	
	Macrocytes	0	1.9
1	Microcytes	2+	13.2
	Color	Degree	%
1	Hypochromic cells	3+	53.9
	Delvelenene ete		
	Polychromasia	0	0.5
	Shape	0 Degree	0.5 %
	Shape Poikilocytosis	0 Degree 0	0.5 %
	Shape Poikilocytosis Schistocytes	0 Degree 0 0	0.5 % 0.8
	Shape Poikilocytosis Schistocytes Echinocytes	0 Degree 0 0 0	0.5 % 0.8 0.3
	Shape Poikilocytosis Schistocytes Echinocytes Elliptocytes	0 Degree 0 0 0	0.5 % 0.8 0.3 0.3
	Shape Polikilocytosis Schistocytes Echinocytes Elliptocytes Ovalocytes	0 Degree 0 0 0 0 0 0	0.5 % 0.8 0.3 0.3 14.7
	Shape Polikilocytosis Schistocytes Echinocytes Elliptocytes Ovalocytes Stomatocytes	0 Degree 0 0 0 0 0 0 0 0	0.5 % 0.8 0.3 0.3 14.7 0.0
	Shape Poikilocytosis Schistocytes Echinocytes Elliptocytes Ovalocytes Stomatocytes Target cells	0 Degree 0 0 0 0 0 0 0 0 0	0.5 % 0.8 0.3 0.3 14.7 0.0 0.0
	Shape Poikilocytosis Schistocytes Echinocytes Elliptocytes Ovalocytes Stomatocytes Target cells Teardrop cells	0 Degree 0 0 0 0 0 0 0 0 0 0 0 0	0.5 % 0.8 0.3 0.3 14.7 0.0 0.0 1.9



Results from manual re-classification

Immature granulocytes accounted for 3% and blasts accounted for 33%; the cell bodies were large and irregular in shape; the cytoplasm was dark blue, and cytoplasm of some cells contained dust-like granules; the chromatins were coarse and granular; 1-2 large and pronounced nucleoli were visible; cup-like blasts were occasionally observed. Pancytopenia, blasts in peripheral blood > 20%, suggesting AML.

Other examinations

ltem	Result
Bone marrow	Abnormal granulocytic hyperplasia; granulocytes at all stages of maturation were visible, with myeloblasts accounting for 27%, mainly intermediate and late-stage granulocytes. Monoblasts and promonocytes accounted for 39% and mature monocytes accounted for 6%. MPO (+); NAP (35%, 54 points). Extracellular iron (+), erythroblasts with intracellular iron were rare. Suggested diagnosis: AML-M4b.
Flow cytometry	A group of abnormal myeloid blasts were observed, accounting for 26.04% of the v cells, which was consistent with the AML phenotype
Chromosome	46,XX,t(6:9)(p23;q34.1)
Gene	DEK-CAN positive. FLT3-ITD allelic ratio: 0%. NPM1(-). FLT3-TKD negative

🖹 Case analysis

- Acute granulocytic leukemia (M4) is classified into four subtypes (per the classification criteria in China), of which M4b is characterized by the hyperplasia of monoblasts and promonocytes, with myeloblasts and promyelocytes accounting for > 20%.
- The MICM diagnosis of this case was AML with t(6;9) (p23;q34). DEK-NUP214 (also known as CAN) is a unique and relatively rare type of AML, accounting for 0.7% to 1.8% of AML. The translocation t(6;9) (p23;q34) results in the fusion of DEK on chromosome 6 and NUP214 on chromosome 9, forming a nucleoporin fusion protein that acts as an aberrant transcription factor and alters nuclear transport by binding soluble transport factors. The median onset ages in children and adults are 13 and 35 years, respectively. Patients often present with anemia and thrombocytopenia or show full blood count decreased. The prognosis is poor. Morphologically, mature type AML and acute myelomonocytic leukemia are the most common. Auer bodies can be observed in one-third of cases. MPO is positive, while NSE can be either positive or negative. About 44%–62% of patients show increased basophils in the bone marrow and peripheral blood (> 2%) (4.6% in peripheral blood for this case), which is rare in other types of AML. Most patients show granulocytic and erythroid dysplasia, and ring sideroblasts are observed in some patients, but megakaryocytic dysplasia is rare.

Case 05 Thrombotic thrombocytopenic purpura (TTP)

Clinical information

The patient was a 64-year-old male who experienced recurrent chest tightness and dizziness for 20 days, and fever for 5 days. He visited a local hospital and was discharged after the symptoms improved. However, he continued to experience recurrent chest tightness and dizziness after discharge, so he returned to the hospital for further examination 10 days later, and the relevant test results are as follows:

ltem	Parameter	Result	ltem	Paramet	Result	
	TBIL	18.9µmol/L	CDC	HGB	73g/L	
	DBIL	7.7µmol/L	CBC	PLT	26*10^9/L	
Chemistry	IBIL	11.2µmol/L			Markedly active bone marrow hyperplasia, increased erythroid proportion,	
	Creatinine	115µmol/L	Bone marrow	Bone marrow		
	LDH	892U/L			decreased myeloid proportion, and poor megakaryocyte platelet production	

Idiopathic thrombocytopenic purpura was suggested and EVANS syndrome was to be excluded. The patient had poor treatment response and experienced recurrent fever. Ten days later, he presented to the emergency department of our hospital with symptoms of delayed response and decreased calculation ability during the emergency treatment.

Sec results

Parameter	Ala	rn	า	Result	Unit
WBC	&			8.37	10^9/L
Neu#	&	R		6.68	10^9/L
Lym#	&			0.87	10^9/L
Mon#				0.75	10^9/L
Eos#		R		0.05	10^9/L
Bas#		R		0.02	10^9/L
IMG#				0.11	10^9/L
Neu%	&	R	н	79.7	%
Lym%	&		L	10.4	%
Mon%				9.0	%
Eos%		R		0.6	%
Bas%		R		0.3	%
IMG%				1.3	%
RBC			L	2.37	10^12/L
HGB			L	80	g/L
HCT			L	23.5	%
MCV				98.9	fL
MCH				33.8	pg
MCHC				341	g/L
RDW-CV			Н	17.2	%
RDW-SD			Н	58.2	fL
PLT	&		L	5	10^9/L
MPV		R		7.9	fL
PDW		R		15.5	
PCT		R	L	0.008	%
P-LCC		R	L	1	10^9/L
P-LCR		R		21.9	%
IPF				5.0	%
RET#		R	Н	0.3752	10^12/L
RET%		R	Н	15.82	%
IRF		R		22.8	%
LFR		R	L	77.2	%
MFR		R		15.5	%
HFR		R	Н	7.3	%
RHE		R		30.4	pg
NRBC#				0.034	10^9/L
NRBC%				0.41	/100WBC



- WBCs were generally normal; moderate anemia and extremely low PLT.
- In the RET scattergram, the RBC cluster extended towards the direction of low forward scattered light, even falling below the PLT particles, suggesting the presence of RBC fragments.

A Peripheral blood morphology examination

	Red b	lood cells		
	Size	Degree	%	1
!	Anisocytosis	3+		
	Macrocytes	0	0.3	4
	Microcytes	0	0.6	
	Color	Degree	%	
!	Hypochromic cells	3+	25.8	
	Polychromasia	0	3.2	
	Shape	Degree	%	
!	Poikilocytosis	3+		
	Schistocytes	3+	9.0	
	Echinocytes	0	0.5	
	Elliptocytes	0	0.2	
	Ovalocytes	0	2.7	
	Stomatocytes	0	0.5	
	Target cells	0	0.2	
	Teardrop cells	0	2.1	



Other examinations

ltem	Parameter	Result		
	TBIL	45.5µmol/L		
	DBIL	8.8µmol/L		
Chemistry	IBIL	36.7µmol/L		
	Creatinine	106µmol/L		
	LDH	1070U/L		
	Activity	<6%		
ADAMIS13	Titer	<0.6BU		

Case analysis

- Due to the failure to diagnose Thrombotic thrombocytopenic purpura (TTP) and initiate corresponding treatment in a timely manner, the patient's condition gradually worsened. The disease progressed from anemia and thrombocytopenia to fever and neurological symptoms, with a gradual decrease in PLT and gradual increases in IBIL and LDH. Finally, idiopathic thrombocytopenic purpura and EVANS syndrome were excluded by the discovery of intravascular hemolysis and schistocytes in peripheral blood morphology examination.
- Patients with Thrombotic thrombocytopenic purpura (TTP) suffer from significant thrombocytopenia and anemia due to loose platelets or fibrin depositing in the numerous small blood vessels, causing damage to passing platelets and RBCs. The decrease in ADAMTS13 activity leads to the accumulation of larger vWF aggregates on endothelial cells, forming many platelet-vWF thrombi that consume platelets, resulting in thrombocytopenia.

Case 06 Chronic myelomonocytic leukemia (CMML)

Clinical information

The patient was a 56-year-old male who was admitted for treatment on Aug. 14 due to fever and hepatosplenomegaly.

G CBC results

Parameter	Alar	m		Result	Unit
WBC			н	57.10	10^9/L
Neu#		R	н	21.86	10^9/L
Lym#		R	н	4.91	10^9/L
Mon#		R	н	30.26	10^9/L
Eos#		R	L	0.01	10^9/L
Bas#		R		0.09	10^9/L
IMG#		R		3.54	10^9/L
Neu%		R	L	38.3	%
Lym%		R	L	8.6	%
Mon%		R	н	53.0	%
Eos%		R	L	0.0	%
Bas%		R		0.1	%
IMG%		R		6.2	%
RBC			L	1.85	10^12/L
HGB			L	54	g/L
НСТ			L	16.6	%
MCV				89.5	fL
MCH				29.3	pg
MCHC				328	g/L
RDW-CV				16.0	%
RDW-SD				52.9	fL
PLT	&		L	60	10^9/L
MPV		R	н	14.6	fL
PDW		R		16.3	
PCT		R	L	0.064	%
P-LCC		R		37	10^9/L
P-LCR		R	н	61.0	%
IPF			н	37.7	%
RET#			L	0.0175	10^12/L
RET%				0.94	%
IRF			н	31.2	%
LFR			L	68.8	%
MFR			н	25.1	%
HFR			Н	6.1	%
RHE				32.1	pg



• WBC increased, mainly Mon; Neu#, Lym#, and Mon# increased; severe anemia; PLT decreased; IPF and IRF increased.

• The boundaries of each particle cluster in the Diff scattergram were clear, and particles in the Mon region were significantly dense and extended in the high-fluorescence direction, suggesting the possible presence of myeloblasts and promyelocytes. The Neu particle cluster also extended upwards to a higher region, suggesting the presence of myelocytes and metamyelocytes. Increased IPF and IRF suggested strong hematopoietic function of erythrocyte and platelet.

Peripheral blood morphology examination

	White	blood cells	
	White blood cells	205	100%
L	Segmented neutrophils	51	24.9
	Band neutrophils	10	4.9
L	Lymphocytes	16	7.8
	Monocytes	103	50.1
	Metamyelocytes	3	1.5
1	Myelocytes	4	2.0
1	Blasts	3	1.5
1	Reactive lymphocytes	15	7.3
	Non-white blood cells	70	%
	Giant platelets	1	
	Large platelets	9	
	Smudge cells	58	28.3
	Р	latelets	
	PLT estimate	Estimated result	Estimation method
	Platelet concentration	54*10^9/L	Automated
	Platelet concentration	66*10^9/L	Manual
	Red	blood cells	
	Size	Degree	%
1	Anisocytosis	2+	
	Macrocytes	0	0.0
	Microcytes	0	0.7
	Color	Degree	%
1	Hypochromic cells	2+	10.8
	Polychromasia	0	0.1
	Shape	Degree	%
	Poikilocytosis	0	
	Schistocytes	0	0.2
	Echinocytes	0	0.3
	Elliptocytes	0	0.3
	Ovalocytes	0	4.6
	Stomatocytes	0	0.0
	Target cells	0	0.4
	Teardrop cells	0	0.2



Results from manual re-classification

Immature granulocytes accounted for 3.5% and myeloblasts and promyelocytes accounted for 6.3%; some cells had larger cell bodies with scant cytoplasm of blue color; chromatins were fine, and nucleoli were visible; some cells had more cytoplasm with dust-like granules, chromatins were coarse, loose, and reticulate, and the nuclei were twisted and folded.

Bone marrow cytology examination

- Myelocytes and cells in earlier stages were visible, with increased proportion of neutrophilic metamyelocytes; unbalanced nucleus and cytoplasm growth as well as degranulation were observed in some cells; eosinophils were visible.
- Blasts accounted for 2.5%; monocyte percentage was high; premonocytes were visible.
- The morphology suggested chronic myelomonocytic leukemia. Further diagnosis was recommended through flow cytometry and CMML-related genetic testing.

📔 Case analysis

- Chronic myelomonocytic leukemia (CMML) was previously considered a subtype of myelodysplastic syndrome (MDS) due to its myelodysplastic and myeloproliferative characteristics. In 2001, WHO classified CMML as a myelodysplastic/myeloproliferative neoplasm (MDS/MPN).
- According to the FAB classification criteria, CMML with WBC < 13 × 10^9/L is the myelodysplastic subtype (MD-CMML), and CMML with WBC ≥ 13 × 10^9/L is the myeloproliferative subtype (MP-CMML).
- According to the 2016 WHO classification criteria, (i) CMML-0 can be diagnosed if the blasts are < 2% in peripheral blood and/or < 5% in bone marrow; (ii) CMML-1 can be diagnosed if the blasts are 2%–4% in peripheral blood and/or 5%–9% in bone marrow; (iii) CMML-2 can be diagnosed if the blasts are 5%–19% in peripheral blood, 10%–19% in bone marrow, and/or if Auer bodies are present;
- The patient in this case had splenomegaly, markedly increased WBCs in peripheral blood, and increased monocytes. Based on the diagnosis report of bone marrow smear, chronic myelomonocytic leukemia was suspected, and flow cytometry and CMML-related genetic testing were required for further diagnosis.

Case 07 MDS with single lineage dysplasia (MDS-SLD)

Clinical information

The patient was a 55-year-old female who was admitted with a history of "diagnosed with systemic lupus erythematosus for 16 years and found to have decreased platelets for more than 1 month". She had visited the hospital 1 month ago for systemic lupus erythematosus and was found to have decreased PLT and moderate anemia. RBC and platelet transfusions were given along with danazol therapy. Hemoglobin and platelet levels increased after treatment. The patient refused bone marrow aspiration and insisted on leaving the hospital. She is currently readmitted due to asthenia.

GRC results

Parameter	Alar	m		Result	Unit
WBC	&			7.66	10^9/L
Neu#	&	R		5.83	10^9/L
Lym#	&			1.53	10^9/L
Mon#				0.30	10^9/L
Eos#		R	L	0.00	10^9/L
Bas#		R		0.00	10^9/L
IMG#				0.19	10^9/L
Neu%	&	R	Н	76.0	%
Lym%	&			20.0	%
Mon%				3.9	%
Eos%		R	L	0.0	%
Bas%		R		0.1	%
IMG%				2.4	%
RBC			L	2.67	10^12/L
HGB			L	96	g/L
НСТ			L	30.1	%
MCV			Н	112.5	fL
МСН			н	36.1	pg
MCHC				321	g/L
RDW-CV			Н	18.4	%
RDW-SD			Н	75.2	fL
PLT		R	L	7	10^9/L
MPV		R		8.5	fL
PDW		R		15.0	
РСТ		R	L	0.006	%
P-LCC		R	L	1	10^9/L
P-LCR		R		18.9	%
NRBC#				0.302	10^9/L
NRBC%				3.95	/100WBC



• WBCs were generally normal; macrocytic anemia; extremely low PLT.

• In the WNB scattergram, a large number of pink particles are visible in the NRBC region.

Parameter	1-31	2-1	2-2	2-4	3-1
WBC	3.63	5.68	6.56	6.24	7.66
PLT	7	8	44	33	7
HGB	71	62	77	77	96
RBC	1.93	1.65	2.10	2.16	2.67

Peripheral blood morphology examination

	White blood	d cells	
	White blood cells	100	100%
н	Segmented neutrophils	81	81.0
	Band neutrophils	3	3.0
L	Lymphocytes	14	14.0
L	Monocytes	1	1.0
	Metamyelocytes	1	1.0
	Non-white blood cells	11	%
1	Nucleated RBCs	1	1.0
	Smudge cells	8	8.0
	Platele	ts	
	PLT estimate	Estimated	Estimation
		result	method
	Platelet concentration	13*10^9/L	Manual
	Red blood	cells	
	Size	Degree	%
1	Anisocytosis	2+	
1	Macrocytes	3+	30.9
	Microcytes	0	1.3
	Color	Degree	%
1	Hypochromic cells	2+	12.1
	Polychromasia	0	0.8
	Shape	Degree	%
1	Poikilocytosis	2+	
	Schistocytes	0	0.6
	Echinocytes	0	0.0
	Elliptocytes	0	0.4
	Ovalocytes	0	13.9
1	Stomatocytes	2+	6.4
	Target cells	0	3.2
	Teardrop cells	0	1.4



🔁 Bone marrow cytology examination

- Markedly active bone marrow hyperplasia, G/E = 0.89/1.
- Granulocytic hyperplasia was fair, mainly intermediate and late-stage granulocytes with visible eosinophiles.
- Active erythroid hyperplasia, mainly polychromatic normoblasts and orthochromatic normoblasts, with mild size variation of mature red blood cells.

CBC results, with increased stomatocytes.

- No megakaryocytes were observed on the slide and few platelets were found.
- The myelogram showed marked hyperplasia, with an inverted M/E ratio; no megakaryocytes and few platelets were observed.

🖹 Case analysis

The preliminary diagnosis of this case was MDS with single lineage dysplasia (MDS-SLD).

According to the WHO's revised classification of MDS (4th edition, 2016), the conventional karyotyping of MDS-SLD is as follows: any karyotype that does not meet the standard of MDS with isolated del(5q), and dysplasia is often predominant in 1 lineage, with dysplasia of 1–2 lineages in the CBC.

Additionally, the presence of increased stomatocytes in this case is often observed in hereditary stomatocytosis, disseminated intravascular coagulation, liver disease, systemic lupus erythematosus, alcohol poisoning, glutathione deficiency, hereditary spherocytosis, thalassemia minor, etc. Due to their poor deformability, stomatocytes are often retained in the splenic sinuses. In the acidic environment of the splenic sinus, the destruction of RBCs is more than three times greater in the spleen than in other sites due to the lack of glucose, reduced available ATP, and further increased cellular permeability to sodium.

Case 08 MDS with excess blasts (MDS-EB)

Clinical information

The patient, an 80-year-old female, was admitted to the hospital in early August with dizziness and anemia.

Gresults

Parameter	Alaı	rm		Result	Unit
WBC	&		н	13.04	10^9/L
Neu#	&	R		5.05	10^9/L
Lym#		R		2.35	10^9/L
Mon#		R		5.62	10^9/L
Eos#		R	L	0.01	10^9/L
Bas#		R		0.01	10^9/L
IMG#		R		1.20	10^9/L
Neu%	&	R	L	38.7	%
Lym%		R	L	18.0	%
Mon%		R	н	43.1	%
Eos%		R	L	0.1	%
Bas%		R		0.1	%
IMG%		R		9.2	%
RBC			L	2.63	10^12/L
HGB			L	75	g/L
НСТ			L	23.4	%
MCV				88.9	fL
MCH				28.4	pg
МСНС				320	g/L
RDW-CV			н	19.0	%
RDW-SD			н	61.9	fL
PLT		R		292	10^9/L
MPV		R		12.0	fL
PDW		R		16.9	
РСТ		R	н	0.352	%
P-LCC		R	н	125	10^9/L
P-LCR		R		42.8	%
NRBC#				0.261	10^9/L
NRBC%				2.00	/100WBC



• WBC was not low, Mon% increased, moderate anemia, normal PLT.

• The presence of grayish-white particles in the DIFF scattergram was mainly due to the extension of the Neu cluster towards the upper left corner. Dense particles in the Mon region led to ill-defined boundaries between Lym and Mon particle clusters, suggesting the possible presence of immature granulocytes, especially promyelocytes and blasts. The PLT histogram showed raised tail. Large platelets or interference from red blood cell fragments and microcytes should be ruled out.

🖉 Peripheral blood morphology examination

	White blo	od cells		Promyelocytes
	White blood cells	201	100%	Tromyclocytes
L	Segmented neutrophils	39	19.4	
	Band neutrophils	12	6.0	
L	Lymphocytes	39	19.4	
Н	Monocytes	31	15.4	
1	Metamyelocytes	12	6.0	
1	Myelocytes	15	7.5	
1	Promyelocytes	4	2.0	
1	Blasts	40	19.8	and the second s
	Reactive lymphocytes	9	4.5	
	Non-white blood cells	78	%	Myeloblasts
1	Nucleated RBCs	8	4.0	,
	Giant platelets	9		
	Large platelets	7		
	Smudge cells	43	21.4	
	Plate	lets		
	PLT estimate	Estimated	Estimation	
		result	method	
	Platelet concentration	241*10^9/L	Automated	
	Platelet concentration	268*10^9/L	Manual	

Results from manual re-classification

Immature granulocytes accounted for 16.9%, of which promyelocytes accounted for 3.4%, with large, round or oval cell bodies; nuclei were deviated to one side; chromatins were coarser than those of blasts; nucleoli were starting to close, and were less prominent than those of blasts; cytoplasm was blue, containing varying numbers of purple-red granules with different morphology and uneven distribution. Myeloblasts and promyelocytes accounted for 18.4%, with large oval cell bodies and blue cytoplasm without granules; chromatins were fine; 2–3 nucleoli could be seen.

The PLT estimate was consistent with CBC, and giant platelets were observed.

Eq Bone marrow cytology examination

- Active nucleated cell hyperplasia in bone marrow. Myeloids accounted for 54.5% and erythroids for 4%, with M:E ratio being 13.63:1.
- Myeloids: Promyelocytes and cells in earlier stages were observed, with a higher proportion of metamyelocytes and segmented neutrophils. Some myeloids exhibited unbalanced nucleus and cytoplasm development as well as degranulation.
- Erythroids: Mainly polychromatic normoblasts and orthochromatic normoblasts, with low proportions of cells at each developmental stage. Erythroblast morphology was generally normal. Mature RBCs varied in sizes.
- One megakaryocyte and small platelet clumps were observed on the slide.
- Blasts accounted for 17%, with medium-sized cell bodies, fine chromatins, and visible nucleoli; cytoplasm was blue and the amount was fair.
- The morphology was consistent with myelodysplastic syndrome with excess blasts (MDS-EB-2). Further diagnosis is recommended by flow cytometry analysis.

📄 Case analysis

MDS with excess blasts (MDS-EB) can be divided into 2 subtypes:

Туре	Dysplasia	Cytopenia	Ring sideroblasts	Blasts in bone marrow and peripheral blood	Conventional karyotyping
MDS-EB1	0–3 lineages	1–3 lineages	Any ratio	5%–9% in bone marrow or 2%–4% in peripheral blood, without Auer bodies	Any karyotype
MDS-EB2	0–3 lineages	1–3 lineages	Any ratio	10%–19% in bone marrow or 5%–19% in peripheral blood or with Auer bodies	Any karyotype

MDS-EB has a higher risk of AML transformation compared with other subtypes, and MDS-EB with excess blasts generally has a poorer prognosis. Treatment options include bone marrow transplantation or demethylation therapy. However, in this case, the patient was elderly, and interventions aimed at improving individual disease resistance and slowing dysplasia.

Case 09 Multiple myeloma (MM)

Clinical information

The patient was a 64-year-old male who was admitted to the emergency room due to severe anemia and lumbar vertebral fracture.

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Sec results

Parameter	Alarm		Result	Unit	RBC	PLT
WBC		L	3.51	10^9/L		
Neu#	R	L	0.86	10^9/L		
Lym#	R		1.58	10^9/L		
Mon#	R		1.06	10^9/L		
Eos#		L	0.01	10^9/L		
Bas#			0.00	10^9/L		
IMG#	R		0.01	10^9/L	0 100 200 fL	0 10 20
Neu%	R	L	24.4	%		
Lym%	R	Н	44.9	%		
Mon%	R	Н	30.2	%		
Eos%		L	0.4	%	DIFF	WNE
Bas%			0.1	%	FL .	EC 1
IMG%	R		0.4	%		
RBC		L	1.22	10^12/L	1	
HGB		L	47	g/L		
НСТ		L	13.8	%		
MCV		Н	112.6	fL		1993 (M)
MCH		Н	38.4	pg		Contraction of Contraction of
MCHC			341	g/L	SS	
RDW-CV			15.6	%		
RDW-SD		Н	63.7	fL	Alarn	n
PLT		L	24	10^9/L		
MPV			9.3	fL	- Pancytopenia	
PDW		Н	18.9		- Blasts?	
PCT		L	0.022	%	- Abn Lymph/blast?	
P-LCC		L	5	10^9/L	- Atypical Lymph?	
P-LCR			21.1	%		
NRBC#			0.000	10^9/L		
NRBC%			0.00	/100WBC		

• Slightly decreased WBC, decreased Neu; severe anemia and MCV was not small; extremely low PLT.

• Due to the decreased RBC/PLT, the area under the histogram was significantly reduced. Abnormal distribution of Lym and Mon particle clusters was observed in the DIFF scattergram, with Lym particles extending towards the upper right and Mon particles distributed in the high-fluorescence region, shifting to the right. Above Mon, some particles were identified as abnormal lymphocytes and blasts. The scattergram of this patient was overall similar to that of lymphocytic leukemia.

Teripheral blood morphology examination

	White blood cells								
	White blood cells	113	100%						
L	Segmented neutrophils	17	15.0						
	Band neutrophils	5	4.4						
н	Lymphocytes	46	40.7						
L	Monocytes	2	1.8						
	Reactive lymphocytes	1	0.9						
1	Plasma cells	42	37.2						
	Non-white blood cells	29	%						
	Smudge cells	17	15.0						
	Platele	ts							
	PLT estimate	Estimated	Estimation						
		result	method						
	Platelet concentration	12*10^9/L	Manual						
	Red blood	l cells							
	Size	Degree	%						
	Anisocytosis	0							
	Macrocytes	0	3.7						
1	Microcytes	2+	11.9						
	Color	Degree	%						
1	Hypochromic cells	3+	44.1						
	Polychromasia	0	0.9						
	Shape	Degree	%						
1	Poikilocytosis	2+							
1	Schistocytes	2+	1.0						
	Echinocytes	0	0.8						
	Elliptocytes	0	0.3						
1	Ovalocytes	2+	11.8						
	Stomatocytses	0	0.0						
	Target cells	0	0.4						
	Teardrop cells	0	0.3						



Results from manual re-classification

The classification results of Neutrophils and lymphocytes were consistent with the CBC results. Few Monocytes and a large number of plasma cells were observed under microscopy. The nuclei of the plasma cells were often round and eccentric, with chromatins agglutinated into large blocks and no nucleolus visible, and the cytoplasm was deep blue and appeared foamy. Rouleaux RBCs were also observed.

Content examinations

ltem	Result	Reference range
Rheumatoid factor	<20	0-20(IU/ml)
Anti-streptolysin "O"	<25	0-100(IU/ml)
C-reactive protein	4.45	0-8(mg/L)
Immunoglobulin A	41	0.7-4.0(g/L)
Immunoglobulin G	4.76	7-16(g/L)
Immunoglobulin M	0.18	0.4-2.3(g/L)
к light chain	512	6.7-22.4(mg/L)
λ light chain	13.4	8.3-27.0(mg/L)
κ light chain/ λ light chain	38.209	0.31-1.56
Immunophenotype electrophoresis	lgA-Kappa lambda monoclonal band	Negative
Total protein	99.5	65-85(g/L)
Albumin	28	40-55(g/L)
Globulin	71.5	20-40(g/L)
A/G ratio	0.39	1.2-2.4
Blood urea nitrogen	17.3	2.8-7.14mmol/L
Creatinine	174.78	40-135µmol/L

Case analysis

- The patient was diagnosed with multiple myeloma lgA-Kappa type.
- Multiple myeloma (MM) is a malignant disease characterized by clonal proliferation of plasma cells. It is more common in middle-aged and elderly population. With the aging of the Chinese population, the incidence of MM is increasing. Due to the massive secretion and deposition of monoclonal antibodies on the bone marrow stroma, osteoclasts are activated, leading to bone disease and hypercalcemia. Moreover, the malignant proliferation of plasma cells inhibits normal hematopoiesis, leading to anemia. The large amount of globulin also results in renal insufficiency. Therefore, the acronym CRAB, which stands for the first letter of the four main symptoms, is used as an alternative name for MM, commonly known as "crab disease".
- Patients with MM are prone to rouleaux formation of RBCs due to the increased fibrinogen and globulin in the blood, which shield the surface potential of RBCs and weaken the mutual repulsion between RBCs. If rouleaux RBCs are observed, attention must be paid to abnormal lymphocyte screening and protein electrophoresis assay to avoid the missed diagnosis of MM.

Case 10 B-cell lymphoblastic leukemia (B-ALL)

Clinical information

The patient was a 7-year-old male who was admitted to the hospital due to intermittent back pain for more than two months, along with chest pain and pain in both lower limbs for over a month. Abdominal ultrasound showed splenomegaly.

G CBC results

Parameter	Ala	arm		Result	Unit	RBC	PLT
WBC		&		6.09	10^9/L		
Neu#	&	R	L	0.88	10^9/L		
Lym#	&	R		3.50	10^9/L		
Mon#		R	L	0.00	10^9/L		
Eos#			Н	1.71	10^9/L		
Bas#				0.00	10^9/L		
IMG#		R		0.09	10^9/L		
Neu%	&	R	L	14.5	%	0 100 200 IL	0 10 20 30 1L
Lym%	&	R		57.3	%		
Mon%		R	L	0.1	%		
Eos%			Н	28.1	%	DIFF	WNB
Bas%				0.0	%		
IMG%		R		1.5	%	FL	FS
RBC				3.86	10^12/L		
HGB			L	107	g/L		
НСТ			L	32.6	%		and the second
MCV				84.4	fL	1	
МСН				27.7	pg		and the second sec
МСНС				328	g/L	SS	FL
RDW-CV				15.3	%		
RDW-SD				45.5	fL		
PLT			L	57	10^9/L	Ala	arm
MPV				8.0	fL	Dia etc2	
PDW				16.0		- Abn Lymph/blast?	
РСТ			L	0.046	%	- Immature Gran?	
P-LCC			L	11	10^9/L		
P-LCR				18.8	%		
NRBC#				0.039	10^9/L		
NRBC%				0.64	/100WBC		

• WBC was not low; Neu and Mon decreased; Eos increased; moderate anemia; PLT decreased.

• In the PLT histogram, the tail of the plot was elevated and in a serration pattern; interference from large platelets, platelet aggregation, or RBC fragments should be ruled out by microscopy. In the DIFF scattergram, the Eos particles was significantly dense and Lym particle cluster extended to the high-fluorescence region. Abnormal cells were suspected.

C Peripheral blood morphology examination

	White	e blood cells		
	White blood cells	110	100%	
L	Segmented neutrophils	7	6.4	
	Band neutrophils	7	6.4	0
	Lymphocytes	46	41.8	
L	Monocytes	1	0.9	
н	Eosinophils	18	16.4	
	Metamyelocytes	2	1.8	
1	Myelocytes	3	2.7	-
1	Blasts	24	21.8	
1	Abnormal lymphocyte	2	1.8	
	Non-white blood cells	76	%	
	Smudge cells	10	9.1	
	P	Platelets		
	PLT estimate	Estimated	Estimation	
		result	method	
	Platelet concentration	58*10^9/L	Automated	-207A
	Platelet concentration	51*10^9/L	Manual	
	Red	blood cells		
	Size	Degree	%	
	Anisocytosis	0		
	Macrocytes	0	0.8	
1	Microcytes	3+	21.3	
	Color	Degree	%	lmr
1	Hypochromic cells	3+	82.1	nro
	Polychromasia	0	0.3	pro
	Shape	Degree	%	pro
1	Poikilocytosis	3+		nuc
1	Schistocytes	3+	2.9	cyte
	Echinocytes	0	0.3	cor
	Elliptocytes	0	1.3	
1	Ovalocytes	2+	13.7	
	Stomatocytes	0	0.0	
1	Target cells	2+	5.1	
	Teardrop cells	0	0.7	



Results from manual re-classification

mmature granulocytes accounted for 4.5%; myeloblasts and promyelocytes accounted for 23.6%. Myeloblasts and promyelocytes had round or oval cell bodies, round or oval nuclei, and fine chromatins; cuts marks were visible; cytoplasm was scant and in blue color. The PLT estimate was consistent with the CBC result. Schistocytes were observed.

Other examinations

ltem	Result
Flow cytometry	Abnormal naive B cells accounted for 97.60% of nucleated cells, expressing CD19, CD22, cCD79a, CD34, CD10, cTdT, CD58, and not expressing CD33, CD15, CD13, CD38, CD81, CD20, CD7, cMPO, cCD3, CD117, CD56, clgM, or CD66c. Immunophenotyping suggested B cell acute lymphoblastic leukemia, with high probability of Common B-ALL
Bone marrow cytology	Lymphocytes accounted for 97.5%, of which myeloblasts and promyelocytes accounted for 97%. Cytochemical staining results: POX (-), AS-DCE (-), a-NAE (+), a-NAF (no inhibition), a-NBE (-), PAS (+). Suggested diagnosis: ALL
Fusion gene	Negative, no large deletion of IKZF1 gene was observed, and ph-like gene rearrangement was negative.
Chromosomal examination	Chromosome karyotyping showed 46, XY(3)

Case analysis

This patient was diagnosed with B-cell acute lymphoblastic leukemia (B-ALL).

Case 11 B-cell lymphoblastic leukemia (B-ALL)

Clinical information

The patient was a 48-year-old female who experienced with flatulent dyspnea and low-grade fever with no obvious cause more than 10 days ago. The patient was alert and oriented with no skin or mucous membrane bleeding. No enlargement of superficial lymph nodes, liver, or spleen was observed.

Sec results

Parameter	Alaı	m		Result	Unit
WBC	&		н	83.29	10^9/L
Neu#	&	R	L	0.98	10^9/L
Lym#	&	R	н	81.62	10^9/L
Mon#		R		0.66	10^9/L
Eos#		R	L	0.01	10^9/L
Bas#		R		0.02	10^9/L
IMG#		R		0.11	10^9/L
Neu%	&	R	L	1.2	%
Lym%	&	R	н	98.0	%
Mon%		R	L	0.8	%
Eos%		R	L	0.0	%
Bas%		R		0.0	%
IMG%		R		0.1	%
RBC			L	2.23	10^12/L
HGB			L	65	g/L
НСТ			L	19.5	%
MCV				87.8	fL
МСН				29.1	pg
MCHC				331	g/L
RDW-CV				13.6	%
RDW-SD				43.5	fL
PLT			L	15	10^9/L
MPV				9.0	fL
PDW				17.0	
РСТ			L	0.013	%
P-LCC			L	3	10^9/L
P-LCR				23.3	%
NRBC#				0.036	10^9/L
NRBC%				0.04	/100WBC







Alarm

- Abn. WBC scattergram
- Abn Lymph/blast?
- Immature Gran?Lymphocytosis
- Thrombocytopenia

- Neutropenia

- Leukocytosis

- Anemia

- WBC increased, predominantly Lym; moderate anemia; extremely low PLT.
- In the DIFF scattergram, Lym particles extended towards the high-fluorescence region, appearing as comet-shaped clusters; neu particles were significantly decreased.

Teripheral blood morphology examination

White blood cells			Myeloblasts and promyelocytes	
	White blood cells	201	100%	
L	Segmented neutrophils	1	0.5	
	Band neutrophils	1	0.5	
	Lymphocytes	49	24.4	
1	Blasts	150	74.6	
	Non-white blood cells	47	%	
	Nucleated RBCs	1	0.5	
	Smudge cells	43	21.4	
	Platele	ts		
	PLT estimate	Estimated	Estimation	
		result	method	A R. A A
	Platelet concentration	55*10^9/L	Automated	NU VIVACI A A
	Platelet concentration	76*10^9/L	Manual	
	Red blood cells			
	Size	Degree	%	
1	Anisocytosis	3+		
	Macrocytes	0	0.3	
1	Microcytes	3+	50.2	
	Color	Degree	%	
1	Hypochromic cells	3+	71.3	
	Polychromasia	0	0.5	
	Shape	Degree	%	WA AUVIN
1	Poikilocytosis	1+		0 0 0 0 0 V
1	Schistocytes	1+	1.9	
	Echinocytes	0	1.4	
	Elliptocytes	0	0.2	
	Ovalocytes	0	11.9	0 00 00 000
	Stomatocytes	0	0.0	
	Target cells	0	0.0	
	Teardrop cells	0	0.9	0 00 000 0000

Results from manual re-classification

Blasts accounted for 74.6%, with variable cell sizes, scant cytoplasm, verrucous protrusions, round or oval nuclei, fine chromatins, and visible nucleoli.

Other examinations

ltem	Result
Bone marrow cytology	Abnormal lymphocyte proliferation, prolymohocytes accounted for 93.5%, and mature lymphocytes accounted for 5.5%. Suggested diagnosis: ALL
Flow cytometry	Abnormal B lymphoblasts accounted for 92% of nucleated cells (Common B-ALL)
Gene	BCR-ABL (p190) positive
Chromosomal examination	46,XX,t(9;22)(q34;q11.2)

Case analysis

- The diagnosis of this case was B-ALL with t(9;22)(q34;q11.2); BCR-ABL1. This is a subtype with poor prognosis, but the prognosis of Ph+ ALL has gradually improved in recent years with the use of tyrosine kinase inhibitors.
- The DIFF scattergram of the two B-ALL cases are very similar, with Lym particles forming comet-shaped clusters extending towards the upper right, and the left edge of the clusters not showing the rightward concave observed in the AML case. Therefore, when similar scattergram are seen in practice, it is important to watch for abnormal lymphocytes.

Case 12 Chronic Lymophocytic leukemia (CLL)

Clinical information

The patient was a 64-year-old female who had been diagnosed with chronic lymphocytic leukemia more than 2 years ago. Considering the patient had no indication for treatment, CBC and blood smear examination were regularly conducted at the Department of Hematology. The results showed fluctuations in WBCs ranging from $78-160 \times 10^{9}$ /L and lymphocytes ranging from $75-154 \times 10^{9}$ /L.

Sec results

Parameter	Alarm		Result	Unit
WBC		Н	131.45	10^9/L
Neu#	R		4.18	10^9/L
Lym#	R	Н	125.25	10^9/L
Mon#	R	Н	1.94	10^9/L
Eos#	R		0.07	10^9/L
Bas#	R		0.01	10^9/L
IMG#	R		0.10	10^9/L
Neu%	R	L	3.2	%
Lym%	R	Н	95.2	%
Mon%	R	L	1.5	%
Eos%	R	L	0.1	%
Bas%	R		0.0	%
IMG%	R		0.1	%
RBC			4.89	10^12/L
HGB			131	g/L
НСТ		L	39.9	%
MCV			81.7	fL
МСН		L	26.7	pg
MCHC			327	g/L
RDW-CV		Н	18.1	%
RDW-SD			53.4	fL
PLT	R		134	10^9/L
MPV	R		9.6	fL
PDW	R		15.7	
РСТ	R		0.129	%
P-LCC	R		34	10^9/L
P-LCR	R		25.6	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC







- Abn Lymph/blast?
- Immature Gran?
- Monocytosis
-
- WBC increased, predominantly Lymphocytes, and Monocytes also increased; RBC and PLT were roughly normal.
- In the DIFF scattergram, the Lym region was significantly dense and some particles extended towards the high-fluorescence region. These particles were classified as Mon by the software. Blood smear re-check was required to confirm whether they were abnormal cells.

FL

😚 Peripheral blood morphology examination

	White bloo	d cells	
	White blood cells	300	100%
L	Segmented neutrophils	6	2.0
	Band neutrophils	1	0.3
н	Lymphocytes	287	95.7
L	Monocytes	2	0.7
1	Blasts	4	1.3
	Non-white blood cells	93	%
	Large platelets	7	
	Smudge cells	83	27.7
	Platele	ts	
	PLT estimate	Estimated	Estimation
		result	method
	Platelet concentration	158*10^9/L	Automated
	Platelet concentration	183*10^9/L	Manual
	Red blood	cells	
	Size	Degree	%
	Anisocytosis	0	
	Macrocytes	0	0.5
1	Microcytes	2+	11.6
	Color	Degree	%
1	Hypochromic cells	3+	20.1
	Polychromasia	0	0.1
	Shape	Degree	%
1	Poikilocytosis	1+	
1	Schistocytes	1+	0.9
	Echinocytes	0	2.1
	Elliptocoytes	0	4.0
	Ovalocytes	0	15.6
	Stomatocytes	0	0.0
	Target cells	0	0.0
	Teardrop cells	0	0.4



Results from manual re-classification

Mature lymphocytes accounted for 97%, with small round cell bodies and scant cytoplasm; the nuclei were round with agglutinated chromatins.

Case analysis

- Chronic lymphocytic leukemia (CLL) is a mature B lymphocyte clonal proliferative tumor. Patients have monoclonal B lymphocytes ≥ 5 × 10^9/L in peripheral blood, and leukemic cell morphology shows mature-appearing small lymphocytes, with occasional naive lymphocytes and a small number of immature or atypical lymphocytes. Neutrophil percentage is decreased, and thrombocytopenia and/or anemia may occur as the disease progresses. Smudge cells are easily seen in peripheral blood.
- The course of the disease is slow, and treatment is usually not necessary. Follow-up should be conducted every 2–6 months. However, treatment can begin if any of the following conditions occur:
 - 1. Evidence of progressive bone marrow failure: manifested as progressive reduction in hemoglobin and/or platelets, with hemoglobin less than 100 g/L and platelets less than 100×10^{9} /L.
 - 2. Massive splenomegaly (e.g., > 6 cm below left costal margin) or progressive or symptomatic splenomegaly.
 - 3. Massive lymphodenopathy (e.g., longest diameter > 10 cm) or progressive or symptomatic lymphodenopathy.
 - 4. Naive lymphocytes \ge 30 \times 10^9/L, with progressive lymphocytosis, e.g., an increase of 50% within 2 months, or lymphocyte doubling time (LDT) < 6 months. When the naive lymphocytes are < 30 \times 10^9/L, LDT cannot be used as the only treatment indication, and other conditions that can cause lymphocytosis, such as infection or steroid use, should be ruled out.
 - 5. Autoimmune hemolytic anemia and/or thrombocytopenia that do not respond well to corticosteroids or other standard of care.
 - 6. Symptomatic or functionally impairing extranodal lesions (skin, kidney, lung, spine, etc.), especially when symptoms cannot be resolved with symptomatic treatment.
 - 7. Presence of at least one of the following disease-related symptoms.
 - **1** Weight loss of \geq 10% for no apparent reason in the past 6 months.
 - ② Severe fatigue (e.g., ECOG performance status ≥ 2; unable to carry out daily activities).
 - Body temperature > 38.0°C with no evidence of infection, lasting for more than 2 weeks.
 - In the second second

Case 13 Mantle cell lymphoma (MCL)

Clinical information

The patient was a 60-year-old male who had been experiencing left knee joint pain and limited mobility for over 2 years, which worsened over the past 3 months. There was mild deformity in the left knee joint, and the patient was admitted to the hospital for "left knee osteoarthritis".

CBC results

Parameter	Alarm		Result	Unit
WBC		н	15.06	10^9/L
Neu#	R	L	1.94	10^9/L
Lym#	R	н	12.77	10^9/L
Mon#	R		0.29	10^9/L
Eos#			0.04	10^9/L
Bas#			0.02	10^9/L
IMG#	R		0.00	10^9/L
Neu%	R	L	12.9	%
Lym%	R	н	84.9	%
Mon%	R	L	1.9	%
Eos%		L	0.2	%
Bas%			0.1	%
IMG%	R		0.0	%
RBC			5.17	10^12/L
HGB			159	g/L
НСТ			46.5	%
MCV			89.9	fL
МСН			30.8	pg
MCHC			342	g/L
RDW-CV			12.8	%
RDW-SD			44.6	fL
PLT			125	10^9/L
MPV			9.4	fL
PDW			15.9	
РСТ			0.117	%
P-LCC		L	26	10^9/L
P-LCR			21.2	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC









WNB

Alarm

- Abn. WBC scattergram
- Abn Lymph/blast?
- Lymphocytosis
- WBC increased, predominantly Lym; Neu decreased; RBC and PLT were normal
- In the DIFF scattergram, Lym region was significantly dense, with a roughly normal overall distribution. Based on the WNB scattergram, abnormal cell alarm was given by nucleus-plasma double-check technology.

Peripheral blood morphology examination 0

White blood cells						
White blood cells	150	100%				
Segmented neutrophils	8	5.3				
Band neutrophils	3	2.0				
Lymphocytes	109	72.7				
Monocytes	3	2.0				
Eosinophils	1	0.7				
Blasts	10	6.7				
Reactive lymphocytes	2	1.3				
Abnormal lymphocyte	14	9.3				
Non-white blood cells	14	%				
Large platelets	1					
Smudge cells	9	6.0				
Platele	ets					
PLT estimate	Estimated	Estimation				
Platelet concentration	128*10/0/	Automated				
Platelet concentration	143*10^9/L	Manual				
Red blood	cells					
Size	Degree	%				
Anisocytosis	0					
Macrocytes	0	0.3				
Microcytes	0	8.3				
Color	Degree	%				
Hypochromic cells	0	0.7				
Polychromasia	0	0.1				
Shape	Degree	%				
Poikilocytosis	0					
Schistocytes	0	0.3				
Echinocytes	0	0.2				
Elliptocytes	0	0.0				
Ovalocytes	0	4.9				
Stomatocytes	0	0.1				
Target cells	0	0.0				
	White blood cells Segmented neutrophils Band neutrophils Lymphocytes Cosinophils Eosinophils Eosinophils Eosinophils Elasts Reactive lymphocytes Abnormal lymphocytes Mon-white blood cells Platelet Smudge cells Platelet concentration Platelet concentration	White blood cells50White blood cells3Segmented neutrophils3Band neutrophils109Monocytes1Cosinophils1Blasts10Reactive lymphocytes2Abnormal lymphocyte14Iarge platelets1Smudge cells2Platelet concentration128*10040Platelet concentration128*10040Platelet concentration128*10040Platelet concentration128*10040Platelet concentration0Macrocytes0Macrocytes0Microcytes0Platelet concentration10State0Microcytes0Microcytes0State0State0Microcytes0State0State0Platelet0Microcytes0Color0State0Stat				



Results from manual re-classification

Lymphocytes accounted for 82% and abnormal lymphocytes accounted for 8%, with blue and clear cytoplasm, fine chromatins, and visible nucleoli.

Other examinations

Item	Result
Bone marrow cytology	The lymphocyte percentage increased, with some irregular morphology, suggesting lymphoproliferative disorders (LPD).
Immunohistochemistry	Multiple clusters of CD3 (+), multiple foci of CD20 (+), multiple foci of CD79A (+), multiple foci of CD5 (+), CD23 (-), CD10 lymphocytes (-), few cyclinD1 (+), ki-67 (15%+) Consistent with those of small B cell lymphoma (tumor cells accounted for approximately 40%)
Flow cytometry	CD5 + CD10 + B lymphocytes accounted for 62.48%, MCL?
Chromosomal examination	46,XY,t(11;14)(q23;q32), add(14)(p11.2)[14]/46,XY[6]

Case analysis

- Mantle cell lymphoma (MCL) is a small-to-medium sized, monomorphic, mature B-cell tumor with specific immunophenotype and reproducible genetic abnormalities. CD5 and SOX11 are typically expressed, and over 95% of patients have CCND1 gene rearrangement (t(11:14)(q13;q32) abnormality), resulting in high nuclear expression of Cyclin D1 protein. MCL mainly affects elderly men and often involves extranodal sites. It has both the characteristics of rapid progression of aggressive lymphoma and incurability of indolent lymphoma. The survival time with multi-drug combination chemotherapy is about 3–5 years, and it currently belongs to incurable NHL.
- The clinical manifestations of MCL are nonspecific, so the diagnosis and prognosis evaluation at the initial visit are critical. Pathological diagnosis is the only means to confirm the diagnosis of MCL.

Case 14 May-Hegglin anomaly

Clinical information

The patient was a 31-year-old female who was admitted in November due to "38 + 5 weeks of pregnancy with ultrasound indicating oligohydramnios".

CBC results

Parameter	Alarm	BC-6800Plus_2 8:05 2022/10/26	BC-6800Plus_1 8:09 2022/10/26	Unit
WBC		9.60	9.16	10^9/L
Neu#	Н	7.80		10^9/L
Lym#		1.24		10^9/L
Mon#		0.47		10^9/L
Eos#		0.07		10^9/L
Bas#		0.02		10^9/L
IMG#		0.09		10^9/L
Neu%	Н	81.2		%
Lym%	L	13.0		%
Mon%		4.9		%
Eos%		0.7		%
Bas%		0.2		%
IMG%		0.9		%
RBC		3.87	3.96	10^12/L
HGB		123	122	g/L
НСТ	L	35.5	36.5	%
MCV		91.9	92.1	fL
МСН		31.8	30.9	pg
MCHC		346	335	g/L
RDW-CV		12.9	13.0	%
RDW-SD		45.9	44.3	fL
PLT	R L	31	60	10^9/L
MPV	RΗ	15.5	15.9	fL
PDW	R	16.6	17.1	
РСТ	R L	0.047	0.056	%
P-LCC	R L	20	38	10^9/L
P-LCR	RΗ	64.0	62.8	%
NRBC#		0.000		10^9/L
NRBC%		0.00		/100WBC



• WBC and RBC were roughly normal; low PLT.

• Low PLT triggered a re-test rule. Upon re-test, PLT-O was found to be 60 × 10^9/L. It was speculated that PLT-I was falsely low due to interference from large platelets or platelet aggregation.

C Peripheral blood morphology examination

White bloc	od cells	
White blood cells	150	100%
Segmented neutrophils Band neutrophils Lymphocytes Monocytes Eosinophils Metamyelocytes Reactive lymphocytes Non-white blood cells Giant platelets	133 3 6 4 1 1 2 97 24	88.6 2.0 4.0 2.7 0.7 0.7 1.3 %
Large platelets	65 7	47
Smuage cells	/ ets	4./
PLT estimate	Estimated result	Estimation method
Platelet concentration	71*10^9/L	Manual
Red blood Size Anisocytosis Macrocytes Microcytes	d cells Degree 0 0 0	% 1.2 5.0
Color	Degree	%
Polychromasia	2+ 0	0.2
Shape	Degree	%
Poikilocytosis	0	
Schistocytes	0	0.0
Echinocytes	0	0.0
Elliptocytes	0	0.1
Ovalocytes	0	2.4
Stomatocytes	0	0.9
larget cells	0	0.0

Results from manual re-classification

The classification results were consistent with the CBC results. Blue, cloudy plaques resembling Dohle bodies, irregular in shape and large in size, were observed in the cytoplasm of neutrophils as well as eosinophils at all stages.

May-Hegglin anomaly was suspected. Additionally, giant and large platelets were observed in 59/100 WBCs, and the PLT estimate was consistent with the PLT-O result.

Case analysis

L

1

- The patient presented with thrombocytopenia, giant platelets, and neutrophil inclusions, which were consistent with the manifestations of May-Hegglin anomaly. Subsequent genetic testing confirmed the presence of a heterozygous MYH9 mutation located at 22q12.3, AD.
- May-Hegglin anomaly is a type of MYH9-related disorder, which is a rare autosomal dominant genetic disorder. Despite a decrease in platelet count, the total platelet volume in the blood is not significantly reduced due to an increase in platelet size. Platelet function remains mostly normal, and bleeding tendencies are mild. Patients are usually diagnosed in adulthood.
- Due to significant thrombocytopenia, most patients are misdiagnosed with ITP at initial diagnosis, and underwent unnecessary hormone therapy or splenectomy.

Case 15 Pelger–Huët anomaly

Clinical information

The patient was a 76-year-old female with poorly controlled type diabetes who visited the Department of Endocrinology of the hospital.

Sec results

Parameter	Alarm		Result	Unit
WBC		L	1.72	10^9/L
Neu#		L	1.28	10^9/L
Lym#		L	0.37	10^9/L
Mon#		L	0.06	10^9/L
Eos#		L	0.01	10^9/L
Bas#			0.00	10^9/L
IMG#			0.00	10^9/L
Neu%		н	74.6	%
Lym%			21.4	%
Mon%			3.6	%
Eos%		L	0.3	%
Bas%			0.1	%
IMG%			0.1	%
RBC		L	2.70	10^12/L
HGB		L	83	g/L
НСТ		L	24.4	%
MCV			90.3	fL
МСН			30.7	pg
MCHC			340	g/L
RDW-CV			13.4	%
RDW-SD			46.0	fL
PLT		L	47	10^9/L
MPV			10.1	fL
PDW			16.3	
РСТ		L	0.047	%
P-LCC		L	12	10^9/L
P-LCR			26.4	%
NRBC#			0.000	10^9/L
NRBC%			0.00	/100WBC









Alarm

- Pancytopenia
- Lymphopenia
- Leukocytopenia
- Anemia
- Thrombocytopenia

• Pancytopenia without abnormal cell alarm.

• No abnormality in histograms or scattergrams.

Peripheral blood morphology examination

White blood cells								
	White blood cells	200	100%					
L	Segmented neutrophils	66	33.0					
!	Band neutrophils	84	42.0					
	Lymphocytes	40	20.0					
L	Monocytes	4	2.0					
	Eosinophils	4	2.0					
	Metamyelocytes	2	1.0					
	Non-white blood cells	27	%					
	Giant platelets	1						
	Large platelets	1						
	Smudge cells	22	11.0					

Results from manual re-classification

The neutrophil nuclei appeared mostly rod-shaped or bi-lobed, with rod-shaped nuclei resembling peanuts or eyeglasses, and lobed nuclei connected by thin threads; the lobes were mostly round or elliptical; the chromatins were dense, deeply stained, and aggregated into small blocks or rod-like structures; hyperlobulated nuclei were not observed.



Case analysis

- Based on the special morphology of neutrophils, the possibility of hereditary Pelger-Huët anomaly was first considered for this patient. Thus, clinical consultation was made, and the patient was advised to undergo a pedigree analysis of immediate family.
- The patient has a son and a daughter who are both healthy. CBC samples were collected from the patient's son and daughter the next day for testing, and the results were normal. The blood cell morphology results are shown below:

White blood cells				Patient's son
	White blood cells	200	100%	Sammented nautronbils % 487%
L	Segmented neutrophils	96	48.0	
!	Band neutrophils	37	18.5	
	Lymphocytes	59	29.5	N C & W W N 3 N C Z 3 N W Z 1
	Monocytes	8	4.0	
	Non-white blood cells	48	%	
	Giant platelets	2		
	Large platelets	б		
	Platelet clumps	1		
	Smudge cells	31	15.5	

White blood cells								
	White blood cells	200	100%					
L	Segmented neutrophils	77	38.5					
1	Band neutrophils	72	36.0					
	Lymphocytes	41	20.5					
	Monocytes	7	3.5					
	Metamyelocytes	1	0.5					
	Myelocytes	1	0.5					
	Non-white blood cells	39	%					
	Giant platelets	1						
	Large platelets	1						
	Smudge cells	20	10.0					

Patient's daughter



- The above results showed that the peripheral blood neutrophil morphologies of the patient's son and daughter were generally the same as that of the patient, and the neutrophil nuclei were mostly rod-shaped or bi-lobed. The MC-80 microscope was used to analyze 200 WBCs, and no cells with 3 or more lobes were observed, so the cell morphology was considered to be congenital Pelger-Huët anomaly.
- Pelger-Huët anomaly is an autosomal dominant genetic abnormality characterized by reduced lobulation of mature neutrophils. It is also known as familial neutrophil anomaly or Pelger-Huët neutrophil anomaly.
- Morphological features: Reduced lobulation of neutrophils. The nuclei were non-lobulated in an oval shape (e.g., kidney-shaped, rod-shaped) or divided into two lobes resembling peanuts, eyeglasses, or dumbbells (similar to panda eyes). The chromatins were concentrated, deeply stained, and aggregated into small blocks or thread-like structures, forming blank gaps in between.

Case 16 Green neutrophilic inclusions

E Clinical information

The patient was a 52-year-old female who experienced pyrexia, diarrhea, headache, fatigue, and nausea one week ago. The highest body temperature was 39.5 °C. She was later admitted to the intensive care unit due to confusion, transient unilateral limb convulsions, and hematemesis. Subsequently, she was diagnosed with hemophagocytic syndrome secondary to severe fever with thrombocytopenia syndrome virus (SFTSV) infection.

🔊 Peripheral blood morphology examination





Results from manual re-classification

Neutrophils exhibited vacuolar degeneration and demonstrated bacterial phagocytosis, along with the presence of blue-green round inclusions in both neutrophils and eosinophils.

Case analysis

- After the discovery of blue-green inclusions, high-sensitivity cardiac troponin I (hs-cTnl), alanine aminotransferase, WBCs, and other test indicators continued to increase or remained at high levels, and the NRBC percentage was as high as 20%. Six days later, the myocardial and pancreatic damage caused by the inflammatory cytokine storm did not show significant improvement, and the patient eventually died.
- Blue-green inclusions are bright green inclusions that appear in the cytoplasm of neutrophils or monocytes, and are usually associated with acute liver failure, lactic acidosis, and other diseases. Their appearance indicates that the patient is in critical conditions, which can alert clinicians to closely monitor the patient's emergency situation, actively treat the underlying disease, protect the liver, and promptly correct lactic acidosis. This may change the patient's outcome.

Long-standing Problems Associated With PLT Aggregation

In vitro platelet aggregation is commonly caused by EDTA-dependent pseudo thrombocytopenia (EDTA-PTCP) in clinical practice. It has been reported that the incidence of EDTA-PTCP is $0.07\% \sim 0.20\%$, while the incidence among platelet donors is 0.2% and that among hospitalized patients is $0.1\% \sim 2.0\%^{[1]}$

Challenges for Clinical Departments in Handling Samples With Low PLT Counts



Complex low PLT count screening may lead to false low count reports **Clinical Disputes**



Complicated time-consuming microscopic examinations add to the workload **Uncontrolled TAT**



Repeated blood collections trigger patient complaints Unsatisfactory Patient Experience

Difficulties Encountered by Traditional Instruments in Handling EDTA-PTCP Samples



Address PLT Aggregation-related Challenges Easily With PLT De-aggregation Technology



New Technology for Interference Elimination

Mindray is committed to solving the genuine concerns of customers and pursuing excellence. The groundbreaking PLT de-aggregation technology consolidates 6 years of collaboration with reputable institutions and guidance from experts, successfully addressing the PLT aggregation-related challenges in clinical laboratory settings.



Process Comparison: De-aggregation Process is 4+ times More Efficient Than

Traditional Process Note: The testing time is estimated from the daily work of grade A tertiary hospitals

Traditional	2 mins 5 mins 00	5 mins 5 mins 20 mins 10 mins 5 mins	5 mins 0
Process	Blood collection Testing	Check Optical re-testing Prepare smears, Using a different Re-testing perform microscopic anticoagulant Additional examination blood collections	Re-check
De-aggregation	2 mins 5 mins	1 mins 	5 mins
Process	Blood collection lesting	Automated optical re-testing (including a waiting time of 13 mins) <	validation

De-aggregation Technology Effectively Addresses Spurious Low PLT Count Due to EDTA-PTCP



Advanced Hematology Analyzer & De-aggregation Technology

The instrument and technology not only can handle the samples affected by EDTA-PTCP, but also automates the entire de-aggregation process without the need for manual intervention. This uniformly controls the TAT and enhances operational efficiency and convenience in laboratories.

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